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Attorney Docket No. 1134R

September 8, 2000

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Box Patent Application  
Washington, DC 20231

**UTILITY PATENT APPLICATION TRANSMITTAL**

Inventor(s): Jonathan P. Duvick, Jacob T. Gilliam, Joyce R. Maddox, Aragula Gururaj Rao, Oswald R. Crasta, Otto Folkerts

Title: Amino Polyol Amine Oxidase Polynucleotides and Related Polypeptides and Methods of Use

**APPLICATION ELEMENTS**

1.  Fee Transmittal Form (Submit an original, and a duplicate for fee processing)
2.  Specification [Total Pages 83]  
(Preferred arrangement set forth below)
  - Descriptive title of the Invention
  - Cross Reference to Related Applications
  - Background of the Invention
  - Brief Summary of the Invention
  - Brief Description of the Drawings (if filed)
  - Detailed Description
  - Claim(s)
  - Abstract of the Disclosure
3.  Drawing(s) (35 USC 113) [Total Sheets 2]
  - a.  Formal
  - b.  Informal

4.  Oath or Declaration [Total Pages \_\_\_\_\_]  
a.  Newly executed (original or copy)  
b.  Copy from a prior application (37 CFR 1.63(d))  
(for continuation/divisional with Box 16 completed)  
i.  DELETION OF INVENTOR(S)  
Signed statement attached deleting inventor(s) named in the prior  
application, see 37 CFR 1.63(d)(2) and 1.33(b).
5.  Microfiche Computer Program (Appendix)
6.  Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)  
a.  Computer Readable Copy  
b.  Paper Copy (identical to computer copy) [Total Pages 87]  
c.  Statement verifying identity to above copies

**ACCOMPANYING APPLICATION PARTS**

7.  Assignment Papers (cover sheet & document(s))
8.  37 CFR 3.73(b) Statement  Power of Attorney  
(where there is an assignee)
9.  English Translation Document (if applicable)
10.  Information Disclosure Statement (IDS/PTO-1449)  Copies of IDS Citations
11.  Preliminary Amendment
12.  Return Receipt Postcard (MPEP 503) (Should be specifically itemized)
13.  Small Entity Statement(s)  Statement filed in prior application  
Status still proper and desired
14.  Certified Copy of Priority document(s)
15.  Other:

Attorney Docket No.: 1134R

16. If a **CONTINUING APPLICATION**, check the appropriate box, and supply the requisite information below and in a preliminary amendment:

Continuation     Divisional     Continuation-in-part (CIP) of prior application  
No. 09/352,159 and 09/352,168.

Prior application information: Examiner \_\_\_\_\_ Group/Art Unit: \_\_\_\_\_

For **CONTINUATION or DIVISIONAL APPS** only: The entire disclosure of the prior application, from which an oath or declaration is supplied under Box 4b, is considered a part of the disclosure of the accompanying continuation or divisional application and is hereby incorporated by reference. The incorporation can only be relied upon when a portion has been inadvertently omitted from the submitted application parts.

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Respectfully submitted,



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AMINO POLYOL AMINE OXIDASE POLYNUCLEOTIDES AND RELATED  
POLYPEPTIDES AND METHODS OF USE

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**Technical Field**

The present invention relates generally to the detection and isolation of fumonisin and AP1 degrading enzymes and to compositions and methods for degradation of 10 fumonisin, a structurally related mycotoxin, or its hydrolysis product AP1. This method has broad application in agricultural biotechnology and crop agriculture and in the improvement of food grain quality.

**Cross Reference To Related Application**

15        This application is a continuation-in-part of U.S. Application No. 09/352,159, which claims the benefit of U.S. Provisional Application No. 60/135,391, filed May 21, 1999 and U.S. Provisional Application No. 60/092,936, filed July, 15 1998 all of which are hereby incorporated by reference. This application also claims the benefit of U.S. 20 Application No. 09/352,168, which is hereby incorporated by reference.

**Background of the Invention**

Fungal diseases are common problems in crop agriculture. Many strides have been made against plant diseases as exemplified by the use of hybrid plants, pesticides and 25 improved agricultural practices. However, as any grower or home gardener can attest, the problems of fungal plant disease continue to cause difficulties in plant cultivation. Thus, there is a continuing need for new methods and materials for solving the problems caused by fungal diseases of plants.

These problems can be met through a variety of approaches. For example, the 30 infectious organisms can be controlled through the use of agents that are selectively biocidal for the pathogens. Another method is interference with the mechanism by which the pathogen invades the host crop plant. Yet another method, in the case of pathogens that cause crop losses, is interference with the mechanism by which the pathogen causes

injury to the host crop plant. Still another method, in the case of pathogens that produce toxins that are undesirable to mammals or other animals that feed on the crop plants, is interference with toxin production, storage, or activity. This invention falls into the latter two categories.

Since their discovery and structural elucidation in 1988 (Bezuidenhout *et al.*, Journal *Chem Soc, Chem Commun* 1988: 743-745 (1988)), fumonisins have been recognized as a potentially serious problem in maize-fed livestock. They are linked to several animal toxicoses including leukoencephalomalacia (Marasas, *et al.*, *Onderstepoort Journal of Veterinary Research* 55: 197-204 (1988); Wilson, *et al.*, *American Association of Veterinary Laboratory Diagnosticians: Abstracts 33rd Annual Meeting*, Denver, Colorado, October 7-9, 1990, Madison, Wisconsin, USA) and porcine pulmonary edema (Colvin, *et al.*, *Mycopathologia* 117: 79-82 (1992)). Fumonisins are also suspected carcinogens (Geary W (1971) *Coord Chem Rev* 7: 81; Gelderblom, *et al.*, *Carcinogenesis* 12: 1247-1251 (1991); Gelderblom, *et al.*, *Carcinogenesis* 13: 433-437 (1992)). *Fusarium* isolates in section *Liseola* produce fumonisins in culture at levels from 2 to >4000 ppm (Leslie, *et al.*, *Phytopathology* 82: 341-345 (1992)). Isolates from maize (predominantly mating population A) are among the highest producers of fumonisin. (Leslie *et al.*, *supra*). Fumonisin levels detected in field-grown maize have fluctuated widely depending on location and growing season, but both preharvest and postharvest surveys of field maize have indicated that the potential for high levels of fumonisins exists (Murphy, *et al.*, *J Agr Food Chem* 41: 263-266 (1993)). Surveys of food and feed products have also detected fumonisin (Holcomb, *et al.*, *J Agr Food Chem* 41: 764-767 (1993); Hopmans, *et al.*, *J Agr Food Chem* 41: 1655-1658 (1993); Sydenham, *et al.*, *J Agr Food Chem* 39: 2014-2018 (1991)). The etiology of *Fusarium* ear mold is poorly understood, although physical damage to the ear and certain environmental conditions can contribute to its occurrence (Nelson, *Mycopathologia* 117: 29-36 (1992)). *Fusarium* can be isolated from most field grown maize, even when no visible mold is present. The relationship between seedling infection and stalk and ear diseases caused by *Fusarium* is not clear. Genetic resistance to visible kernel mold has been identified (Gendloff, *et al.*, *Phytopathology* 76: 684-688 (1986); Holley, *et al.*, *Plant Dis* 73: 578-580 (1989)), but the relationship to visible mold to fumonisin production has yet to be elucidated.

Fumonisins have been shown in *in vitro* mammalian cell studies to inhibit sphingolipid biosynthesis through inhibition of the enzyme sphingosine N-acetyl transferase, resulting in the accumulation of the precursor sphinganine (Norred, *et al.*, *Mycopathologia* 117: 73-78 (1992); Wang, *et al.*, *Biol Chem* 266: 14486 (1991); Yoo, *et al.*, *Toxicol Appl Pharmacol* 114: 9-15 (1992); Nelson, *et al.*, *Annu Rev Phytopathol* 31:233-252 (1993)). It is likely that inhibition of this pathway accounts for at least some of fumonisin's toxicity, and support for this comes from measures of sphinganine: sphingosine ratios in animals fed purified fumonisin (Wang, *et al.*, *J Nutr* 122: 1706-1716 (1992)). Fumonisins also affect plant cell growth (Abbas, *et al.*, *Weed Technol* 6: 548-552 (1992); Vanasch, *et al.*, *Phytopathology* 82: 1330-1332 (1992); Vesonder, *et al.*, *Arch Environ Contam Toxicol* 23: 464-467 (1992)). Kuti *et al.*, (Abstract, Annual Meeting American Phytopathological Society, Memphis, TN: APS Press 1993) reported on the ability of exogenously added fumonisins to accelerate disease development and increase sporulation of *Fusarium moniliforme* and *Fusarium oxysporum* on tomato.

Enzymes that degrade the fungal toxin fumonisin to its de-esterified form (e.g. AP1 from FB1) have been identified in US patent no. 5,716,820, issued February 10, 1998, US patent no. 5,792,931, issued August 11, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997, and all hereby incorporated by reference. It is understood that AP1 as used here designates the hydrolyzed form of any fumonisin, FB1, FB2, FB3, FB4, or any other AP1-like compounds, including synthetically produced AP1 like compounds, that contain a C-2 or C-1 amine group and one or more adjacent hydroxyl groups. Plants expressing a fumonisin esterase enzyme, infected by fumonisin producing fungus, and tested for fumonisin and AP1 were found to have low levels of fumonisin but high levels of AP1. AP1 is less toxic than fumonisin to plants and probably also to animals but contamination with AP1 is still a concern (Lamprecht, *et al.*, *Phytopathology*, 84:383-391 (1991)). The preferred result would be complete detoxification of fumonisin to a non-toxic form. Therefore enzymes capable of degrading AP1 are necessary for the further detoxification of fumonisin.

The present invention provides newly discovered polynucleotides and related polypeptides of amino polyol amine oxidase (abbreviated APAO, formerly known as AP1 catabolase, US patent no. 5,716,820, *supra*, US patent no. 5,792,931, *supra*; US patent no.

6,025,188, *supra*, pending US application no. 08/888,950, *supra*; trAPAO is the abbreviation for a truncated, but still functional APAO), capable of oxidatively deaminating the AP1 to a compound identified as the 2-oxo derivative of AP1 or its cyclic ketal form (abbreviated as 2-OP, formerly called AP1-N1, US patent no. 5,716,820, US 5 patent no. 5,792,931, US patent no. 6,025,188, *supra*; pending US application no. 08/888,950, *supra*), isolated from *Exophiala spinifera*, ATCC 74269. The partially purified APAO enzyme from *Exophiala spinifera* has little or no activity on intact FB1, a form of fumonisins. However, recombinant APAO enzyme from *Exophiala spinifera*, expressed in *E. coli*, has significant but reduced activity on intact FB1 and other B-series 10 fumonisins. APAO or trAPAO thus could potentially be used without fumonisin esterase since the amine group is the major target for detoxification. Alternatively, fumonisin esterase and APAO (or trAPAO) can be used together for degrading toxins.

APAO is a type of flavin amine oxidase (EC 1.4.3.4, enzyme class nomenclature, see *Enzyme Nomenclature 1992*, Recommendations of the Nomenclature Committee of the 15 IUBMB on the Nomenclature and Classification of Enzymes, Academic Press, Inc. (1992)). One class of flavin amine oxidases in mammals is known as monoamine oxidases, where they participate in the conversion of amines involved in neuronal function. A prokaryotic flavin amine oxidase that deaminates putrescine has been described (Ishizuka *et al.*, *J. Gen Microbiol.* 139:425-432 (1993)). A single fungal gene, from 20 *Aspergillus niger* has been cloned (Schilling *et al.*, *Mol Gen Genet.* 247:430-438 (1995)). It deaminates a variety of alkyl and aryl amines, but when tested for its ability to oxidize AP1, was found to not contain AP1 oxidizing activity.

The toxicity of fumonisins and their potential widespread occurrence in food and feed makes it imperative to find detoxification or elimination strategies to remove the 25 compound from the food chain.

### **Summary of the Invention**

The present invention provides polynucleotides and related polypeptides of newly discovered APAOs. SEQ ID NO: 5 contains the nucleotide sequence of an active, 30 truncated APAO (trAPAO), SEQ ID NO: 10 contains the nucleotide sequence of trAPAO with an additional lysine and SEQ ID NO: 22, 35, 37, 39, 41, 43, and 45 comprise full length nucleotide sequences of APAOs isolated from different organisms. In addition,

APAO can be modified to eliminate glycosylation sites and/or cysteine residues, for example, see SEQ ID NOS: 32, 48, 50, and 52. Another aspect of the present invention is the method of predicting possible mutagenesis sites on APAO by developing a 3-dimensional model of APAO and then identifying the possible sites that may contribute to misfolding of the protein. The present invention also includes the 3-dimensional model of APAO generated by a computer modeling program, preferably the *Modeler* program. For expression in a plant, the polynucleotide of the present invention can be operably linked to a targeting sequence. It is an object of the present invention to provide transgenic plants comprising the nucleic acids of the present invention.

Therefore, in one aspect, the present invention relates to an isolated APAO encoding polynucleotide ligated to a fumonisin esterase encoding polynucleotide wherein the APAO encoding polynucleotide comprises a member selected from (a) a polynucleotide encoding a polypeptide of the present invention; (b) a polynucleotide having at least 70% sequence identity to the polynucleotides of the present invention; and (c) a polynucleotide of the present invention. The isolated nucleic acid can be DNA. The isolated nucleic acid can also be RNA. Examples of fumonisin esterase genes include, but are not limited to ESP1 and BEST1.

In another aspect, the present invention relates to vectors comprising the polynucleotides of the present invention, including ligated and non-ligated polynucleotides. Also the present invention relates to recombinant expression cassettes, comprising a polynucleotide of the present invention operably linked to a promoter.

In another aspect, the present invention is directed to a host cell into which has been introduced the polynucleotides of the present invention, including a host cell comprising a fumonisin esterase ligated to an APAO or a fumonisin esterase not ligated to an APAO.

In yet another aspect, the present invention relates to a transgenic plant or plant cell comprising a recombinant expression cassette with a promoter operably linked to any of the isolated polynucleotides of the present invention. Preferred plants containing the recombinant expression cassette of the present invention include but are not limited to maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley, tomato, and millet. The present invention also provides transgenic seed from the transgenic plant.

In another aspect, the present invention relates to an isolated protein selected from the group consisting of (a) a polypeptide comprising at least 70% sequence identity to a polypeptide of the present invention; (b) a polypeptide encoded by a nucleic acid of the present invention; and (c) a polypeptide characterized by a polypeptide of the present invention.

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This invention further provides methods of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a structurally related mycotoxin breakdown product, by applying APAO as a spray or wash. Additionally, fumonisins and related mycotoxins can be degraded by the application of both fumonisin esterase enzymes and APAO enzymes. Mycotoxins can be degraded in harvested grain, during the processing of harvested grain, in animal feed, or in plant tissue as, for example, during the use of the plant for silage or as a spray on grain, fruit or vegetables. Further, this invention provides methods of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a structurally related mycotoxin breakdown product, by transforming the APAO polynucleotide, alone or in combination with polynucleotides encoding a fumonisin esterase, into plant cells.

The polynucleotides of the present invention can also be used as a selectable marker for plant transformation. By transforming plant cells with an expression cassette comprising a polynucleotide of the present invention and then placing the plant cells on media containing FB1, AP1 or a phytotoxic analog, only the plant cells expressing the polynucleotide of the present invention would survive.

Another embodiment of the present invention is the use of the enzyme fumonisin esterase and APAO by themselves or in combination as reagents for detecting fumonisin and structurally related toxins.

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### **Brief Description of the Drawings**

Figure 1 shows a 3-dimensional model of APAO (1B) based on the crystal structure of a related amine oxidase from maize, maize polyamine oxidase (MPAO) (1A). The sites for possible mutation of APAO to alter glycosylation sites or cysteine residues are shown.

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Figure 2 shows a 3-dimensional model of APAO (2B) based on the crystal structure of a related amine oxidase from maize MPAO (2B). The substrate binding holes are shown as circles.

## Detailed Description of the Invention

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Unless mentioned otherwise, the techniques employed or contemplated herein are standard methodologies well known to one of ordinary skill in the art. The materials, methods and examples are illustrative only and not limiting. The following is presented by way of illustration and is not intended to limit the scope of the invention.

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of botany, microbiology, tissue culture, molecular biology, chemistry, biochemistry and recombinant DNA technology, which are within the skill of the art. Such techniques are explained fully in the literature. See, e.g., J. H. Langenheim and K. V. Thimann, *Botany: Plant Biology and Its Relation to Human Affairs* (1982) John Wiley; *Cell Culture and Somatic Cell Genetics of Plants*, Vol. 1 (I. K. Vasil, ed. 1984); R. V. Stanier, J. L. Ingraham, M. L. Wheelis, and P. R. Painter, *The Microbial World*, (1986) 5th Ed., Prentice-Hall; O. D. Dhringra and J. B. Sinclair, *Basic Plant Pathology Methods*, (1985) CRC Press; Maniatis, Fritsch & Sambrook, *Molecular Cloning: A Laboratory Manual* (1982); *DNA Cloning*, Vols. I and II (D. N. Glover ed. 1985); *Oligonucleotide Synthesis* (M. J. Gait ed. 1984); *Nucleic Acid Hybridization* (B. D. Hames & S. J. Higgins eds. 1984); and the series *Methods in Enzymology* (S. Colowick and N. Kaplan, eds., Academic Press, Inc.).

Units, prefixes, and symbols may be denoted in their SI accepted form. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. Numeric ranges are inclusive of the numbers defining the range. Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes. The terms defined below are more fully defined by reference to the specification as a whole.

In describing the present invention, the following terms will be employed, and are intended to be defined as indicated below.

By "microbe" is meant any microorganism (including both eukaryotic and prokaryotic microorganisms), such as fungi, yeast, bacteria, actinomycetes, algae and protozoa, as well as other unicellular structures.

A "fumonisin-producing microbe" is any microbe capable of producing the mycotoxin fumonisin or analogs thereof. Such microbes are generally members of the fungal genus *Fusarium*, as well as recombinantly derived organisms, which have been genetically altered to enable them to produce fumonisin or analogs thereof.

By "degrading fumonisin" is meant any modification to fumonisin, AP1, or any derivative of fumonisin or AP1 which causes a decrease or loss in its toxic activity, such as degradation to less than 1%, 5%, 10%, or 50% of original toxicity, with less than 10% being preferred. Such a change can comprise cleavage of any of the various bonds, oxidation, reduction, the addition or deletion of a chemical moiety, or any other change that affects the activity of the molecule. In a preferred embodiment, the modification includes hydrolysis of the ester linkage in the molecule as a first step and then oxidative deamination. Furthermore, chemically altered fumonisin can be isolated from cultures of microbes that produce an enzyme of this invention, such as growing the organisms on media containing radioactively-labeled fumonisin, tracing the label, and isolating the degraded toxin for further study. The degraded fumonisin can be compared to the active compound for its phytotoxicity or mammalian toxicity in known sensitive species, such as porcines, rabbits, and equines or in cell or tissue culture assays. Such toxicity assays are known in the art. For example, in plants a whole leaf bioassay can be used in which solutions of the active and inactive compound are applied to the leaves of sensitive plants. The leaves may be treated *in situ* or, alternatively, excised leaves may be used. The relative toxicity of the compounds can be estimated by grading the ensuing damage to the plant tissues and by measuring the size of lesions formed within a given time period. Other known assays can be performed at the cellular level, employing standard tissue culture methodologies e.g., using cell suspension cultures.

By "fumonisin esterase" is meant any enzyme capable of hydrolysis of the ester linkage in fumonisin or a structurally similar molecule such as AAL toxin. Two examples of such enzymes are ESP1 and BEST1 found in US patent no. 5,716,820, issued February 10, 1998; US patent no. 5,792,931, issued August 11, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997.

By "structurally related mycotoxin" is meant any mycotoxin having a chemical structure related to a fumonisin or AP1 such as AAL toxin, fumonisin B1, fumonisin B2, fumonisin B3, fumonisin B4, fumonisin C1, fumonisin A1 and A2, and their analogs or hydrolyzed forms, as well as other mycotoxins having similar chemical structures, 5 including synthetically made analogs that contain a C-2 or C-1 amine group and one or more adjacent hydroxyl groups, that would be expected to be degraded by the activity of an enzyme of the present invention. The present invention is the first flavin amine oxidase known to attack a primary amine not located at C-1 (i.e. C-2 of AP1) and resulting in a keto rather than an aldehydic product.

10 It is understood that "AP1" or "amino polyol" as used here is to designate the hydrolyzed form of any fumonisin, FB1, FB2, FB3, FB4, AAL, or any other AP1-like compound, including a compound made synthetically, that contains a C-2 or C-1 amine group and one or more adjacent hydroxyl groups.

15 By "amplified" is meant the construction of multiple copies of a nucleic acid sequence or multiple copies complementary to the nucleic acid sequence using at least one of the nucleic acid sequences as a template. Amplification systems include the polymerase chain reaction (PCR) system, ligase chain reaction (LCR) system, nucleic acid sequence based amplification (NASBA, Cangene, Mississauga, Ontario), Q-Beta Replicase systems, transcription-based amplification system (TAS), and strand displacement amplification 20 (SDA). See, e.g., *Diagnostic Molecular Microbiology: Principles and Applications*, D. H. Persing *et al.*, Ed., American Society for Microbiology, Washington, DC (1993). The product of amplification is termed an amplicon.

25 The term "conservatively modified variants" applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refer to those nucleic acids that encode identical or conservatively modified variants of the amino acid sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein. For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any 30 of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are "silent variations" and represent one species of conservatively modified variation. Every nucleic acid sequence herein that encodes a polypeptide also

describes every possible silent variation of the nucleic acid. One of ordinary skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine, one exception is *Micrococcus rubens*, for which GTG is the methionine codon (Ishizuka, et al., *J. Gen'l Microbiol.*, 139:425-432 (1993)) can be  
5 modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid, which encodes a polypeptide of the present invention, is implicit in each described polypeptide sequence and incorporated herein by reference.

As to amino acid sequences, one of skill will recognize that individual substitutions, deletions or additions to a nucleic acid, peptide, polypeptide, or protein  
10 sequence which alters, adds or deletes a single amino acid or a small percentage of amino acids in the encoded sequence is a "conservatively modified variant" when the alteration results in the substitution of an amino acid with a chemically similar amino acid. Thus, any number of amino acid residues selected from the group of integers consisting of from 1 to 15 can be so altered. Thus, for example, 1, 2, 3, 4, 5, 7, or 10 alterations can be made.  
15 Conservatively modified variants typically provide similar biological activity as the unmodified polypeptide sequence from which they are derived. For example, substrate specificity, enzyme activity, or ligand/receptor binding is generally at least 30%, 40%, 50%, 60%, 70%, 80%, or 90%, preferably 60-90% of the native protein for it's native substrate. Conservative substitution tables providing functionally similar amino acids are  
20 well known in the art.

The following six groups each contain amino acids that are conservative substitutions for one another:

- 1) Alanine (A), Serine (S), Threonine (T);
- 2) Aspartic acid (D), Glutamic acid (E);
- 25 3) Asparagine (N), Glutamine (Q);
- 4) Arginine (R), Lysine (K);
- 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and
- 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

See also, Creighton (1984) Proteins W.H. Freeman and Company.

30 As used herein, "consisting essentially of" means the inclusion of additional sequences to an object polynucleotide where the additional sequences do not selectively hybridize, under stringent hybridization conditions, to the same cDNA as the

polynucleotide and where the hybridization conditions include a wash step in 0.1X SSC and 0.1% sodium dodecyl sulfate at 65°C.

By "encoding" or "encoded", with respect to a specified nucleic acid, is meant comprising the information for translation into the specified protein. A nucleic acid encoding a protein may comprise non-translated sequences (e.g., introns) within translated regions of the nucleic acid, or may lack such intervening non-translated sequences (e.g., as in cDNA). The information by which a protein is encoded is specified by the use of codons. Typically, the amino acid sequence is encoded by the nucleic acid using the "universal" genetic code. However, variants of the universal code, such as is present in some plant, animal, and fungal mitochondria, the bacterium *Mycoplasma capricolum* (*Proc. Natl. Acad. Sci. (USA)*, 82: 2306-2309 (1985)), or the ciliate *Macronucleus*, may be used when the nucleic acid is expressed using these organisms.

When the nucleic acid is prepared or altered synthetically, advantage can be taken of known codon preferences of the intended host where the nucleic acid is to be expressed. For example, although nucleic acid sequences of the present invention may be expressed in both monocotyledonous and dicotyledonous plant species, sequences can be modified to account for the specific codon preferences and GC content preferences of monocotyledonous plants or dicotyledonous plants as these preferences have been shown to differ (Murray *et al.* *Nucl. Acids Res.* 17: 477-498 (1989) and herein incorporated by reference). Thus, the maize preferred codon for a particular amino acid might be derived from known gene sequences from maize. Maize codon usage for 28 genes from maize plants is listed in Table 4 of Murray *et al.*, *supra*.

As used herein, "heterologous" in reference to a nucleic acid is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by deliberate human intervention. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form. A heterologous protein may originate from a foreign species or, if from the same species, is substantially modified from its original form by deliberate human intervention.

By "host cell" or "recombinantly engineered cell" is meant a cell, which contains a vector and supports the replication and/or expression of the expression vector. Host cells

may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, *Pichia*, insect, plant, amphibian, or mammalian cells. Preferably, host cells are monocotyledonous or dicotyledonous plant cells, including but not limited to maize, sorghum, sunflower, soybean, wheat, alfalfa, rice, cotton, canola, barley, millet, and tomato. A particularly  
5 preferred monocotyledonous host cell is a maize host cell.

The term "hybridization complex" includes reference to a duplex nucleic acid structure formed by two single-stranded nucleic acid sequences selectively hybridized with each other.

10 The term "introduced" in the context of inserting a nucleic acid into a cell, means "transfection" or "transformation" or "transduction" and includes reference to the incorporation of a nucleic acid into a eukaryotic or prokaryotic cell where the nucleic acid may be incorporated into the genome of the cell (e.g., chromosome, plasmid, plastid or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (e.g., transfected mRNA).

15 The term "isolated" refers to material, such as a nucleic acid or a protein, which is substantially or essentially free from components which normally accompany or interact with it as found in its naturally occurring environment. The isolated material optionally comprises material not found with the material in its natural environment. Nucleic acids, which are "isolated", as defined herein, are also referred to as "heterologous" nucleic acids.

20 Unless otherwise stated, the term "APAO nucleic acid" means a nucleic acid comprising a polynucleotide ("APAO polynucleotide") encoding an APAO polypeptide. The term APAO, unless otherwise stated can encompass both APAO and the functional, truncated version of APAO designated trAPAO.

25 As used herein, "nucleic acid" includes reference to a deoxyribonucleotide or ribonucleotide polymer in either single- or double-stranded form, and unless otherwise limited, encompasses known analogues having the essential nature of natural nucleotides in that they hybridize to single-stranded nucleic acids in a manner similar to naturally occurring nucleotides (e.g., peptide nucleic acids).

30 By "nucleic acid library" is meant a collection of isolated DNA or RNA molecules, which comprise and substantially represent the entire transcribed fraction of a genome of a specified organism. Construction of exemplary nucleic acid libraries, such as genomic and cDNA libraries, is taught in standard molecular biology references such as Berger and

Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology*, Vol. 152, Academic Press, Inc., San Diego, CA (Berger); Sambrook *et al.*, *Molecular Cloning - A Laboratory Manual*, 2nd ed., Vol. 1-3 (1989); and *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (1994 Supplement).

As used herein "operably linked" includes reference to a functional linkage between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. Generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in the same reading frame.

The term "ligated" or "ligation" refers to the process of forming phosphodiester bonds between two or more polynucleotides, which most often are double stranded DNAs. Techniques for ligation are well known in the art and protocols are described in standard laboratory manuals and references, such as, Sambrook, *et al. Molecular Cloning: A Laboratory Manual*, 2ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1989). The two polynucleotides can include, but are not limited to, a polynucleotide, which can function as a promoter, ligated to a polynucleotide capable of encoding a polypeptide or linking two polynucleotides each capable of encoding a polypeptide. In the case of joining two polynucleotides that each encode a polypeptide, a polynucleotide spacer region between the two polynucleotides may or may not be present. The spacer region may encode a polypeptide containing a protease cleavage site. Optionally, the spacer region may contain a polynucleotide cleavage site such as but not limited to a site for RNase cleavage or a self-cleaving ribozyme (See, e.g., Tanner, *FEMS Microbiol Rev*, 23(3):257-75 (1999)). Alternatively, the transcription of the two or more ligated polynucleotides may result in a polycistronic message. An example of a spacer sequence that would direct translation of downstream coding sequences is an intervening ribosomal entry site (IRES) (See, e.g., Liu, *et al.*, *Anal Biochem*, 280(1):20-28 (2000)). The length of the spacer region may be of any length that results in a functional polypeptide or polypeptides. For example, the spacer region may be from 1 nucleotide to 1000 nucleotides, preferably 24 nucleotides in length.

As used herein, the term "plant" includes reference to whole plants, plant organs (e.g., leaves, stems, roots, etc.), seeds and plant cells and progeny of same. Plant cell, as used herein includes, without limitation, seeds suspension cultures, embryos, meristematic regions, callus tissue, leaves, roots, shoots, gametophytes, sporophytes, pollen, and microspores. The class of plants, which can be used in the methods of the invention, is generally as broad as the class of higher plants amenable to transformation techniques, including both monocotyledonous and dicotyledonous plants including species from the genera: *Cucurbita*, *Rosa*, *Vitis*, *Juglans*, *Fragaria*, *Lotus*, *Medicago*, *Onobrychis*, *Trifolium*, *Trigonella*, *Vigna*, *Citrus*, *Linum*, *Geranium*, *Manihot*, *Daucus*, *Arabidopsis*,  
5 *Brassica*, *Raphanus*, *Sinapis*, *Atropa*, *Capsicum*, *Datura*, *Hyoscyamus*, *Lycopersicon*,  
10 *Nicotiana*, *Solanum*, *Petunia*, *Digitalis*, *Majorana*, *Ciahorium*, *Helianthus*, *Lactuca*,  
*Bromus*, *Asparagus*, *Antirrhinum*, *Heterocallis*, *Nemesis*, *Pelargonium*, *Panieum*,  
*Pennisetum*, *Ranunculus*, *Senecio*, *Salpiglossis*, *Cucumis*, *Browaalia*, *Glycine*, *Pisum*,  
*Phaseolus*, *Lolium*, *Oryza*, *Avena*, *Hordeum*, *Secale*, *Allium*, and *Triticum*. A particularly  
15 preferred plant is *Zea mays*.

As used herein, "polynucleotide" includes reference to a deoxyribopolynucleotide, ribopolynucleotide, or analogs thereof that have the essential nature of a natural ribonucleotide in that they hybridize, under stringent hybridization conditions, to substantially the same nucleotide sequence as naturally occurring nucleotides and/or allow  
20 translation into the same amino acid(s) as the naturally occurring nucleotide(s). A polynucleotide can be full-length or a subsequence of a native or heterologous structural or regulatory gene. Unless otherwise indicated, the term includes reference to the specified sequence as well as the complementary sequence thereof. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides" as that term is intended herein. Moreover, DNAs or RNAs comprising unusual bases, such as inosine, or  
25 modified bases, such as tritylated bases, to name just two examples, are polynucleotides as the term is used herein. It will be appreciated that a great variety of modifications have been made to DNA and RNA that serve many useful purposes known to those of skill in the art. The term polynucleotide as it is employed herein embraces such chemically, enzymatically  
30 or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including *inter alia*, simple and complex cells.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers.

5 As used herein "promoter" includes reference to a region of DNA upstream from the start of transcription and involved in recognition and binding of RNA polymerase and other proteins to initiate transcription. A "plant promoter" is a promoter capable of initiating transcription in plant cells. Exemplary plant promoters include, but are not limited to, those that are obtained from plants, plant viruses, and bacteria which comprise 10 genes expressed in plant cells such *Agrobacterium* or *Rhizobium*. Examples are promoters that preferentially initiate transcription in certain tissues, such as leaves, roots, seeds, fibres, xylem vessels, tracheids, or sclerenchyma. Such promoters are referred to as "tissue preferred". A "cell type" specific promoter primarily drives expression in certain cell types in one or more organs, for example, vascular cells in roots or leaves. An "inducible" or 15 "regulatable" promoter is a promoter, which is under environmental control. Examples of environmental conditions that may effect transcription by inducible promoters include anaerobic conditions or the presence of light. Another type of promoter is a developmentally regulated promoter, for example, a promoter that drives expression during pollen development. Tissue preferred, cell type specific, developmentally regulated, and 20 inducible promoters constitute the class of "non-constitutive" promoters. A "constitutive" promoter is a promoter, which is active under most environmental conditions.

The term "APAO polypeptide or trAPAO polypeptide" refers to one or more amino acid sequences. The term is also inclusive of fragments, variants, homologs, alleles or precursors (e.g., preproproteins or proproteins) thereof. An "APAO or trAPAO protein" 25 comprises an APAO or trAPAO polypeptide.

As used herein "recombinant" includes reference to a cell or vector, that has been modified by the introduction of a heterologous nucleic acid or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found in identical form within the native (non-recombinant) form of the cell or express native 30 genes that are otherwise abnormally expressed, under expressed or not expressed at all as a result of deliberate human intervention. The term "recombinant" as used herein does not encompass the alteration of the cell or vector by naturally occurring events (e.g.,

spontaneous mutation, natural transformation/transduction/transposition) such as those occurring without deliberate human intervention.

As used herein, a "recombinant expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements, 5 which permit transcription of a particular nucleic acid in a target cell. The recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid to be transcribed, and a promoter.

10 The term "residue" or "amino acid residue" or "amino acid" are used interchangeably herein to refer to an amino acid that is incorporated into a protein, polypeptide, or peptide (collectively "protein"). The amino acid may be a naturally occurring amino acid and, unless otherwise limited, may encompass known analogs of natural amino acids that can function in a similar manner as naturally occurring amino 15 acids.

The term "selectively hybridizes" includes reference to hybridization, under stringent hybridization conditions, of a nucleic acid sequence to a specified nucleic acid target sequence to a detectably greater degree (e.g., at least 2-fold over background) than its hybridization to non-target nucleic acid sequences and to the substantial exclusion of 20 non-target nucleic acids. Selectively hybridizing sequences typically have about at least 40% sequence identity, preferably 60-90% sequence identity, and most preferably 100% sequence identity (i.e., complementary) with each other.

The terms "stringent conditions" or "stringent hybridization conditions" include reference to conditions under which a probe will hybridize to its target sequence, to a 25 detectably greater degree than other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences can be identified which can be up to 100% complementary to the probe (homologous probing). Alternatively, stringency conditions can be adjusted to 30 allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Optimally, the probe is approximately 500 nucleotides in length,

but can vary greatly in length from less than 500 nucleotides to equal to the entire length of the target sequence.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide or Denhardt's. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the  $T_m$  can be approximated from the equation of Meinkoth and Wahl, *Anal. Biochem.*, 138:267-284 (1984):  $T_m = 81.5 \text{ } ^\circ\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61 (\% \text{ form}) - 500/L$ ; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe.  $T_m$  is reduced by about 1 °C for each 1% of mismatching; thus,  $T_m$ , hybridization and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with ≥90% identity are sought, the  $T_m$  can be decreased 10 °C. Generally, stringent conditions are selected to be about 5 °C lower than the thermal melting point ( $T_m$ ) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4 °C lower than the thermal melting point ( $T_m$ ); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10 °C lower than the thermal melting point ( $T_m$ ); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20 °C

lower than the thermal melting point ( $T_m$ ). Using the equation, hybridization and wash compositions, and desired  $T_m$ , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a  $T_m$  of less than 45 °C (aqueous solution) or 32 °C  
5 (formamide solution) it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen, *Laboratory Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 "Overview of principles of hybridization and the strategy of nucleic acid probe assays", Elsevier, New York (1993); and *Current  
10 Protocols in Molecular Biology*, Chapter 2, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995). Unless otherwise stated, in the present application high stringency is defined as hybridization in 4X SSC, 5X Denhardt's (5g Ficoll, 5g polyvinylpyrrolidone, 5 g bovine serum albumin in 500ml of water), 0.1 mg/ml boiled salmon sperm DNA, and 25 mM Na phosphate at 65°C, and a wash in 0.1X SSC, 0.1%  
15 SDS at 65°C.

As used herein, "transgenic plant" includes reference to a plant, which comprises within its genome a heterologous polynucleotide. Generally, the heterologous polynucleotide is stably integrated within the genome such that the polynucleotide is passed on to successive generations. The heterologous polynucleotide may be integrated into the genome alone or as part of a recombinant expression cassette. "Transgenic" is used herein to include any cell, cell line, callus, tissue, plant part or plant, the genotype of which has been altered by the presence of heterologous nucleic acid including those transgenics initially so altered as well as those created by sexual crosses or asexual propagation from the initial transgenic. The term "transgenic" as used herein does not encompass the alteration of the genome (chromosomal or extra-chromosomal) by conventional plant breeding methods or by naturally occurring events such as random cross-fertilization, non-recombinant viral infection, non-recombinant bacterial transformation, non-recombinant transposition, or spontaneous mutation.  
20  
25

As used herein, "vector" includes reference to a nucleic acid used in transfection of a host cell and into which can be inserted a polynucleotide. Vectors are often replicons. Expression vectors permit transcription of a nucleic acid inserted therein.  
30

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides or polypeptides: (a) “reference sequence”, (b) “comparison window”, (c) “sequence identity”, (d) “percentage of sequence identity”, and (e) “substantial identity”.

5 (a) As used herein, “reference sequence” is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

10 (b) As used herein, “comparison window” means includes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence may be compared to a reference sequence and wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

20 Methods of alignment of nucleotide and amino acid sequences for comparison are well known in the art. The local homology algorithm (Best Fit) of Smith and Waterman, Adv. Appl. Math may conduct optimal alignment of sequences for comparison. 2: 482 (1981); by the homology alignment algorithm (GAP) of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970); by the search for similarity method (Tfasta and Fasta) of Pearson and Lipman, *Proc. Natl. Acad. Sci.* 85: 2444 (1988); by computerized implementations of these 25 algorithms, including, but not limited to: CLUSTAL in the PC/Gene program by Intelligenetics, Mountain View, California, GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, Wisconsin, USA; the CLUSTAL program is well described by Higgins and Sharp, *Gene* 73: 237-244 (1988); Higgins and Sharp, *CABIOS* 5: 151-153 (1989); Corpet, *et al.*, *Nucleic Acids Research* 16: 10881-90 (1988); Huang, *et al.*, *Computer Applications in the Biosciences* 8: 155-65 (1992), and Pearson, *et al.*, *Methods in Molecular Biology* 24: 307-331 (1994). The preferred program to use for optimal

global alignment of multiple sequences is PileUp (Feng and Doolittle, *Journal of Molecular Evolution*, 25:351-360 (1987) which is similar to the method described by Higgins and Sharp, *CABIOS*, 5:151-153 (1989) and hereby incorporated by reference). The BLAST family of programs which can be used for database similarity searches includes:

- 5 BLASTN for nucleotide query sequences against nucleotide database sequences; BLASTX  
for nucleotide query sequences against protein database sequences; BLASTP for protein  
query sequences against protein database sequences; TBLASTN for protein query  
sequences against nucleotide database sequences; and TBLASTX for nucleotide query  
sequences against nucleotide database sequences. See, *Current Protocols in Molecular  
10 Biology*, Chapter 19, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New  
York (1995).

GAP uses the algorithm of Needleman and Wunsch (J. Mol. Biol. 48: 443-453,  
1970) to find the alignment of two complete sequences that maximizes the number of  
matches and minimizes the number of gaps. GAP considers all possible alignments and  
15 gap positions and creates the alignment with the largest number of matched bases and the  
fewest gaps. It allows for the provision of a gap creation penalty and a gap extension  
penalty in units of matched bases. GAP must make a profit of gap creation penalty number  
of matches for each gap it inserts. If a gap extension penalty greater than zero is chosen,  
GAP must, in addition, make a profit for each gap inserted of the length of the gap times  
20 the gap extension penalty. Default gap creation penalty values and gap extension penalty  
values in Version 10 of the Wisconsin Genetics Software Package are 8 and 2,  
respectively. The gap creation and gap extension penalties can be expressed as an integer  
selected from the group of integers consisting of from 0 to 100. Thus, for example, the gap  
creation and gap extension penalties can be 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 30, 40, 50,  
25 or greater.

GAP presents one member of the family of best alignments. There may be many  
members of this family, but no other member has a better quality. GAP displays four  
figures of merit for alignments: Quality, Ratio, Identity, and Similarity. The Quality is the  
metric maximized in order to align the sequences. Ratio is the quality divided by the  
30 number of bases in the shorter segment. Percent Identity is the percent of the symbols that  
actually match. Percent Similarity is the percent of the symbols that are similar. Symbols  
that are across from gaps are ignored. A similarity is scored when the scoring matrix value

for a pair of symbols is greater than or equal to 0.50, the similarity threshold. The scoring matrix used in Version 10 of the Wisconsin Genetics Software Package is BLOSUM62 (see Henikoff & Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

Unless otherwise stated, sequence identity/similarity values provided herein refer to  
5 the value obtained using the BLAST 2.0 suite of programs using default parameters.  
Altschul *et al.*, *Nucleic Acids Res.* 25:3389-3402 (1997).

As those of ordinary skill in the art will understand, BLAST searches assume that  
proteins can be modeled as random sequences. However, many real proteins comprise  
regions of nonrandom sequences, which may be homopolymeric tracts, short-period  
10 repeats, or regions enriched in one or more amino acids. Such low-complexity regions  
may be aligned between unrelated proteins even though other regions of the protein are  
entirely dissimilar. A number of low-complexity filter programs can be employed to  
reduce such low-complexity alignments. For example, the SEG (Wooten and Federhen,  
*Comput. Chem.*, 17:149-163 (1993)) and XNU (Claverie and States, *Comput. Chem.*,  
15 17:191-201 (1993)) low-complexity filters can be employed alone or in combination.

(c) As used herein, “sequence identity” or “identity” in the context of two nucleic  
acid or polypeptide sequences includes reference to the residues in the two sequences,  
which are the same when aligned for maximum correspondence over a specified  
comparison window. When percentage of sequence identity is used in reference to  
20 proteins it is recognized that residue positions which are not identical often differ by  
conservative amino acid substitutions, where amino acid residues are substituted for other  
amino acid residues with similar chemical properties (e.g. charge or hydrophobicity) and  
therefore do not change the functional properties of the molecule. Where sequences differ  
in conservative substitutions, the percent sequence identity may be adjusted upwards to  
25 correct for the conservative nature of the substitution. Sequences, which differ by such  
conservative substitutions, are said to have “sequence similarity” or “similarity”. Means  
for making this adjustment are well known to those of skill in the art. Typically this  
involves scoring a conservative substitution as a partial rather than a full mismatch,  
thereby increasing the percentage sequence identity. Thus, for example, where an identical  
30 amino acid is given a score of 1 and a non-conservative substitution is given a score of  
zero, a conservative substitution is given a score between zero and 1. The scoring of  
conservative substitutions is calculated, *e.g.*, according to the algorithm of Meyers and

Miller, *Computer Applic. Biol. Sci.*, 4: 11-17 (1988) e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California, USA).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

(e) (i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has between 50-100% sequence identity, preferably at least 50% sequence identity, preferably at least 60% sequence identity, preferably at least 70%, more preferably at least 80%, more preferably at least 90% and most preferably at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of between 40-100%, preferably at least 55%, preferably at least 60%, more preferably at least 70%, 80%, 90%, and most preferably at least 95%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. The degeneracy of the genetic code allows for many amino acids substitutions that lead to variety in the nucleotide sequence that code for the same amino acid, hence it is possible that the DNA sequence could code for the same polypeptide but not hybridize to each other under stringent conditions. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is that the polypeptide, which the first

nucleic acid encodes, is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e) (ii) The terms "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with between 55-100% sequence identity to a reference sequence preferably at least 55% sequence identity, preferably 60% preferably 70%, more preferably 80%, most preferably at least 90% or 95% sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970). An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution. In addition, a peptide can be substantially identical to a second peptide when they differ by a non-conservative change if the epitope that the antibody recognizes is substantially identical. Peptides, which are "substantially similar" share sequences as, noted above except that residue positions, which are not identical, may differ by conservative amino acid changes.

### Fumonisin Degrading Organisms

The present invention is based on the discovery of organisms with the ability to degrade the mycotoxin fumonisin. In a search for a biological means of detoxifying fumonisins, several dematiaceous hyphomycetes were isolated from field-grown maize kernels. The fungi were found to be capable of growing on fumonisin B1 or B2 (FB1 or FB2) as a sole carbon source, degrading it partially or completely in the process. One species, identified as *Exophiala spinifera*, a "black yeast", was recovered from maize seed from diverse locations in the southeastern and south central US. The enzyme-active strain of *Exophiala spinifera* (ATCC 74269) was deposited (see US patent no. 5,716,820, issued February 10, 1998, US patent no. 5,792,931, issued August 11, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997). Other enzyme-active strains of *Exophiala spinifera* were used to isolate APAO polynucleotides. Isolate ESP002 was isolated from palm trees (ATCC 26089) and isolate ESP003 was isolated from maize seed. Another fungus from which APAO polynucleotides were isolated was *Rhinocladiella atrovirens* (RAT 011).

## Nucleic Acids

The present invention provides, *inter alia*, isolated nucleic acids of RNA, DNA, and analogs and/or chimeras thereof, comprising an APAO or trAPAO polynucleotide.

5 The present invention also includes polynucleotides optimized for expression in different organisms. For example, for expression of the polynucleotide in a maize plant, the sequence can be altered to account for specific codon preferences and to alter GC content as according to Murray *et al., supra*. Maize codon usage for 28 genes from maize plants is listed in Table 4 of Murray, *et al., supra*.

10 The APAO or trAPAO nucleic acids of the present invention comprise isolated APAO or trAPAO polynucleotides which, are inclusive of:

(a) a polynucleotide encoding an APAO or trAPAO polypeptide of the sequences shown in SEQ ID NOS: 36, 38, 40, 42, 44, and 46, and conservatively modified and polymorphic variants thereof;

15 (b) a polynucleotide which selectively hybridizes to a polynucleotide of (a) or (b);  
(c) a polynucleotide having at least 50% sequence identity with polynucleotides of (a) or (b);  
(d) complementary sequences of polynucleotides of (a), (b), or (c); and  
(e) a polynucleotide comprising at least 25 contiguous nucleotides from a  
20 polynucleotide of (a), (b), (c), or (d).

In addition, polynucleotides are presented that are a fusion of an APAO or trAPAO polynucleotide and the polynucleotide of a fumonisin esterase. The invention encompasses the sequences from *Exophiala* or *Rhinocladiella* as well as sequences having sequence similarity with such sequences. It is recognized that the sequences of the  
25 invention can be used to isolate corresponding sequences in other organisms. Methods such as PCR, hybridization, and the like can be used to identify sequences having substantial sequence similarity to the sequences of the invention. See, for example, Sambrook, *et al.*, (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Planview, New York) and Innis *et al.*, (1990) *PCR Protocols: Guide to Methods and Applications* (Academic Press, New York). Coding sequences isolated based on their sequence identity to the entire fumonisin degrading coding

sequences set forth herein or to fragments thereof are encompassed by the present invention.

It is recognized that the sequences of the invention can be used to isolate similar sequences from other fumonisin degrading organisms. Likewise sequences from other 5 fumonisin degrading organisms may be used in combination with the sequences of the present invention. See, for example, copending application entitled "Compositions and Methods for Fumonisin Detoxification", U.S. application serial number 60/092,953, filed concurrently herewith and herein incorporated by reference.

Plasmids containing the polynucleotide sequences of the invention were deposited 10 with American Type Culture Collection (ATCC), Manassas, Virginia, and assigned Accession Nos. 98812, 98813, 98814, 98815, 98816, and PTA-32. These deposits will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. These deposits were made merely as a convenience for those of skill in the art and are not an admission that a 15 deposit is required under 35 U.S.C. § 112.

### **Construction of Nucleic Acids**

The isolated nucleic acids of the present invention can be made using (a) standard recombinant methods, (b) synthetic techniques, or combinations thereof. In some 20 embodiments, the polynucleotides of the present invention will be cloned, amplified, or otherwise constructed from a fungus or bacteria.

The nucleic acids may conveniently comprise sequences in addition to a polynucleotide of the present invention. For example, a multi-cloning site comprising one or more endonuclease restriction sites may be inserted into the nucleic acid to aid in 25 isolation of the polynucleotide. Also, translatable sequences may be inserted to aid in the isolation of the translated polynucleotide of the present invention. For example, a hexahistidine marker sequence provides a convenient means to purify the proteins of the present invention. The nucleic acid of the present invention - excluding the polynucleotide sequence - is optionally a vector, adapter, or linker for cloning and/or expression of a 30 polynucleotide of the present invention. Additional sequences may be added to such cloning and/or expression sequences to optimize their function in cloning and/or expression, to aid in isolation of the polynucleotide, or to improve the introduction of the

polynucleotide into a cell. Typically, the length of a nucleic acid of the present invention less than the length of its polynucleotide of the present invention is less than 20 kilobase pairs, often less than 15 kb, and frequently less than 10 kb. Use of cloning vectors, expression vectors, adapters, and linkers is well known in the art. Exemplary nucleic acids include such vectors as: M13, lambda ZAP Express, lambda ZAP II, lambda gt10, lambda gt11, pBK-CMV, pBK-RSV, pBluescript II, lambda DASH II, lambda EMBL 3, lambda EMBL 4, pWE15, SuperCos 1, SurfZap, Uni-ZAP, pBC, pBS+/-, pSG5, pBK, pCR-Script, pET, pSPORTK, p3'SS, pGEM, pSK+/-, pGEX, pSPORTI and II, pOPRSVI CAT, pOPI3 CAT, pXT1, pSG5, pPbac, pMbac, pMC1neo, pOG44, pOG45, pFRT $\beta$ GAL, pNEO $\beta$ GAL, pRS403, pRS404, pRS405, pRS406, pRS413, pRS414, pRS415, pRS416, lambda MOSSlox, and lambda MOSElox. Optional vectors for the present invention, include but are not limited to, lambda ZAP II, and pGEX. For a description of various nucleic acids see, for example, Stratagene Cloning Systems, Catalogs 1995, 1996, 1997 (La Jolla, CA); and, Amersham Life Sciences, Inc, Catalog '97 (Arlington Heights, IL).

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### Synthetic Methods for Constructing Nucleic Acids

The isolated nucleic acids of the present invention can also be prepared by direct chemical synthesis by methods such as the phosphotriester method of Narang *et al.*, *Meth. Enzymol.* 68: 90-99 (1979); the phosphodiester method of Brown *et al.*, *Meth. Enzymol.* 68: 109-151 (1979); the diethylphosphoramidite method of Beaucage *et al.*, *Tetra. Lett.* 22: 1859-1862 (1981); the solid phase phosphoramidite triester method described by Beaucage and Caruthers, *Tetra. Letts.* 22(20): 1859-1862 (1981), e.g., using an automated synthesizer, e.g., as described in Needham-VanDevanter *et al.*, *Nucleic Acids Res.*, 12: 6159-6168 (1984); and, the solid support method of US Patent No. 4,458,066. Chemical synthesis generally produces a single stranded oligonucleotide. This may be converted into double stranded DNA by hybridization with a complementary sequence, or by polymerization with a DNA polymerase using the single strand as a template. One of skill will recognize that while chemical synthesis of DNA is limited to sequences of about 100 bases, longer sequences may be obtained by the ligation of shorter sequences.

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## **UTRs and Codon Preference**

In general, translational efficiency has been found to be regulated by specific sequence elements in the 5' non-coding or untranslated region (5' UTR) of the RNA. Positive sequence motifs include translational initiation consensus sequences (Kozak, 5 *Nucleic Acids Res.* 15:8125 (1987)) and the 5<G> 7 methyl GpppG RNA cap structure (Drummond *et al.*, *Nucleic Acids Res.* 13:7375 (1985)). Negative elements include stable intramolecular 5' UTR stem-loop structures (Muesing *et al.*, *Cell* 48:691 (1987)) and AUG sequences or short open reading frames preceded by an appropriate AUG in the 5' UTR (Kozak, *supra*, Rao *et al.*, *Mol. and Cell. Biol.* 8:284 (1988)). Accordingly, the present 10 invention provides 5' and/or 3' UTR regions for modulation of translation of heterologous coding sequences.

Further, the polypeptide-encoding segments of the polynucleotides of the present invention can be modified to alter codon usage. Altered codon usage can be employed to alter translational efficiency and/or to optimize the coding sequence for expression in a 15 desired host or to optimize the codon usage in a heterologous sequence for expression in maize. Codon usage in the coding regions of the polynucleotides of the present invention can be analyzed statistically using commercially available software packages such as "Codon Preference" available from the University of Wisconsin Genetics Computer Group (see Devereaux *et al.*, *Nucleic Acids Res.* 12: 387-395 (1984)) or MacVector 4.1 (Eastman 20 Kodak Co., New Haven, Conn.). Thus, the present invention provides a codon usage frequency characteristic of the coding region of at least one of the polynucleotides of the present invention. The number of polynucleotides (3 nucleotides per amino acid) that can be used to determine a codon usage frequency can be any integer from 3 to the number of polynucleotides of the present invention as provided herein. Optionally, the 25 polynucleotides will be full-length sequences. An exemplary number of sequences for statistical analysis can be at least 1, 5, 10, 20, 50, or 100.

## **Sequence Shuffling**

The present invention provides methods for sequence shuffling using 30 polynucleotides of the present invention, and compositions resulting therefrom. Sequence shuffling is described in PCT publication No. 96/19256. See also, Zhang, J.- H., *et al.* *Proc. Natl. Acad. Sci. USA* 94:4504-4509 (1997) and Zhao, *et al.*, *Nature Biotech* 16:258-

261 (1998). Generally, sequence shuffling provides a means for generating libraries of polynucleotides having a desired characteristic, which can be selected or screened for. Libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides , which comprise sequence regions, which have substantial 5 sequence identity and can be homologously recombined in vitro or in vivo. The population of sequence-recombined polynucleotides comprises a subpopulation of polynucleotides which possess desired or advantageous characteristics and which can be selected by a suitable selection or screening method. The characteristics can be any property or attribute capable of being selected for or detected in a screening system, and 10 may include properties of: an encoded protein, a transcriptional element, a sequence controlling transcription, RNA processing, RNA stability, chromatin conformation, translation, or other expression property of a gene or transgene, a replicative element, a protein-binding element, or the like, such as any feature which confers a selectable or detectable property. In some embodiments, the selected characteristic will be an altered 15  $K_m$  and/or  $K_{cat}$  over the wild-type protein as provided herein. In other embodiments, a protein or polynucleotide generated from sequence shuffling will have a substrate binding affinity greater than the non-shuffled wild-type polynucleotide. In yet other embodiments, a protein or polynucleotide generated from sequence shuffling will have an altered pH optimum as compared to the non-shuffled wild-type polynucleotide. The increase in such 20 properties can be at least 110%, 120%, 130%, 140% or greater than 150% of the wild-type value.

### **Recombinant Expression Cassettes**

The present invention further provides recombinant expression cassettes comprising a nucleic acid of the present invention. A nucleic acid sequence coding for the 25 desired polynucleotide of the present invention, for example a cDNA or a genomic sequence encoding a polypeptide long enough to code for an active protein of the present invention, can be used to construct a recombinant expression cassette which can be introduced into the desired host cell. A recombinant expression cassette will typically 30 comprise a polynucleotide of the present invention operably linked to transcriptional initiation regulatory sequences which will direct the transcription of the polynucleotide in the intended host cell, such as tissues of a transformed plant.

For example, plant expression vectors may include (1) a cloned plant gene under the transcriptional control of 5' and 3' regulatory sequences and (2) a dominant selectable marker. Such plant expression vectors may also contain, if desired, a promoter regulatory region (e.g., one conferring inducible or constitutive, environmentally- or 5 developmentally-regulated, or cell- or tissue-specific/selective expression), a transcription initiation start site, a ribosome binding site, an RNA processing signal, a transcription termination site, and/or a polyadenylation signal.

A plant promoter fragment can be employed which will direct expression of a polynucleotide of the present invention in all tissues of a regenerated plant. Such 10 promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the 1'- or 2'- promoter derived from T-DNA of *Agrobacterium tumefaciens*, the Smas promoter, the cinnamyl alcohol dehydrogenase promoter (US Patent No. 5,683,439), the Nos promoter, the rubisco promoter, the GRP1-8 15 promoter, the 35S promoter from cauliflower mosaic virus (CaMV), as described in Odell et al., (1985), *Nature*, 313:810-812, rice actin (McElroy et al., (1990), *Plant Cell*, 163-171); ubiquitin (Christensen et al., (1992), *Plant Mol. Biol.* 12:619-632; and Christensen, et al., (1992), *Plant Mol. Biol.* 18:675-689); pEMU (Last, et al., (1991), *Theor. Appl. Genet.* 81:581-588); MAS (Velten et al., (1984), *EMBO J.* 3:2723-2730); and maize H3 20 histone (Lepetit et al., (1992), *Mol. Gen. Genet.* 231:276-285; and Atanassvoa et al., (1992), *Plant Journal* 2(3):291-300), the Rsyn7 as described in published PCT Application WO 97/44756, ALS promoter, as described in published PCT Application WO 96/30530, and other transcription initiation regions from various plant genes known to those of skill. 25 For the present invention ubiquitin is the preferred promoter for expression in monocot plants.

Alternatively, the plant promoter can direct expression of a polynucleotide of the present invention in a specific tissue or may be otherwise under more precise environmental or developmental control. Such promoters are referred to here as "inducible" promoters. Environmental conditions that may effect transcription by 30 inducible promoters include pathogen attack, anaerobic conditions, or the presence of light. Examples of inducible promoters are the Adh1 promoter, which is inducible by hypoxia or

cold stress, the Hsp70 promoter, which is inducible by heat stress, and the PPDK promoter, which is inducible by light.

Examples of promoters under developmental control include promoters that initiate transcription only, or preferentially, in certain tissues, such as leaves, roots, fruit, seeds, or flowers. The operation of a promoter may also vary depending on its location in the genome. Thus, an inducible promoter may become fully or partially constitutive in certain locations.

If polypeptide expression is desired, it is generally desirable to include a polyadenylation region at the 3'-end of a polynucleotide coding region. The polyadenylation region can be derived from a variety of plant genes, or from T-DNA. The 3' end sequence to be added can be derived from, for example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene. Examples of such regulatory elements include, but are not limited to, 3' termination and/or polyadenylation regions such as those of the *Agrobacterium tumefaciens* nopaline synthase (nos) gene (Bevan et al., (1983), *Nucl. Acids Res.* 12:369-385); the potato proteinase inhibitor II (PINII) gene (Keil, et al., (1986), *Nucl. Acids Res.* 14:5641-5650; and An et al., (1989), *Plant Cell* 1:115-122); and the CaMV 19S gene (Mogen et al., (1990), *Plant Cell* 2:1261-1272).

An intron sequence can be added to the 5' untranslated region or the coding sequence of the partial coding sequence to increase the amount of the mature message that accumulates in the cytosol. Inclusion of a spliceable intron in the transcription unit in both plant and animal expression constructs has been shown to increase gene expression at both the mRNA and protein levels up to 1000-fold. Buchman and Berg, *Mol. Cell Biol.* 8: 4395-4405 (1988); Callis et al., *Genes Dev.* 1: 1183-1200 (1987). Such intron enhancement of gene expression is typically greatest when placed near the 5' end of the transcription unit. Use of maize introns Adh1-S intron 1, 2, and 6, the Bronze-1 intron are known in the art. See generally, *The Maize Handbook*, Chapter 116, Freeling and Walbot, Eds., Springer, New York (1994).

Plant signal sequences, including, but not limited to, signal-peptide encoding DNA/RNA sequences which target proteins to the extracellular matrix of the plant cell (Dratewka-Kos, et al., (1989), *J. Biol. Chem.* 264:4896-4900), the *Nicotiana plumbaginifolia* extension gene (DeLoose, et al., (1991), *Gene* 99:95-100), signal peptides

which target proteins to the vacuole like the sweet potato sporamin gene (Matsuka, et al., (1991), *PNAS* 88:834) and the barley lectin gene (Wilkins, et al., (1990), *Plant Cell*, 2:301-313), signal peptides which cause proteins to be secreted such as that of PR1b (Lind, et al., (1992), *Plant Mol. Biol.* 18:47-53), or the barley alpha amylase (BAA) (Rahmatullah, et al., *Plant Mol. Biol.* 12:119 (1989)) and hereby incorporated by reference), or from the present invention the signal peptide from the ESP1 or BEST1 gene, or signal peptides which target proteins to the plastids such as that of rapeseed enoyl-Acp reductase (Verwaert, et al., (1994), *Plant Mol. Biol.* 26:189-202) are useful in the invention. The barley alpha amylase signal sequence operably linked to the trAPAO or APAO polynucleotide is the preferred construct for expression in maize for the present invention.

The vector comprising the sequences from a polynucleotide of the present invention will typically comprise a marker gene, which confers a selectable phenotype on plant cells. Usually, the selectable marker gene will encode antibiotic resistance, with suitable genes including genes coding for resistance to the antibiotic spectinomycin (e.g., the aada gene), the streptomycin phosphotransferase (SPT) gene coding for streptomycin resistance, the neomycin phosphotransferase (NPTII) gene encoding kanamycin or geneticin resistance, the hygromycin phosphotransferase (HPT) gene coding for hygromycin resistance, genes coding for resistance to herbicides which act to inhibit the action of acetolactate synthase (ALS), in particular the sulfonylurea-type herbicides (e.g., the acetolactate synthase (ALS) gene containing mutations leading to such resistance in particular the S4 and/or Hra mutations), genes coding for resistance to herbicides which act to inhibit action of glutamine synthase, such as phosphinothricin or basta (e.g., the *bar* gene), or other such genes known in the art. The *bar* gene encodes resistance to the herbicide basta, and the ALS gene encodes resistance to the herbicide chlorsulfuron.

Alternatively, the invention, itself, could be used as a method for selection of transformants, in other words as a selectable marker. An APAO or trAPAO polynucleotide operably linked to a promoter and then transformed into a plant cell by any of the methods described in the present application would express the degradative enzyme. When the plant cells are placed in the presence of fumonisin, AP1, or a phytotoxic analog in culture only the transformed cells would be able to grow. In another embodiment, the plant cell could be transformed with both a polynucleotide for APAO and a polynucleotide for fumonisin esterase. The selective agent in this case could be either AP1 or fumonisin

or any structural analog. Thus, growth of plant cells in the presence of a mycotoxin favors the survival of plant cells that have been transformed to express the coding sequence that codes for one of the enzymes of this invention and degrades the toxin. When the APAO or trAPAO cassette with or without the fumonisin esterase polynucleotide, is co-transformed 5 with another gene of interest and then placed in the presence of fumonisin, AP1 or a phytotoxic analog, this invention would allow for selection of only those plant cells that contain the gene of interest. In the past antibiotic resistance genes have been used as selectable markers. Given the current concerns by consumers and environmentalist over use of antibiotic genes and the possibility of resistant microorganisms arising due to this 10 use, a non-antibiotic resistant selectable marker system such as the present invention, fulfills this very important need.

Typical vectors useful for expression of genes in higher plants are well known in the art and include vectors derived from the tumor-inducing (Ti) plasmid of *Agrobacterium tumefaciens* described by Rogers *et al.*, Meth. In Enzymol., 153:253-277 (1987). These 15 vectors are plant integrating vectors in that on transformation, the vectors integrate a portion of vector DNA into the genome of the host plant. Exemplary *A. tumefaciens* vectors useful herein are plasmids pKYLX6 and pKYLX7 of Schardl *et al.*, Gene, 61:1-11 (1987) and Berger *et al.*, Proc. Natl. Acad. Sci. U.S.A., 86:8402-8406 (1989). Another 20 useful vector herein is plasmid pBI101.2 that is available from CLONTECH Laboratories, Inc. (Palo Alto, CA).

### **Expression of Proteins in Host Cells**

Using the nucleic acids of the present invention, one may express a protein of the present invention in a recombinantly engineered cell such as bacteria, yeast, insect, 25 mammalian, or preferably plant cells. The cells produce the protein in a non-natural condition (e.g., in quantity, composition, location, and/or time), because they have been genetically altered through human intervention to do so.

It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of a nucleic acid encoding a protein of the 30 present invention. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of isolated nucleic acids encoding a protein of the present invention will typically be achieved by operably linking, for example, the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding a protein of the present invention. To obtain high level expression of a cloned gene, it is desirable to construct expression vectors which contain, at the minimum, a strong promoter, such as ubiquitin, to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator. Constitutive promoters are classified as providing for a range of constitutive expression. Thus, some are weak constitutive promoters, and others are strong constitutive promoters. Generally, by "weak promoter" is intended a promoter that drives expression of a coding sequence at a low level. By "low level" is intended at levels of about 1/10,000 transcripts to about 1/100,000 transcripts to about 1/500,000 transcripts. Conversely, a "strong promoter" drives expression of a coding sequence at a "high level", or about 1/10 transcripts to about 1/100 transcripts to about 1/1,000 transcripts.

One of skill would recognize that modifications could be made to a protein of the present invention without diminishing its biological activity. Some modifications may be made to facilitate the cloning, expression, or incorporation of the targeting molecule into a fusion protein. Such modifications are well known to those of skill in the art and include, for example, a methionine added at the amino terminus to provide an initiation site, or additional amino acids (e.g., poly His) placed on either terminus to create conveniently located restriction sites or termination codons or purification sequences.

25

#### *A. Expression in Prokaryotes*

Prokaryotic cells may be used as hosts for expression. Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Commonly used prokaryotic control sequences which are defined herein to include 30 promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang et al., Nature 198:1056 (1977)),

the tryptophan (trp) promoter system (Goeddel et al., Nucleic Acids Res. 8:4057 (1980)) and the lambda derived P L promoter and N-gene ribosome binding site (Shimatake et al., Nature 292:128 (1981)). The inclusion of selection markers in DNA vectors transfected in *E. coli* is also useful. Examples of such markers include genes specifying resistance to 5 ampicillin, tetracycline, or chloramphenicol.

The vector is selected to allow introduction of the gene of interest into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected 10 with the plasmid vector DNA. Expression systems for expressing a protein of the present invention are available using *Bacillus sp.* and *Salmonella* (Palva, et al., Gene 22: 229-235 (1983); Mosbach, et al., Nature 302: 543-545 (1983)). The pGEX-4T-1 plasmid vector from Pharmacia is the preferred *E. coli* expression vector for the present invention.

15      *B. Expression in Eukaryotes*

A variety of eukaryotic expression systems such as yeast, insect cell lines, plant and mammalian cells, are known to those of skill in the art. As explained briefly below, the present invention can be expressed in these eukaryotic systems. In some embodiments, transformed/transfected plant cells, as discussed *infra*, are employed as expression systems 20 for production of the proteins of the instant invention.

Synthesis of heterologous proteins in yeast is well known. Sherman, F., et al., *Methods in Yeast Genetics*, Cold Spring Harbor Laboratory (1982) is a well recognized work describing the various methods available to produce the protein in yeast. Two widely utilized yeasts for production of eukaryotic proteins are *Saccharomyces cerevisiae* and 25 *Pichia pastoris*. Vectors, strains, and protocols for expression in *Saccharomyces* and *Pichia* are known in the art and available from commercial suppliers (e.g., Invitrogen). Suitable vectors usually have expression control sequences, such as promoters, including 3-phosphoglycerate kinase or alcohol oxidase, and an origin of replication, termination sequences and the like as desired.

30      A protein of the present invention, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates or the

pellets. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassay of other standard immunoassay techniques.

The sequences encoding proteins of the present invention can also be ligated to various expression vectors for use in transfecting cell cultures of, for instance, mammalian, insect, or plant origin. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. A number of suitable host cell lines capable of expressing intact proteins have been developed in the art, and include the HEK293, BHK21, and CHO cell lines. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (e.g., the CMV promoter, a HSV *tk* promoter or *pgk* (phosphoglycerate kinase) promoter), an enhancer (Queen *et al.*, *Immunol. Rev.* 89: 49 (1986)), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites (e.g., an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells useful for production of proteins of the present invention are available, for instance, from the American Type Culture Collection Catalogue of Cell Lines and Hybridomas (7th edition, 1992).

Appropriate vectors for expressing proteins of the present invention in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth, and *Drosophila* cell lines such as a Schneider cell line (See Schneider, *J. Embryol. Exp. Morphol.* 27: 353-365 (1987)).

As with yeast, when higher animal or plant host cells are employed, polyadenylation or transcription terminator sequences are typically incorporated into the vector. An example of a terminator sequence is the polyadenylation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VP1 intron from SV40 (Sprague, *et al.*, *J. Virol.* 45: 773-781 (1983)). Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors. Saveria-Campo, M., Bovine Papilloma Virus DNA a Eukaryotic Cloning Vector in *DNA Cloning Vol. II a Practical Approach*, D.M. Glover, Ed., IRL Press, Arlington, Virginia pp. 213-238 (1985).

In addition, one of the genes for fumonisins esterase or the APAO or trAPAO placed in the appropriate plant expression vector can be used to transform plant cells. The

enzyme can then be isolated from plant callus or the transformed cells can be used to regenerate transgenic plants. Such transgenic plants can be harvested, and the appropriate tissues (seed or leaves, for example) can be subjected to large scale protein extraction and purification techniques, and the fumonisin degradation enzymes or APAO can be isolated  
5 for use in fumonisin and fumonisin hydrolysis product detoxification processes.

### **Plant Transformation Methods**

Numerous methods for introducing foreign genes into plants are known and can be used to insert an APAO or trAPAO polynucleotide into a plant host, including biological  
10 and physical plant transformation protocols. See, for example, Miki et al., (1993), “Procedure for Introducing Foreign DNA into Plants”, In: *Methods in Plant Molecular Biology and Biotechnology*, Glick and Thompson, eds., CRC Press, Inc., Boca Raton, pages 67-88. The methods chosen vary with the host plant, and include chemical transfection methods such as calcium phosphate, microorganism-mediated gene transfer  
15 such as *Agrobacterium* (Horsch, et al., (1985), *Science* 227:1229-31), electroporation, micro-injection, and biolistic bombardment.

Expression cassettes and vectors and *in vitro* culture methods for plant cell or tissue transformation and regeneration of plants are known and available. See, for example, Gruber, et al., (1993), “Vectors for Plant Transformation” In: *Methods in Plant Molecular  
20 Biology and Biotechnology*, Glick and Thompson, eds. CRC Press, Inc., Boca Raton, pages 89-119.

### ***Agrobacterium*-mediated Transformation**

The most widely utilized method for introducing an expression vector into plants is based on the natural transformation system of *Agrobacterium*. *A. tumefaciens* and *A.  
25 rhizogenes* are plant pathogenic soil bacteria, which genetically transform plant cells. The Ti and Ri plasmids of *A. tumefaciens* and *A. rhizogenes*, respectively, carry genes responsible for genetic transformation of plants. See, for example, Kado, (1991), *Crit. Rev. Plant Sci.* 10:1. Descriptions of the *Agrobacterium* vector systems and methods for  
30 *Agrobacterium*-mediated gene transfer are provided in Gruber et al., *supra*; Miki, et al., *supra*; and Moloney et al., (1989), *Plant Cell Reports* 8:238.

Similarly, the gene can be inserted into the T-DNA region of a Ti or Ri plasmid derived from *A. tumefaciens* or *A. rhizogenes*, respectively. Thus, expression cassettes can be constructed as above, using these plasmids. Many control sequences are known which when coupled to a heterologous coding sequence and transformed into a host organism 5 show fidelity in gene expression with respect to tissue/organ specificity of the original coding sequence. See, e.g., Benfey, P. N., and Chua, N. H. (1989) *Science* 244: 174-181. Particularly suitable control sequences for use in these plasmids are promoters for constitutive leaf-specific expression of the gene in the various target plants. Other useful control sequences include a promoter and terminator from the nopaline synthase gene 10 (NOS). The NOS promoter and terminator are present in the plasmid pARC2, available from the American Type Culture Collection and designated ATCC 67238. If such a system is used, the virulence (*vir*) gene from either the Ti or Ri plasmid must also be present, either along with the T-DNA portion, or via a binary system where the *vir* gene is 15 present on a separate vector. Such systems, vectors for use therein, and methods of transforming plant cells are described in US Pat. No. 4,658,082; US application Ser. No. 913,914, filed Oct. 1, 1986, as referenced in US Patent 5,262,306, issued November 16, 1993 to Robeson, et al.; and Simpson, R. B., et al. (1986) *Plant Mol. Biol.* 6: 403-415 (also referenced in the '306 patent); all incorporated by reference in their entirety.

Once constructed, these plasmids can be placed into *A. rhizogenes* or *A. tumefaciens* and these vectors used to transform cells of plant species, which are ordinarily 20 susceptible to *Fusarium* or *Alternaria* infection. Several other transgenic plants are also contemplated by the present invention including but not limited to soybean, corn, sorghum, alfalfa, rice, clover, cabbage, banana, coffee, celery, tobacco, cowpea, cotton, melon and pepper. The selection of either *A. tumefaciens* or *A. rhizogenes* will depend on the plant 25 being transformed thereby. In general *A. tumefaciens* is the preferred organism for transformation. Most dicotyledonous plants, some gymnosperms, and a few monocotyledonous plants (e.g. certain members of the *Liliales* and *Arales*) are susceptible to infection with *A. tumefaciens*. *A. rhizogenes* also has a wide host range, embracing most dicots and some gymnosperms, which includes members of the *Leguminosae*, *Compositae*, 30 and *Chenopodiaceae*. Monocot plants can now be transformed with some success. European Patent Application Publication Number 604 662 A1 to Hiei *et al.* discloses a method for transforming monocots using *Agrobacterium*. Saito *et al.* discloses a method

for transforming monocots with *Agrobacterium* using the scutellum of immature embryos (European Application 672 752 A1). Ishida *et al.* discusses a method for transforming maize by exposing immature embryos to *A. tumefaciens* (Ishida *et al.*, *Nature Biotechnology*, 1996, 14:745-750).

Once transformed, these cells can be used to regenerate transgenic plants, capable of degrading fumonisin. For example, whole plants can be infected with these vectors by wounding the plant and then introducing the vector into the wound site. Any part of the plant can be wounded, including leaves, stems and roots. Alternatively, plant tissue, in the form of an explant, such as cotyledonary tissue or leaf disks, can be inoculated with these vectors, and cultured under conditions, which promote plant regeneration. Roots or shoots transformed by inoculation of plant tissue with *A. rhizogenes* or *A. tumefaciens*, containing the gene coding for the fumonisin degradation enzyme, can be used as a source of plant tissue to regenerate fumonisin-resistant transgenic plants, either via somatic embryogenesis or organogenesis. Examples of such methods for regenerating plant tissue are disclosed in Shahin, E. A. (1985) *Theor. Appl. Genet.* 69:235-240; US Pat. No. 4,658,082; Simpson, R. B., et al. (1986) *Plant Mol. Biol.* 6: 403-415; and U.S. patent applications Ser. Nos. 913,913 and 913,914, both filed Oct. 1, 1986, as referenced in U.S. Patent 5,262,306, issued November 16, 1993 to Robeson, et al.; the entire disclosures therein incorporated herein by reference.

## Direct Gene Transfer

Despite the fact that the host range for *Agrobacterium*-mediated transformation is broad, some major cereal crop species and gymnosperms have generally been recalcitrant to this mode of gene transfer, even though some success has recently been achieved in rice (Hiei *et al.*, (1994), *The Plant Journal* 6:271-282). Several methods of plant transformation, collectively referred to as direct gene transfer, have been developed as an alternative to *Agrobacterium*-mediated transformation.

A generally applicable method of plant transformation is microprojectile-mediated transformation, where DNA is carried on the surface of microprojectiles measuring about 1 to 4  $\mu\text{m}$ . The expression vector is introduced into plant tissues with a biostatic device that accelerates the microprojectiles to speeds of 300 to 600 m/s which is sufficient to penetrate the plant cell walls and membranes. (Sanford *et al.*, (1987), *Part. Sci. Technol.* 5:27;

Sanford, 1988, *Trends Biotech* 6:299; Sanford, (1990), *Physiol. Plant* 79:206; Klein et al., (1992), *Biotechnology* 10:268).

Another method for physical delivery of DNA to plants is sonication of target cells as described in Zang et al., (1991), *BioTechnology* 9:996. Alternatively, liposome or 5 spheroplast fusions have been used to introduce expression vectors into plants. See, for example, Deshayes et al., (1985), *EMBO J.* 4:2731; and Christou et al., (1987), *PNAS USA* 84:3962. Direct uptake of DNA into protoplasts using  $\text{CaCl}_2$  precipitation, polyvinyl alcohol, or poly-L-ornithine has also been reported. See, for example, Hain et al., (1985), *Mol. Gen. Genet.* 199:161; and Draper et al., (1982), *Plant Cell Physiol.* 23:451.

10 Electroporation of protoplasts and whole cells and tissues has also been described. See, for example, Donn et al., (1990), In: *Abstracts of the VIIth Int'l. Congress on Plant Cell and Tissue Culture IAPTC*, A2-38, page 53; D'Halluin et al., (1992), *Plant Cell* 4:1495-1505; and Spencer et al., (1994), *Plant Mol. Biol.* 24:51-61.

15 Thus, polynucleotide encoding a polypeptide able to degrade fumonisin or AP1 can be isolated and cloned in an appropriate vector and inserted into an organism normally sensitive to the *Fusarium* or its toxins. Furthermore, the polynucleotide imparting fumonisin or AP1 degradative activity can be transferred into a suitable plasmid, and transformed into a plant. Thus, a fumonisin or AP1 degrading transgenic plant can be produced. Organisms expressing the polynucleotide can be easily identified by their 20 ability to degrade fumonisin or AP1. The protein capable of degrading fumonisin or AP1 can be isolated and characterized using techniques well known in the art.

### **APAO or trAPAO in a Transgenic Plant**

25 Fumonisin esterase reduces but does not eliminate the toxicity of fumonisins. Therefore a second enzymatic modification to further reduce or abolish toxicity is desirable. The partially purified APAO enzyme from *Exophiala spinifera* has little or no activity on intact FB1, a form of fumonisin. However, recombinant APAO enzyme from *Exophiala spinifera*, expressed in *E. coli*, has significant but reduced activity on intact FB1 and other B-series fumonisins. APAO or trAPAO thus could potentially be used without 30 fumonisin esterase since the amine group is the major target for detoxification. Alternatively, the two genes, fumoninsin esterase and APAO (or trAPAO) can be used together for degrading toxins.

APAO is predicted to be an enzyme that, when by itself or co-expressed in a heterologous expression system along with fumonisin esterase (either ESP1 or BEST1), will result in the production of 2-oxo-FB1 and/or 2-oxo pentol (2-OP) from fumonisin B1. The substrate range of recombinant, *E. coli*-expressed APAO is limited to fumonisins and their hydrolysis products and does not include amino acids, sphingolipid precursors such as phytosphingosine, or polyamines such as spermidine. Thus, APAO is highly specific for fumonisin-like amines, and thus would have little deleterious effect on other cellular metabolites. In addition, if it is extracellularly localized, it will limit any contact with biologically important amines that might also be substrates. The end result will be a more effective detoxification of fumonisins than can be achieved with esterase alone.

The oxidase activity of APAO is predicted to result in generation of hydrogen peroxide in stoichiometric amounts relative to AP1 or fumonisin oxidized. This may prove to be an additional benefit of this enzyme, since hydrogen peroxide is both antimicrobial and is thought to contribute to the onset of a defense response in plants (Przemylaw, *Biochem J.*, 322:681-692 (1997), Lamb, *et al.*, *Ann Rev Plant Physiol Plant Mol Bio* 48:251-275 (1997), and Alvarez, *et al.*, *Oxidative Stress and the Molecular Biology of Antioxidant Defenses*, Cold Spring Harbor Press, 815-839 (1997)).

Because one of the embodiments of the present invention is to have both a fumonisin esterase polynucleotide and an APAO or trAPAO polynucleotide present in a plant, there are several ways to introduce more than one polynucleotide in a plant. One way is to transform plant tissue with polynucleotides to both fumonisin esterase and APAO or trAPAO at the same time. In some tissue culture systems it is possible to transform callus with one polynucleotide and then after establishing a stable culture line containing the first polynucleotide, transform the callus a second time with the second polynucleotide. One could also transform plant tissue with one polynucleotide, regenerate whole plants, then transform the second polynucleotide into plant tissue and regenerate whole plants. The final step would then be to cross a plant containing the first polynucleotide with a plant containing the second polynucleotide and select for progeny containing both polynucleotides.

Another method is to create a fusion protein between esterase and APAO or trAPAO, preferably with a spacer region between the two polypeptides. Both enzymes would be

active although tethered to each other. In addition, an enzyme cleavage site engineered in the spacer region, would allow cleavage by an endogenous or introduced protease.

Transgenic plants containing both a fumonisin esterase enzyme and/or the APAO enzyme and thus able to degrade fumonisin or a structurally related mycotoxin would be  
5 able to reduce or eliminate the pathogenicity of any microorganism that uses fumonisin or a structurally related mycotoxin as a mode of entry to infect a plant. Fungal pathogens frequently use toxins to damage plants and weaken cell integrity in order to gain entry and expand infection in a plant. By preventing the damage induced by a toxin, a plant would be able to prevent the establishment of the pathogen and thereby become tolerant or  
10 resistant to the pathogen.

Another benefit of fumonisin degradation is the production of hydrogen peroxide. When fumonisin or AP1 is oxidatively deaminated at C-2, as occurs by exposure to APAO or trAPAO enzyme, hydrogen peroxide is produced as a by-product. Hydrogen peroxide production can trigger enhanced resistance responses in a number of ways. 1)  
15 Hydrogen peroxide has direct antimicrobial activity. 2) Hydrogen peroxide acts as a substrate for peroxidases associated with lignin polymerization and hence cell wall strengthening. 3) Via still to be determined mechanisms, hydrogen peroxide acts as a signal for activation of expression of defense related genes, including those that result in stimulation of salicylic acid accumulation. Salicylic acid is thought to act an endogenous  
20 signal molecule that triggers expression of genes coding for several classes of pathogenesis-related proteins. Moreover, salicylic acid may set up the oxidative burst and thus act in a feedback loop enhancing its own synthesis. Salicylic acid may also be involved in hypersensitive cell death by acting as an inhibitor of catalase, an enzyme that removes hydrogen peroxide. 4) Hydrogen peroxide may trigger production of additional  
25 defense compounds such as phytoalexins, antimicrobial low molecular weight compounds. For a review on the role of the oxidative burst and SA please see Lamb, C. and Dixon, R.A., *Ann. Rev. Plant Physiol. Plant Mol. Biol.*, 48: 251-275 (1997).

### **Detoxification of Harvested Grain, Silage, or Contaminated Food Crop**

30 The present invention also relates to a method of detoxifying a fumonisin or a structurally related mycotoxin with an APAO enzyme during the processing of grain for animal or human food consumption, during the processing of plant material for silage, or

food crops contaminated with a toxin producing microbe, such as but not limited to, tomato. Since the atmospheric ammoniation of corn has proven to be an ineffective method of detoxification (see B. Fitch Haumann, *INFORM* 6:248-257 (1995)), such a methodology during processing is particularly critical where transgenic detoxification is  
5 not applicable.

In one embodiment of the present invention, fumonisin degradative enzymes are presented to grain, plant material for silage, or a contaminated food crop, or during the processing procedure, at the appropriate stages of the procedure and in amounts effective for detoxification of fumonisins and structurally related mycotoxins. Detoxification by the  
10 enzymes, microbial strains, or an engineered microorganism can occur not only during the processing, but also any time prior or during the feeding of the grain or plant material to an animal or incorporation of the grain or food crop into a human food product, or before or during ingestion of the food crop.

Another embodiment of the present invention is the engineering of a bacterium or  
15 fungus to express the detoxification enzymes and then using the bacterium or fungus rather than the enzyme itself. There are a number of microbes that could be engineered to express the polynucleotides of the present invention. One could also activate, either inducibly or constitutively, the endogenous genes for fumonisin esterase or APAO. By overexpressing the degradative enzymes and then treating plants, seed, or silage with the  
20 microorganism, it would be possible to degrade fumonisin *in situ*.

The polynucleotides of the invention can be introduced into microorganisms that multiply on plants (epiphytes) to deliver enzymes to potential target crops. Epiphytes can be gram-positive or gram-negative bacteria, for example.

The microorganisms that have been genetically altered to contain at least one  
25 degradative polynucleotide and resulting polypeptide may be used for protecting agricultural crops and products. In one aspect of the invention, whole, i.e. unlysed, cells of the transformed organism are treated with reagents that prolong the activity of the enzyme produced in the cell when the cell is applied to the environment of a target plant. A secretion leader may be used in combination with the gene of interest such that the  
30 resulting enzyme is secreted outside the host cell for presentation to the target plant.

The degradative enzymes can be fermented in a bacterial host and the resulting bacteria processed and used as a microbial spray. Any suitable microorganism can be used

for this purpose. See, for example, Gaertner, *et al.* (1993) in *Advanced Engineered Pesticides*, (ed. Kim, Marcel Dekker, New York).

The enzymes or microorganisms can be introduced during processing in appropriate manners, for example as a wash or spray, or in dried or lyophilized form or 5 powered form, depending upon the nature of the milling process and/or the stage of processing at which the enzymatic treatment is carried out. See generally, Hoseney, R.C., *Principles of Cereal Science and Technology*, American Assn. of Cereal Chemists, Inc., 1990 (especially Chapters 5, 6 and 7); Jones, J.M., *Food Safety*, Eagan Press, St. Paul, MN, 1992 (especially Chapters 7 and 9); and Jelen, P., *Introduction to Food Processing*, 10 Restan Publ. Co., Reston, VA, 1985. Processed grain or silage to be used for animal feed can be treated with an effective amount of the enzymes in the form of an inoculant or probiotic additive, for example, or in any form recognized by those skilled in the art for use in animal feed. The enzymes of the present invention are expected to be particularly useful in detoxification during processing and/or in animal feed prior to its use, since the 15 enzymes display relatively broad ranges of pH activity. The esterase from *Exophiala spinifera*, ATCC 74269, showed a range of activity from about pH 3 to about pH 6, and the esterase from the bacterium of ATCC 55552 showed a range of activity from about pH 6 to about pH 9 (US patent no. 5,716,820, *supra*). The APAO enzyme from *Exophiala spinifera* (ATCC 74269) has a pH range of activity from pH 6 to pH 9.

20

### **Genetic Engineering of Ruminant Microorganisms**

Ruminant microorganisms can be genetically engineered to contain and express either the fumonisin esterase enzymes or APAO, or a combination of the enzymes. The 25 genetic engineering of microorganisms is now an art recognized technique, and ruminant microorganisms so engineered can be added to feed in any art recognized manner, for example as a probiotic or inoculant. In addition, microorganisms capable of functioning as bioreactors can be engineered so as to be capable of mass producing either the fumonisin esterases or the APAO enzyme.

## Use of the Fumonisins Esterase and APAO Enzymes for Detection of Reagents for Fumonisins and Related Compounds

Another embodiment of the present invention is the use of the enzymes of the present invention as detection reagents for fumonisins and related compounds. The 5 enzymes of the present invention can be used as detection reagents because of the high specificity of the esterase and deaminase enzymes, and the fact that hydrolysis followed by amine oxidation can be monitored by detection of hydrogen peroxide or ammonia using standard reagents (analogous to a glucose detection assay using glucose oxidase). Hydrogen peroxide is often measured by linking a hydrogen peroxide-dependent 10 peroxidase reaction to a colored or otherwise detectable peroxidase product (e.g. Demmano, *et al.*, *European Journal of Biochemistry* 238(3): 785-789 (1996)). Ammonia can be measured using ion-specific electrodes : Fritzsche, *et al.*, *Analytica Chimica Acta* 244(2): 179-182 (1991); West, *et al.*, *Analytical Chemistry* 64(5): 533-540 (1992), and all herein incorporated by reference) or by GC or other chromatographic method.

15 For example, recombinant or non-recombinant, active fumonisin esterase (ESP1 or BEST) and APAO proteins are added in catalytic amounts to a sample tube containing an unknown amount of fumonisins (FB1, FB2, FB3, FB4, or partial or complete hydrolysis products of these). The tube is incubated under pH and temperature conditions sufficient to convert any fumonisin in the sample to AP1 or to 2-oxo-FB1, and correspondingly the 20 AP1 to 2-OP, ammonia, and hydrogen peroxide. Alternatively, APAO or trAPAO is added in catalytic amounts to a sample tube containing an unknown amount of fumonisins (FB1, FB2, FB3, FB4, or partial or complete hydrolysis products of these). The tube is incubated under pH and temperature conditions sufficient to convert any fumonisin in the sample to 2-oxo FB1, ammonia, and hydrogen peroxide. Then suitable reagents are added for 25 quantification of the hydrogen peroxide or ammonia that were generated stoichiometrically from fumonisins. By comparison with control tubes that received no esterase or APAO enzyme, the amount of fumonisin present can be calculated in direct molar proportion to the hydrogen peroxide or ammonia detected, relative to a standard curve.

This invention can be better understood by reference to the following non-limiting 30 examples. It will be appreciated by those skilled in the art that other embodiments of the invention may be practiced without departing from the spirit and the scope of the invention as herein disclosed and claimed.

## Example 1

**Fungal and bacterial isolates.** *Exophiala* isolates from maize were isolated as described in US patent no. 5,716,820, issued February 10, 1998; US patent no. 6,025,188, 5 issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997, and herein incorporated by reference.

**Isolation methods.** Direct isolation of black yeasts from seed was accomplished by plating 100 microliters of seed wash fluid onto YPD or Sabouraud agar augmented with cycloheximide (500 mg/liter) and chloramphenicol (50 mg/liter). Plates were incubated at 10 room temperature for 7-14 days, and individual pigmented colonies that arose were counted and cultured for analysis of fumonisin-degrading ability as described in US patent no. 5,716,820, issued February 10, 1998; US patent no. 6,025,188, issued February 15, 2000; and pending US application no. 08/888,950, filed July 7, 1997.

**Analysis of fumonisins and metabolism products.** Analytical thin-layer chromatography was carried out on 100% silanized C18 silica plates (Sigma #T-7020; 10 x 15 10 cm; 0.1 mm thick) by a modification of the published method of Rottinghaus (Rottinghaus, *et al.*, *J Vet Diagn Invest*, 4: 326 (1992), and herein incorporated by reference).

To analyze fumonisin esterase activity sample lanes were pre-wet with methanol to 20 facilitate sample application. After application of from 0.1 to 2 µl of aqueous sample, the plates were air-dried and developed in MeOH:4% KCl (3:2) or MeOH:0.2 M KOH (3:2) and then sprayed successively with 0.1 M sodium borate (pH 9.5) and fluorescamine (0.4 mg/ml in acetonitrile). Plates were air-dried and viewed under long wave UV.

For analysis of APAO activity, an alternative method was used. Equal volumes of 25 sample and <sup>14</sup>C-AP1 (1 mg/ml, pH 8, 50 mM sodium phosphate) were incubated at room temperature for one to six days. Analytical thin-layer chromatography was then carried out on C60 HPK silica gel plates (Whatman #4807-700; 10x10 cm; 0.2 mm thick). After application of from 0.1 to 2 µl of aqueous sample, the plates were air-dried and developed in CHCl<sub>3</sub>:MeOH:CH<sub>3</sub>COOH:H<sub>2</sub>O (55:36:8:1). Plates were then air dried, and exposed to 30 PhosphorImager screen (Molecular Dynamics) or autoradiographic film. A Storm™ PhosphorImager (Molecular Dynamics) was used to scan the image produced on the screen.

**Alkaline hydrolysis of FB1 to AP1.** FB1 or crude fumonisin C<sub>8</sub> material was suspended in water at 10-100 mg/ml and added to an equal volume of 4 N NaOH in a screw-cap tube. The tube was sealed and incubated at 60°C for 1 hr. The hydrolysate was cooled to RT and mixed with an equal volume of ethyl acetate, centrifuged at 1000 RCF 5 for 5 minute and the organic (upper) layer recovered. The pooled ethyl acetate layers from two successive extractions were dried under N<sub>2</sub> and resuspended in distilled H<sub>2</sub>O. The resulting material (the aminopentol of FB1 or “AP1”) was analyzed by TLC.

**Enzyme activity of culture filtrate and mycelium.** *Exophiala spinifera* isolate 2141.10 was grown on YPD agar for 1 week, and conidia were harvested, suspended in 10 sterile water, and used at 10<sup>5</sup> conidia per ml to inoculate sterile Fries mineral salts medium containing 1 mg/ml purified FB1 (Sigma Chemical Co.). After 2 weeks incubation at 28°C in the dark, cultures were filtered through 0.45 micron cellulose acetate filters, and rinsed with Fries mineral salts. Fungal mycelium was suspended in 15 mL of 0.1% FB1, pH 5.2 + 1 mM EDTA + 3 µg/mL Pepstatin A + 1.5 µg/mL Leupeptin and disrupted in a 15 Bead Beater™ using 0.1 mm beads and one minute pulses, with ice cooling. Hyphal pieces were collected by filtering through Spin X™ (0.22 µm), and both mycelial supernatant and original culture filtrates were assayed for fumonisin modification by methods outlined above.

**Preparation of crude culture filtrate.** Agar cultures grown as above were used to 20 inoculate YPD broth cultures (500 ml) in conical flasks at a final concentration of 10<sup>5</sup> conidia per ml culture. Cultures were incubated 5 days at 28°C without agitation and mycelia harvested by filtration through 0.45 micron filters under vacuum. The filtrate was discarded and the mycelial mat was washed and resuspended in sterile carbon-free, low mineral salts medium (1 g/liter NH<sub>3</sub>NO<sub>4</sub>; 1 g/liter NaH<sub>2</sub>PO<sub>4</sub>; 0.5 g/liter MgCl<sub>2</sub>; 0.1 g/liter 25 NaCl; 0.13 g/liter CaCl<sub>2</sub>; 0.02 g/liter FeSO<sub>4</sub> · 7H<sub>2</sub>O, pH 4.5) containing 0.5 mg/ml alkaline hydrolyzed crude FB1. After 3-5 days at 28°C in the dark with no agitation the cultures were filtered through low protein binding 0.45 micron filters to recover the culture filtrate. Phenylmethyl sulfonyl fluoride (PMSF) was added to a concentration of 2.5 mM and the culture filtrate was concentrated using an Amicon™ YM10 membrane in a stirred cell at 30 room temperature, and resuspended in 50 mM sodium acetate, pH 5.2 containing 10 mM CaCl<sub>2</sub>. The crude culture filtrate (approx. 200-fold concentrated) was stored at -20°C.

To obtain preparative amounts of enzyme-hydrolyzed fumonisin, 10 mg. of FB1 (Sigma) was dissolved in 20 mL of 50 mM sodium acetate at pH 5.2 + 10 mM CaCl<sub>2</sub>, and 0.25 mL of 200x concentrated crude culture filtrate of 2141.10 was added. The solution was incubated at 37°C for 14 hours, and then cooled to room temperature. The reaction  
5 mixture was brought to approx. pH 9.5 by addition of 0.4 mL of 4 N KOH, and the mixture was extracted twice with 10 mL ethyl acetate. The combined organic layers were dried under N<sub>2</sub> and resuspended in dH<sub>2</sub>O. 2.5 milligrams of organic extracted material were analyzed by Fast Atom Bombardment (FAB) mass spectrometry. The resulting mass spectrum showed a major ion at M/z (+1)=406 mass units, indicating the major product of  
10 enzymatic hydrolysis was AP1, which has a calculated molecular weight of 405.

### Example 2

#### Preparation of AP1-induced and non-induced mycelium.

Liquid cultures of *Exophiala spinifera* isolate 2141.10 were prepared from YPD agar plates (Yeast Extract 10 gm, Bacto-Peptone 20 gm, Dextrose 0.5 gm, and Bacto-Agar 15 gm per liter of water). Aliquots (400-500 uL) of a water suspension of *E. spinifera* cells from YPD agar were spread uniformly onto 150 x 15 mm YPD agar plates with 4 mm sterile glass beads. The plates were incubated at room temperature for 6-7 days. The  
15 mycelia/conidia were transferred from the agar plates into Mineral Salts Medium (MSM) (Na<sub>2</sub>HPO<sub>4</sub>·7H<sub>2</sub>O 0.2 gm, NH<sub>4</sub>Cl 1.0 gm, CaCl<sub>2</sub>·2H<sub>2</sub>O 0.01 gm, FeSO<sub>4</sub>·7H<sub>2</sub>O 0.02 gm per liter of distilled water, pH 4.5) and centrifuged at 5000 x g, 4°C, 20 minutes to pellet the cells. The cell pellet was rinsed once in 40 ml MSM and recentrifuged. The rinsed cell pellet was used to inoculate MSM at a 1:19 ratio of packed cells: MSM. The culture to be  
20 induced was supplemented with AP1 to a final concentration of 0.5-1.0 mg/ml and incubated at 28 °C, 100 rpm, in the dark to induce catabolic enzymes. The non-induced cultures did not receive AP1 but were grown on media containing 4-ABA at the same concentration as AP1. The supernatants were removed by filtration through 0.45 cellulose acetate. The remaining mycelial mat was washed with sterile MSM and then frozen in  
25 liquid nitrogen for storage.  
30

### Example 3

#### Effect of FB1 and AP1 on maize coleoptiles

Maize coleoptiles from 4 day dark-grown germinated maize seeds were excised above the growing point and placed in 96-well microtiter plates in the presence of 60 microliters of sterile distilled water containing FB1 or AP1 at approximately equimolar concentrations of 1.5, .5, .15, .05, .015, .005, .0015, or .0005 millimolar, along with water controls. After 2 days in the dark at 28° C the coleoptiles were placed in the light and incubated another 3 days. Injury or lack thereof was evaluated as follows:

10

	0	.0005	.0015	.005	.015	.05	.15	.5	1.5	mM
FB1	-	-	-	-	+/-	+	+	+	+	
AP1	-	-	-	-	-	-	-	-	+	

+ = brown necrotic discoloration of coleoptile

- = no symptoms (same as water control)

The results (see table above) indicate there is at least a 30-fold difference in toxicity between FB1 and AP1 to maize coleoptiles of this genotype. This is in general agreement with other studies where the toxicity of the two compounds was compared for plant tissues: In *Lemna* tissues, AP1 was approx. 40-fold less toxic (Vesonder *et al.*, "Arch Environ Contam Toxicol 23: 464-467 (1992)"). Studies with both AAL toxin and FB1 in tomato also indicate the hydrolyzed version of the molecule is much less toxic (Gilchrist *et al.*, *Mycopathologia* 117: 57-64 (1992)). Lamprecht *et al.* also observed an approximate 20 100-fold reduction in toxicity to tomato by AP1 versus FB1 (Lamprecht *et al.*, *Phytopathology* 84: 383391 (1994))

### Example 4

#### Effect of FB1 and AP1 on maize tissue cultured cells (Black Mexican Sweet, BMS)

FB1 or AP1 at various concentrations was added to suspensions of BMS cells growing in liquid culture medium in 96-well polystyrene plates. After 1 week the cell density in wells was observed under low power magnification and growth of toxin-treated wells was compared to control wells that received water. Growth of BMS cells was significantly inhibited at 0.4 micromolar FB1, but no inhibition was observed until 40 micromolar AP1. This represents an approximate 100-fold difference in toxicity to maize tissue cultured cells. Similarly Van Asch et al. (VanAsch *et al.*, *Phytopathology* 82: 1330-

1332 (1992)) observed significant inhibition of maize callus grown on solid medium at 1.4 micromolar FB1. AP1 was not tested in that study, however.

### Example 5

5

#### APAO Activity

A cell-free extract that contains the deaminase activity was obtained by subjecting substrate-induced *Exophiala spinifera* cells to disruption using a Bead Beater™ in 50 mM Na-phosphate, pH 8.0, and recovering the cell-free supernatant by centrifugation and .45 micron filtration. Catabolic activity is assayed by incubating extracts with AP1 (hydrolyzed fumonisin B1 backbone) or <sup>14</sup>C-labelled AP1 with the extract and evaluating by TLC on C18 or C60 silica. The product 2-OP has a lower R<sub>f</sub> than AP1 and is detected either by radiolabel scan or by H<sub>2</sub>S<sub>0</sub><sub>4</sub> spray/charring of the TLC plate. 2-OP does not react with the amine reagent, fluorescamine, that is routinely used to detect AP1 on TLC plates, suggesting that the amine group is missing or chemically modified. Activity is greater at 10 37°C than at room temperature, but following 30 min. at 65°C or 100°C (no AP1 catabolic activity remained). Activity is maximal at pH 9. At pH 9, complete conversion to 2-OP occurred in 30 minutes. Activity is retained by 30,000 dalton molecular weight cutoff membrane, but only partially retained by 100,000 dalton molecular weight cutoff membrane. Other amine-containing substrates were tested for modification by the crude 15 extract. Fumonisin, with tricarballylic acids attached, is not modified by the extract, indicating that ester-hydrolysis must occur first for the APAO to be able to be effective in modifying FB1 (as noted below, the *E. coli*-expressed, recombinant APAO enzyme does in fact oxidize FB1 although at a lower rate than AP1). Other long-chain bases (sphingosine, sphinganine, and phytosphingosine) are apparently not modified by the crude APAO, 20 suggesting the enzyme(s) is specific for the fumonisin backbone. Preparative amounts of the product, named 2-OP, have also been purified and analyzed by C13 nmr. The results indicate that 2-OP has a keto group at carbon 2 instead of an amine, consistent with an oxidative deamination by an amine oxidase. The C13 nmr data also indicate that 2-OP spontaneously forms an internal hemiketal between C-1 and C-5, resulting in a 5- 25 membered ring with a new chiral center at C-2. All other carbon assignments are as in AP1, thus 2-OP is a compound of composition C<sub>22</sub>H<sub>44</sub>O<sub>6</sub>, FW 404. The product of the 30

enzyme acting on hydrolyzed fumonisins would not be expected to display any significant toxicity.

Other enzymes were tested for their ability to modify AP1. All enzymes were assayed by radiolabeled TLC, as described above, under optimal conditions at 37° Celsius, 5 overnight or longer. The results are as follows:

Deaminating	EC	Source	Result
Monoamine Oxidase	1.4.3.4	bovine plasma	negative
D-amino oxidase	1.4.3.3	porcine kidney; TypeX	negative
L-amino oxidase	1.4.3.2	C.adamanteus venom; TypeI	negative
Tyramine oxidase	1.4.3.4	Arthrobacter spp	negative
Methylamine dehydrogenase	1.4.99.3	Paracoccus denitrificans	negative
Aralkyl amine dehydrogenase	1.4.99.4	Alcaligenes faecalis	negative
Phenylalanine ammonia lyase	4.3.1.5	Rhodotorula glutinis; TypeI	negative
Histidine ammonia lyase	4.3.1.3	Pseudomonas fluorescens	negative
L-aspartase	4.3.1.1	Hafnia alvei (Bacterium cadaveris)	negative
Tyrosine oxidase	1.14.18.1	mushroom	negative
Lysine oxidase	1.4.3.14	Trichoderma viride	negative
Diamine oxidase	1.4.3.6	porcine kidney	negative

The results were negative for each enzyme tested. Therefore isolates from the American Type Culture Collection (ATCC) were collected. The ATCC isolates selected 10 were listed as containing amine-modifying enzymes or were capable of growth/utilization on amine-containing substrates. The isolates were tested to determine if they could grow on or utilize AP1 as the sole carbon source and if any could modify AP1 to a new compound(s). The nitrogen sources that were used in liquid cultures were AP1 0.1% (w/v), s-butylamine 0.1% (v/v), n-butylamine 0.1% (v/v), and ammonium nitrate 0.2% 15 (w/v). These were prepared in Vogel's Minimal Media (without NH<sub>4</sub>NO<sub>3</sub>) containing 2% sucrose. The isolates were inoculated into the various media and monitored for growth over 2-3 weeks. They were also assayed with the <sup>14</sup>C-radiolabeled TLC assay for AP1 modification. In summary, none of the isolates tested exhibited modification of AP1 *in vivo*. Clearly the APAO enzyme is unique and unusual in its ability to modify the AP1 20 toxin.

## EXAMPLE 6

### Isolation of the trAPAO Polynucleotide

The trAPAO polynucleotide was identified using a proprietary transcript imaging 25 method that compares transcript patterns in two samples and allows cloning of differentially expressed fragments. This technology was developed by CuraGen® (New

Haven, CT). (see Published PCT patent application no. WO 97/15690, published May 1, 1997, and hereby incorporated by reference) Fluorescently-tagged, PCR amplified cDNA fragments representing expressed transcripts can be visualized as bands or peaks on a gel tracing, and the cDNA from differentially expressed (induced or suppressed) bands can be 5 recovered from a duplicate gel, cloned and sequenced. Known cDNAs can be identified without the need for cloning, by matching the predicted size and partially known sequence of specific bands on the tracing.

In the present invention two RNA samples were obtained from cultures of *E. spinifera* grown for a specified period in a mineral salts medium containing either AP1 (induced 10 condition), or gamma-aminobutyric acid (ABA; non-induced condition) as a sole carbon source. In the induced condition, fumonisin esterase and APAO enzyme activities are detected, whereas in the non-induced condition these activities are not detected. The methods used for induction of APAO and detection of activity are described earlier (see Example 2 and Example 5). RNA was extracted from induced mycelium by Tri-Reagent 15 methods (Molecular Research Center Inc., Cincinnati, Ohio) only grinding a frozen slurry of tissue and Tri-Reagent with a mortar and pestle until almost melted and adding an additional extraction after the phase separation by extracting the aqueous phase one time with phenol, and two times with a phenol:chloroform:isoamyl alcohol mixture. The RNA's were submitted for CuraGen® transcript imaging to detect cDNA fragments that 20 are induced specifically in the presence AP1. In the resulting gel tracing several bands were found which showed induction of at least 2-fold and up to 79-fold or even 100-fold or more in AP1. In the resulting gel tracing several bands were found which showed induction of at least 10-fold in AP1-grown cells as compared to cells grown in ABA. The sequence of two highly induced bands can be found in Table 1.

25

**TABLE 1**

**Nucleotide sequence of two CuraGen® bands that were identified as strongly induced by AP1 in cultures of *Exophiala spinifera*.**

30 >k0n0-395.5\_b (SEQ ID NO: 1)  
GGGCCCGGCGTTCTCGTAGGCTGCGCGGAGTTGGTCCCAGACAGACTTTGTCGTACCTGCTTG  
GACTGTTGGGACCACCTCCGTCGGGTCTCCGACCATGAAACAGGTAAATGGACCATGTCGAT  
CGACGTCGATGCTGGTATCTCTGGCAAATGAGATGGGGTCACAGCTCGATTGGAGGACGCCGA  
GAAGCCTTGTTCGCGCCACCAACGGCTTGTCCCATAACGAAGACTATCTTGCTATAGTAGCCCAGG  
35 ATAGAATTTCGCCAATGCTGCTTCTCGGCGGGAAAGAGGTGGTAAAATGTCAAGGTGGGAT  
ACAAGGTTGTCGGTAACGAAACCANCACCTTTGCTTCGGAACACGGCGC

>r0c0-182.3\_6 (SEQ ID NO: 2)  
GAATTTCCGCCAATGCTGCTTCGGCGGGAAAGAGGTGGTAAAAATGTCAAGGTGGATACA  
AGGTTGTCGGTAACGAAACCACCACTTTGCTCGAACACCGCGCCGAGGCCATCGTAC  
TGTACAGCCGGATGCCGACTGCTCAATTCAAGCGACGGGGTGGTGAAGGTGCAC

5

Two of the highly induced bands, k0n0-395.5, and r0c0-182.3 showed significant sequence homology to a family of enzymes, flavin-containing amine oxidases (EC 1.4.3.4), that oxidize primary amines to an aldehyde or ketone, releasing ammonia and 10 hydrogen peroxide (Table 2).

TABLE 2

15 Identification of a putative flavin amine oxidase from *E. spinifera*: AP1-induced transcript fragments with amine oxidase homology. BLAST 2.0 default parameters.

Clone ID	Size	Best Hit	Best Hit Name, source	Prob	from	to	Likely function
k0n0-395.5	395 bp	P40974	putrescine oxidase, <i>Micrococcus rubens</i> , EC 1.4.3.10 Length = 478	8.0 e -07	276	333	oxidation of C-2 amine of AP1
r0c0-182.3 (contigs with k0n0-395)	182 bp	P12398	monoamine oxidase type A (MAO-A) [ <i>Bos taurus</i> ] Length = 527	0.0039	238	296	oxidation of C-2 amine of AP1

20 The chemical structure of the primary product of AP1 deamination is thought to be a 2-keto compound which cyclizes to a hemiketal at carbons 2 and 5. Therefore it is predicted that this induced enzyme is responsible for deamination of AP1.

Using sequence derived from k0n0-395.5, a partial cDNA was obtained by 3' and 5' RACE-PCR (Chenchik, *et al.*, CLONTECHniques X 1:5-8 (1995); Chenchik, *et al.*, A new 25 method for full-length cDNA cloning by PCR. In *A Laboratory Guide to RNA: Isolation, Analysis, and Synthesis*. Ed. Krieg, P.A. (Wiley-Liss, Inc.), 273-321 (1996)). A RACE cloning kit from CLONTECH was used, to obtain the RACE amplicons. Briefly, poly A+ RNA is transcribed to make first strand cDNA using a "lock-docking" poly T, cDNA synthesis primer, the second strand is synthesized and the

Marathon cDNA adaptor is ligated to both ends of the ds cDNA. Diluted template is then used with the Marathon adapter primer and in separate reactions either a 5' Gene Specific Primer (GSP) or a 3'GSP is used to produce the 3' or 5' RACE amplicon. After characterization of the RACE product(s) and sequencing, full-length cDNAs may be generated by 1) end-to-end PCR using distal 5' and 3' GSPs with the adapter-ligated ds cDNA as template, or 2) the cloned 5' and 3'-RACE fragments may be digested with a restriction enzyme that cuts uniquely in the region of overlap, the fragments isolated and ligated. Subsequently, the RACE-generated full-length cDNAs from 1) and 2) may be cloned into a suitable vector.

In combination with the supplied adapter primer the following gene specific primers were used: for 3' RACE the oligonucleotide N21965: 5'-TGGTTTCGTTACCGACAACCTTGTATCCC-3' (SEQ ID NO: 3) and for 5' race, the oligonucleotide N21968: 5'-GAGTTGGTCCCAGACAGACTTTGTCGT-3' (SEQ ID NO: 4. The polynucleotide sequence of the trAPAO polynucleotide, k0n0-395\_6.5, from *Exophiala spinifera* is shown in SEQ ID NO: 5. The polypeptide sequence of trAPAO is shown in SEQ ID NO: 6.

A second clone of APAO containing an unspliced intron was also found. The polynucleotide sequence of trAPAO-I polynucleotide, k0n0-395\_5.4, the intron containing clone, from *Exophiala spinifera*, can be found in SEQ ID NO: 7. The polypeptide sequence of trAPAO-I with the intron spliced out is shown in SEQ ID NO: 8. The polypeptide sequence of trAPAO-I without the intron spliced out is shown in SEQ ID NO: 9.

## EXAMPLE 7

### Heterologous Expression of trAPAO

Protein alignments generated with PileUp (GCG) indicate that k0n0-395\_6.5 (trAPAO) is similar in size to other flavin amine oxidases and is close to being full length with respect to the amino terminus of their class of proteins. The k0n0-395\_6.5 sequence contains a complete  $\beta$ - $\alpha$ - $\beta$  fold that is required for dinucleotide (FAD) binding, close to the amino end. The k0n0-395 sequence appears to lack only a variable amino terminal segment that varies in length from 5 amino acids in rat monoamine oxidases A & B to 40 amino acids in length in *Aspergillus* MAO-N. The function of these amino terminal extensions is not known; they are not recognizable as secretion signals. Based on the likely

localization of the *Exophiala* APAO outside the cell membrane, the prediction is that k0n0-395 would have a signal sequence similar to that of the fumonisin esterase cloned from the same organism (US patent no. 5,716,820, *supra*). Using GenomeWalker™, it is possible to clone the 5' end of the transcript and upstream genomic regulatory elements.

5 However, the signal sequence is not expected to be critical to the functionality of the enzyme; in fact, the preferred strategy for heterologous expression in maize and *Pichia pastoris* involves replacing the endogenous signal sequence (if present) with an optimized signal sequence for the organism, e.g. barley alpha amylase for maize and the yeast alpha factor secretion signal for *Pichia*. In maize transformed with fumonisin esterase, the

10 barley alpha amylase signal sequence gave higher amounts of functional protein than the native fungal signal, therefore replacement of the native fungal signal sequence is a logical optimization step. Since many of the amine oxidases have a positively charged amino acid near the N-terminus and upstream of the dinucleotide binding site, an additional optimization step included adding a codon for the lysine (K) to the N-terminus of the

15 trAPAO clone (k0n0-395\_6.5, SEQ ID NO: 5). This clone is designated K:trAPAO and can be seen in SEQ ID NOS: 10 and 11. The extra lysine is at amino acid 1 and nucleotides 1-3.

#### EXAMPLE 8

##### 20 *Pichia* Expression of trAPAO

For optimum expression of trAPAO in *Pichia pastoris* the alpha mating factor signal peptide was operably linked in-frame with K:trAPAO coding sequence and can be seen in SEQ ID NOS: 16 and 17. The nucleotide sequence of clone pPicZalphaA:K:trAPAO contains a PCR-amplified insert comprising the k0n0-395 open reading frame with an additional lysine residue at the amino terminus, with a 5' EcoRI site and 3' NotI site for in-frame cloning into the alpha factor secretion vector pPicZalphaA. Nucleotides 1-267 contain the yeast α mating factor secretion signal. The amino acid sequence, shown in SEQ ID NO: 17, contains the trAPAO polypeptide produced from pPicZalphaA:K:trAPAO following transformation into *Pichia pastoris*.

30 For cloning into expression vectors, two cloning strategies were used. The cDNA k0n0-395\_5.4 was generated by using end-to-end PCR using distal 5' and 3' GSPs with the adapter-ligated double stranded cDNA as a template. Each oligonucleotide primer was

designed with 5' restriction enzyme sites that contain a 23-25 bp of anchored gene sequence. The 3' primer also included the stop codon. The primer sequences are N23256: 5'-ggggaattcAAAGACAACGTTGCGGACGTGGTAG-3' (SEQ ID NO: 12) and N23259: 5'-ggggcgccgcCTATGCTGCTGGCACCAAGGCTAG-3' (SEQ ID NO: 13). A second  
5 method was used to generate k0n0-395\_6.5. 5' RACE and 3' RACE products using a distal primer containing the necessary restriction enzyme sites, stop codon, etc as described above and paired with a "medial" GSP. The "medial primers" N21965: 5'-TGGTTTCGTTACCGACAACCTTGTATCCC-3' (SEQ ID NO: 14) for 3' RACE and for  
10 5' race, the oligonucleotide N21968: 5'-GAGTTGGTCCCAGACAGACTTTGTCGT-3'  
15 (SEQ ID NO: 15). Adapter-ligated double stranded cDNA was used as template. The isolated 5' and 3'-RACE fragments were digested with a restriction enzyme that cuts uniquely in the region of overlap, in this case Bgl I, isolated and ligated into the expression vector. The digestible restriction sites allow cloning of the inserts in-frame into EcoRI/NotI digested pPicZalphaA. pPicZalphaA is an *E. coli* compatible *Pichia* expression vector containing a functional yeast alpha factor secretion signal and peptide processing sites, allowing high efficiency, inducible secretion into the culture medium of *Pichia*. The resulting 1.4 kb bands were cloned into EcoRI/NotI digested pPicZalphaA plasmid.

SEQ ID NO: 16 contains the polynucleotide sequence of clone pPicZalphaA:K:trAPAO, a PCR-amplified insert that comprises the k0n0-395 open  
20 reading frame with an additional lysine residue at the amino terminus, and a 5' EcoRI site and 3' NotI site for in-frame cloning into the alpha factor secretion vector pPicZalphaA. SEQ ID NO: 17 contains the amino acid sequence of the trAPAO polypeptide produced from pPicZalphaA:K:trAPAO following transformation into *Pichia pastoris*. The alpha factor secretion signal and a lysine are added.

25 *Pichia* was transformed as described in Invitrogen Manual, Easy Select<sup>TM</sup> *Pichia* Expression Kit, Version B, #161219, with the trAPAO polynucleotide as described above with either an intron (trAPAO-I, negative control, no expression of active trAPAO since *Pichia* does not splice introns very efficiently) or without an intron (capable of making an active APAO protein). The *Pichia* culture fluids and pellets were assayed for APAO  
30 activity as described earlier.

The set of frozen six day *Pichia* culture cell pellets contained two samples with intron (SEQ ID NO: 7) in gene construct, # 11, # 14, and two samples without intron in

gene construct (SEQ ID NO: 5), #6, # 52. The six day culture fluids from the same cultures were used to spike with crude fungal enzyme for positive controls.

The 50  $\mu$ l cell pellets were resuspended in 150  $\mu$ l cold 50mM Na-phosphate, pH 8.0, and divided into two fresh 500  $\mu$ l tubes. One tube was kept on ice with no treatment, 5 the pellet suspension, and one tube was used for lysis. An equal volume of 0.1 mm zirconia-silica beads was added to each tube. The tubes were BeadBeat<sup>TM</sup> for 15 seconds then cooled on ice 5 minutes. This was repeated three times. The crude lysate was then transferred to another tube for assay or lysate suspension.

The TLC assays were performed as follows, the samples are 1) pellet suspensions; 10 10  $\mu$ l; 2) lysate suspensions; 10  $\mu$ l; 3) media controls-mixed 5  $\mu$ l media with 5  $\mu$ l crude fungal enzyme; 10  $\mu$ l; 4) positive control-used crude fungal enzyme undiluted; 10  $\mu$ l; 5) substrate control-used 50mM Na-phosphate, pH8.0; 10  $\mu$ l. Ten microliters of each sample plus 10  $\mu$ l of <sup>14</sup>C-AP1 (1 mg/ml, 50 mM Na-phosphate, pH 8) was incubated at room temperature for 6 days. One microliter of the sample was spotted onto C18 and C60 TLC 15 plates. The C18 plates were developed in MeOH:4% KCl (3:2). The C60 plates were developed in CHCl<sub>3</sub>:MeOH:CH<sub>3</sub>COOH:H<sub>2</sub>O (55:36:8:1). The plates were then air dried and then exposed to a PhosphorScreen<sup>TM</sup> for 2-3 days. A Storm<sup>TM</sup> PhosphorImager was used to develop the images.

A positive TLC result is obtained if an additional radioactive spot appears at a 20 lower Rf of the produced AP1 modification earlier identified as 2-OP, a deaminated product of AP1. In samples # 6 and # 52 (without intron) the AP1-modifying enzyme activity (conversion of AP1 to 2-OP) was detected in pellet suspensions and pellet lysates, although the majority of activity was associated with the pellet suspensions. In samples #11 and #14 (with intron) a minimal amount of AP1-modifying enzyme activity was 25 detectable in the pellet lysate of # 14 only, which indicates Pichia cannot process the intron efficiently.

This experiment verified APAO activity can be detected in *Pichia* transformants, which verifies that trAPAO as described functions correctly in degrading AP1. The activity is associated with cell suspensions, which show higher activity than pellet lysates. 30 Pellet lysates may show less activity due to release of endogenous proteases during lysis of the cells.

## EXAMPLE 9

### **Expression of trAPAO or APAO in *E. coli***

The vector for expressing K:trAPAO in *E. coli* is pGEX-4T-1. This vector is a prokaryotic glutathione S-transferase (GST) fusion vector for inducible, high-level intracellular expression of genes or gene fragments as fusions with *Schistosoma japonicum* GST. GST gene fusion vectors include the following features, a lac promoter for inducible, high-level expression; an internal lac Iq gene for use in any *E. coli* host; and the thrombin factor Xa or PreScission Protease recognition sites for cleaving the desired protein from the fusion product. The insert of interest, k0n0-395\_6.5 (K:trAPAO) or APAO, was subcloned into the 5' EcoRI site and a 3' NotI site allowing in-frame expression of the GST:K:trAPAO or GST:APAO fusion peptide.

The polynucleotide sequence of the GST:K:trAPAO fusion can be found in SEQ ID NO: 18. The GST fusion with polylinker can be found at nucleotides 1 to 687. The K:trAPAO can be found at nucleotides 688 to 2076. The resulting polypeptide for the GST:K:trAPAO fusion can be seen at SEQ ID NO: 19. Amino acids 1 to 229 represent the GST fusion plus polylinker and amino acids 230 to 692 represent the K:trAPAO portion of the fusion.

*E. coli* was transformed with the pGEX-4T-1 vector containing K:trAPAO or APAO as described in BRL catalogue, Life Technologies, Inc. catalogue; Hanahan, D., *J. Mol. Biol.* 166:557 (1983) Jessee, J. *Focus* 6:4 (1984); King, P.V. and Blakesley, R., *Focus* 8:1, 1 (1986), and hereby incorporated by reference. The transformed *E. coli* was induced by addition of IPTG (isopropyl b-D-thiogalactopyranoside). Four samples of soluble extract and four samples of insoluble inclusion bodies were tested for trAPAO or APAO activity as described in Example 8. APAO activity was present in all soluble samples and two insoluble samples. Highest activity was found at 10 uM IPTG induction. Thus the pGEX-4T-1 vector containing K:trAPAO or APAO is capable of producing active APAO enzyme in *E. coli*.

## EXAMPLE 10

30

### **The Complete Nucleotide Sequence of the *Exophiala* APAO Gene**

Using Genome Walker, the complete nucleotide sequence of the *Exophiala* APAO gene was recovered. The nucleotide sequence described in SEQ ID NO: 5 is missing a

portion of the 5' end of the native gene. The missing portion of the 5' end of the native gene is not necessary for expression of an active APAO enzyme, as can be seen in Examples 8 and 9. The complete nucleotide sequence of APAO can be seen in SEQ ID NO: 22. The translation of SEQ ID NO: 22 can be found in SEQ ID NO: 23.

5

## EXAMPLE 11

### Expression of APAO and ESP1 in transgenic maize callus

One of the preferred constructs for expression in maize is the nucleotide sequence of the trAPAO operably linked to the barley alpha amylase signal sequence. The nucleotide sequence of K:trAPAO translational fusion with barley alpha amylase signal sequence, for expression and secretion of the mature trAPAO in maize can be seen in SEQ ID NO: 20. Nucleotides 1-72, represent the barley alpha amylase signal sequence; nucleotides 73-75, represent the added lysine residue; and nucleotides 76 -1464 , represent the trAPAO cDNA. The amino acid sequence translation of SEQ ID NO: 20 can be found in SEQ ID NO: 21. Amino acids 1 to 24 represent the barley alpha amylase signal sequence and amino acids 25 to 463 is the sequence of K:trAPAO.

Maize embryos were transformed with linear DNA (insert, lacking a bacterial antibiotic resistance marker), derived from constructs containing three transcription units: 1) a PAT selectable marker gene (Wohlleben *et al.*, *Gene* 70, 25-37 (1988)), 2) fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence, and 3) full length APAO without or with an amino-terminal barley alpha amylase signal sequence, (P13603, comprising a PAT selectable marker operably linked to a 35S promoter, fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence and the ubiquitin promoter, and APAO operably linked to the ubiquitin promoter and P13611, comprising a 25 PAT selectable marker operably linked to the 35S promoter, fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence and the ubiquitin promoter and APAO operably linked to a barley alpha amylase signal sequence and the ubiquitin promoter). In these constructs both ESP1 and APAO were linked to the maize ubiquitin promoter and first intron. In a third construct, the same three transcriptional units were 30 cloned into an Agrobacterium T1 vector (P15258, the construct comprises a PAT selectable marker, fumonisin esterase ESP1 operably linked to a barley alpha amylase signal sequence and APAO). Stably transformed callus or T0 plants regenerated from callus were tested for ESP1 and APAO activity in buffer extracts of leaf tissue, using

radiolabeled FB1 and/or AP1 and C18 thin-layer chromatography. Positive controls consist of non-transformed tissue spiked with *E. coli*-expressed recombinant ESP1 or APAO. The results indicate that both ESP1 and APAO activities can be detected in transgenic maize callus and plants.

5

**Expression of ESP1 and APAO in transgenic callus**

Construct	Sample ID Number	ESP1 activity (TLC)	APAO activity (TLC)
13603	3065.031-2	+	+
13603	3065.034-3	+	+
13603	3065.1117-3	+	+
13603	3065.11s7-n13	+	+
13603	3065.117-2	+	+
13603	3065.1115-2	+	+
13603	3065.1115-6	+	+
13603	3065.1112-1	+	+
13603	3065.118-6	+	+
13603	3065.11s3-1	+	+
13603	3065.11s1-13	+	+
13603	2805.762-2	+	+
13603	3065.1110-2	+	+
13603	3065.039-2	+	+
13611	3065.293-3	+	+
13611	3065.263-1	+	+
13611	3070.24.2.3	+	+

Transgenic plants were regenerated from the transgenic callus positive for both  
10 ESP1 and APAO activity by standard methods known in the art. Enzyme activity was  
tested as described previously. As can be seen below transgenic maize plants can  
successfully express both ESP1 and APAO enzymes.

15

**Expression of APAO and ESP1 in transgenic maize plants (T0)**

Construct	Sample ID Number	ESP1 activity (TLC)	APAO activity (TLC)
13603	910080	+	+
13603	910081	+	+
13603	917065	+	+

Another preferred construct for expression of APAO in a plant is targeting the  
20 APAO to the peroxisome. Maize embryos were bombarded with insert containing APAO

operably linked to ubiquitin promoter and a peroxisomal targeting sequence (Gould, *et al.*, *J Cell Biol* 108:1657-1664 (1989)); ESP1 operably linked to ubiquitin promoter and the barley alpha amylase signal sequence; and a selectable marker of PAT operably linked to the 35S promoter (construct number I14952). Negative controls were unbombarded embryos/callus. Positive controls were unbombarded embryos/callus spiked with purified enzyme. Transformed callus was then tested for ESP1 or APAO activity as previously described. Out of 67 samples tested 18 samples contained both ESP1 activity and APAO activity. Peroxisomally targeted APAO and apoplast targeted fumonisin esterase can both be successfully expressed in a plant cell.

Another preferred construct for expression of APAO in a plant is targeting the APAO to the mitochondrial membrane. A C-terminal extension is required for targeting monoamine oxidases MAO-A and MAO-B to mammalian outer mitochondrial membranes. A MAO-A, MAO-B, or functionally similar C-terminal extension can be ligated in-frame to APAO or trAPAO to facilitate localization of this enzyme to the mitochondrial membrane of maize or other transformed species.

## EXAMPLE 12

### Comparison of APAO Sequence With Other Sequences

The *Exophiala* cDNA APAO (SEQ ID NO: 22) contains an 1800 bp open reading frame coding for a 600 amino acid polypeptide (SEQ ID NO: 23) with divergent homology to two classes of proteins. The carboxy three-fourths of APAO (amino acids 137 to 593) is strongly homologous to flavin amine oxidases, a group of enzymes catalyzing the oxidative deamination of primary amines at carbon 1. The amine oxidase function of the carboxy terminal domain was confirmed by expression of a truncated APAO polypeptide (from 137 to 600) in both *Pichia pastoris* and *E. coli*, using AP1 as a substrate (see Example 9). The amino terminal portion of APAO, in contrast, (from approx. 5 to 134) shows significant homology to a group of small deduced open reading frames (ORFs) reported in several bacteria and blue-green algae, as well as several higher organisms. These ORFs code for small proteins of unknown function, ranging in size from 14 to 17 kDa. The juxtaposition of these divergent homologies in a single polypeptide has not been reported previously.

Flavin amine oxidases (E.C. 4.1.4.3) are a group of flavoenzymes found in both higher and lower organisms, and serve a variety of functions in catabolism. They catalyze the oxidative deamination of primary amino groups located at the C-1 position of a variety of substrates, resulting in an aldehyde product plus ammonia and hydrogen peroxide. The 5 APAO enzymes of the present invention are the first flavin amine oxidase known to attack a primary amine not located at C-1 (i.e. C-2 of AP1) and resulting in a keto rather than aldehydic product. However, amino acid oxidases, while not closely related to flavin amine oxidases, are flavoenzymes that oxidize a C-2 amine adjacent to a C-1 carboxyl group.

10 The monoamine oxidases MAO A & B, (from human, bovine, and trout), are localized in the mitochondrial outer membrane of higher organisms and regulate the level of neurotransmitters. Microbial examples include a fungal amine oxidase (*Aspergillus niger* (*niger*) MAO-N) involved in amine catabolism, and a bacterial putrescine oxidase from a gram (+) bacterium (*Micrococcus rubens*.). The primary polypeptides vary in 15 length from 478 to 527 amino acids, and share regions of high amino acid sequence conservation at the 5' end as well as at various points through the coding region. Protein alignments generated with PileUp (GCG) indicate that trAPAO contains all conserved domains found in this class of proteins including those near the 5' end.

20 The amine oxidase domain of trAPAO contains several key features shared by this class of enzymes, including an amino-terminal dinucleotide (ADP) binding region characterized by a beta-alpha-beta stretch containing three invariant glycines (G -X-G-X-X-G) in the beta-alpha turn. In trAPAO, this sequence is (DVVVVGAGLSG). This region is involved in FAD binding. Absent are several features unique to the mammalian 25 amine oxidases, including several important cysteine residues (Wu *et al.*, *Mol Pharm* 43:888 (1993)), one of which (Cys-406 of MAO-A) is involved in covalent binding of FAD, and a carboxy-terminal extension that has been demonstrated to be involved in transporting to and anchoring the MAO in the outer mitochondrial membrane. The *Aspergillus* enzyme MAO-N has been demonstrated to contain non-covalent FAD, and also lacks the conserved cysteine. Therefore it is possible that the APAO enzyme has a 30 non-covalent FAD. The *Aspergillus* MAO-N has a carboxy-terminal tripeptide Ala-Arg-Leu that is involved in peroxisomal targeting and localization; this sequence is absent from *Exophiala* MAO.

The amine oxidase domain of trAPAO contains a total of seven cysteines, compared to ten for the *Aspergillus* enzyme and only two for the *Micrococcus* enzyme. The mammalian MAO enzymes contain variable numbers of cysteines (at least ten), some of which are highly conserved (including the FAD binding residue mentioned above). The 5 trAPAO sequence also has two putative glycosylation sites (NDS, NQS) towards the amino end.

The purpose of the amino-terminal extension of APAO and the basis for its homology to a group of 14-17 kDa proteins is not clear. In *Synechocystis*, a similar polypeptide ORF is located immediately upstream of the NADP-dependent glutamine 10 dehydrogenase (gdhA) and has been shown to be required for functional expression of gdhA (Chavez et al, 1995). However, in trAPAO the domain is clearly not necessary for enzymatic activity, as shown by the results of the expression experiments using the truncated APAO. An interesting clue comes from the frequent association of this small 15 ORF with gene clusters involved in oxidoreductase activity in bacteria, or induced by heat stress in mice, suggesting a possible role in redox protection. A byproduct of amine oxidase activity is hydrogen peroxide. Flavoenzymes and other redox enzymes are often susceptible to inactivation by hydrogen peroxide (Schrader et al., *App Microb Biotechnol* 45:458; Aguiree, et al., *J Bacteriol* 171:6243 (1989)), and it is possible that this protein has a protective role against oxidants such as hydrogen peroxide. Alternatively, this domain 20 could be involved in enzyme function, localization or association of the enzyme with other structures. No signal peptide region can be detected in this amino terminal region.

In multiple sequence alignment using GCG PileUp, trAPAO is most similar to putrescine oxidase of *Micrococcus rubens*, Swissprot accession number P40974, (30% identical amino acids, 40% similar). Homology with several mammalian monoamine 25 oxidases A and B, Swissprot accession numbers P21397 (*Homo Sapiens* mao a), P19643 (*Rattus norvegicus* mao b), P21396 (*Rattus norvegicus* mao a), and P21398 (*Bos taurus* mao a), is somewhat less, ranging from 25 to 28% identity and 36 to 40% similarity. Homology to the only other fungal flavin amine oxidase known, MAO-N from *Aspergillus niger* (Swissprot accession number P46882), is somewhat lower (24% identical, 34% 30 similar). The microbial enzymes are considerably divergent from each other, while the mammalian monoamine oxidases share 65 to 87% identity.

The amino terminal domain (ATD) of APAO also shows homology to a 14.5 kD protein from human and rat phagocytes that shows translational inhibition activity *in vitro* (Swissprot accession # P52758, P52759) Schmiedeknecht, *et al.*, *Eur J Biochem* 242 (2), 339-351 (1996), and includes a heat-responsive protein from mouse (Samuel, *et al.*, 5 *Hepatology* 25 (5), 1213-1222 (1997)). This suggests that this family of proteins is involved in regulating cellular metabolism. No example exists in which this domain is fused to a larger protein domain, however, making APAO unique. Without intending to be limited by theory, all of this suggests, that this domain plays a regulatory role in APAO gene expression, possibly to prevent translation of the message when it is not needed. This 10 raises the question of how translation of the message is restored when active enzyme is required by the *Exophiala* cell. Possibly there are alternative start sites that begin downstream of the inhibitor domain; or proteolysis, complexing, degradation, or phosphorylation/ dephosphorylation of the inhibitor domain when it is not needed. The first possibility is less likely because there are no other ATG codons prior to the ATG at 15 122-124 that constitutes the predicted start site of APAO. The second possibility cannot be easily tested, although there is a casein kinase site in the ATD. Alternative roles for the ATD include oligomerization of the APAO protein, or anchoring the protein to some intracellular site, such as the membrane.

A parallel example of regulatory control over another flavoenzyme, human flavin 20 monooxygenase 4 (FMO-4), by a C-terminal extention has been reported (Itagaki, *et al.*, *J of Biol Chem* 271(33): 20102-20107 (1996)). In this case the introduction of a stop codon prior to the 81 base C-terminal extension allowed expression of active enzyme in heterologous systems. The role of the C-terminal portion was not elucidated, however. In another example, alternative splicing led to a shorter gene product that complexed with and 25 interfered with the function of the normally spliced version (Quinet, *et al.*, *J of Biol Chem* 268(23): 16891-16894 (1993)). In another case, an alternative splicing-generated insert in another protein led to inhibition of cell growth (Bhat, *et al.*, *Protein Engineering* 9(8): 713-718 (1996)). In yet another variation, fas/Apo1 splicing variants prevent apoptosis, apparently through a 49 amino acid domain shared by all variants ((Papoff, *et al.*, *J of 30 Immunology* 156(12): 4622-4630 (1996)).

### EXAMPLE 13

#### Making a Chimera Protein Containing Fumonisin Esterase and APAO activity in the Same Polypeptide.

5       The enzyme activities of fumonisin esterase and APAO can be combined in a single polypeptide by using the open reading frames together either with or without a spacer region between the two polypeptides. This creates a hybrid protein with dual enzyme activities that can be exported as a unit to the apoplast, and will allow both enzyme activities to be conveniently localized to the same area of the cell wall. The two cDNAs  
10      can be combined in either order, but the preferred method is to link them in the order NH<sub>3</sub>-Esterase:APAO-COOH. The spacer, if present, may consist of a short stretch of amino acids such as GGGSGGGS, or a set of amino acids that comprises a protease cleavage site that can be acted on by an apoplastic protease. This would result in the production of stoichiometric amounts of both esterase and APAO enzymes in the apoplast.  
15      Alternatively, a polycystronic message could be engineered which is capable of direct translation of a downstream sequence, for example inclusion of an IRES sequence in the spacer region or a polynucleotide spacer region containing a polynucleotide cleavage site that can be recognized by RNase or is a self-cleaving ribozyme. The length of the splice site could be of any length that ensures proper translation of the polynucleotide.

20      The esterase-APAO ligated protein can be made with any fumonisin esterase, including but not limited to, the fumonisin esterase from *E. spinifera* (ESP1) or fumonisin esterase from bacterium (BEST1). Since the pH range for maximum activity of BEST1 is similar to that of APAO (range 6.0 to 8.0), these may present the most effective combination in fusion form. In addition, any of the polynucleotides of the present  
25      invention, including APAO mutated to improve expression, may be used for an esterase-APAO ligation. As described in previous examples these fusion sequences can be placed in the appropriate expression vectors and used to express proteins in either bacteria or plants.

30      The nucleotide sequence of ESP1 contains three nucleotide differences and three corresponding amino acid differences for the ESP1 sequence disclosed in pending US application no. 08/888,950, filed July 7, 1997 and US patent no. 6,025,188, issued February 15, 2000. Both the sequences disclosed in the present application and the sequences disclosed in the pending US applications contain functional fumonisin esterase

genes. For the purposes of the present invention, either the original ESP1 sequences or the ESP1 sequences may be used in combination with the APAO sequences or in fusion sequences. The nucleotide sequence of a BAA:ESP1:trAPAO construct for plant expression can be found in SEQ ID NO: 24 and the translation in SEQ ID NO: 25. The 5 nucleotide sequence for a BAA:BEST1:K:trAPAO construct for plant expression can be found in SEQ ID NO: 26 and the translation in SEQ ID NO: 27. The nucleotide sequence of a GST:ESP1:K:trAPAO fusion for bacterial expression in a pGEX-4T-1 or similar vector can be found in SEQ ID NO: 28 and the translation in SEQ ID NO: 29. The nucleotide sequence for a GST:BEST1:K:trAPAO fusion for bacterial expression in a 10 pGEX-4T-1 or similar vector can be seen in SEQ ID NO: 30 and the translation in SEQ ID NO: 31.

15 **EXAMPLE 14**  
**APAO Substrate Studies**

The following assay was used to determine the substrate specificity of the APAO enzyme. Reaction mix: 436  $\mu$ l of 200 mM Na-phosphate, pH8.0; 50  $\mu$ l substrate (10 mM); 20 2  $\mu$ l Amplex Red (1 mg in 200  $\mu$ l DMSO); and 2  $\mu$ l of Peroxidase (5000 U/ml). The APAO enzyme was recombinant enzyme produced as GST fusion in *E. coli*, purified over a glutathione affinity column and cleaved with thrombin to remove the GST. All components were mixed at room temperature. The initial rate was determined in a spectrophotometer at 572 nm over one minute by absorbance units/second (BLANK). Ten 25 microliters of APAO at 70  $\mu$ g/ml was added and mixed. The initial rate was again determined at 572 nm over one minute in absorbance units/second (SAMPLE). The rates were converted to absorbance units/minute. The BLANK value was subtracted from the SAMPLE value. The absorbance units were converted to  $\mu$ M H<sub>2</sub>O<sub>2</sub> wherein 1  $\mu$ M H<sub>2</sub>O<sub>2</sub> equals 0.138 absorbance units at pH 8.0.

30

35

## SUBSTRATES FOR APAO

SUBSTRATE	RATE μM H <sub>2</sub> O <sub>2</sub> /min
1 mM Fumonisin B1	0.1429
1 mM AP1	0.8876
0.5mg/mL Fumonisin B2	0.3058
1 mM Fumonisin B3	0.1449
0.5mg/mL Fumonisin B4	0.1728
1 mM norepinephrine	0.0087
1 mM epinephrine	0.0071
1 mM dopamine	0.0040
1 mM spermine	0.0002

**NOT SUBSTRATES FOR APAO** (defined as compounds resulting in less than 1%

5 conversion to hydrogen peroxide by APAO relative to AP1 under similar conditions of time, pH, temperature, and substrate concentration): 2-phenylethylamine, spermidine, EDTA-Na<sub>2</sub>, tryptamine, putrescine, benzamidine, serotonin, cadaverine, Pefabloc SC, tyramine, 1,3-diaminopropane, leupeptin, histamine, hydroxylamine, aprotinin, deprenyl, 10 Fumonisin C4, isoniazid, sphingosine, phenelzine, sphinganine, phytosphingosine, D-alanine, DL-alanine, L-arginine, L-asparagine, L-aspartic acid, D-aspartic acid, L-cysteine, L-glutamine, L-glutamic acid, glycine, L-histidine, L-isoleucine, L-leucine, L-lysine, DL-lysine, L-methionine, DL-methionine, L-phenylalanine, L-proline, L-threonine, L-tryptophan, L-tyrosine, L-valine.

15

## EXAMPLE 15

### Sites on APAO for Possible Mutagenesis

Some cytosolic enzymes, when engineered for secretion by fusion with a heterologous signal peptide, lack function due to glycosylation at one or more potential glycosylation sites (amino acid consensus sequence N-X-S/T) that are not normally glycosylated in the native environment (Farrell *et al.*, *Plant Mol Biol* 15(6):821-5 (1990)). Since APAO lacks a recognizable signal sequence, it may be cytoplasmically localized in *Exophiala spinifera*, although secretion by some other method not involving a signal peptide cannot be ruled out. APAO contains two potential glycosylation sites, which may be glycosylated when APAO is secreted in a plant or other eukaryotic cell. Other modifications to APAO can be made to improve its expression in a plant system, including

site-directed mutagenesis to remove selected cysteine residues, which may be detrimental to proper folding when the protein is secreted into the endomembrane system for delivery to the apoplast.

Knowledge of the 3-dimensional structure of APAO would help to evaluate the likelihood that particular amino acids could contribute to misfolding, and increase the odds of making rational changes in the APAO sequence for successful secretion. To this end a 3-dimensional model of APAO was developed based on the crystal structure of a related amine oxidase from maize, maize polyamine oxidase or MPAO (Binda *et al.*, *Structure* 7:265-276 (1999)). The model was derived by automated modeling using the program *Modeler* (Molecular Simulations, Inc., San Diego, CA) and the resulting 3-D structure showed excellent fit based on an RMS deviation of 0.68 Å for the backbone coordinates of the two structures. The 3-D model of APAO based on MPAO is shown in Figures 1 and 2. Some of the possible mutations of APAO, which would result in removal of glycosylation sites or removal of cysteine residues can be seen below and in Figure 1.

15

**Table of site-directed mutagenesis vectors and enzyme assay results.**

Residue number	1	2	3	4	5	6	7	8	9	Glyc Site 1a,b	Glyc Site 2a,b	E coli expression vector, APAO or trAPAO activity	Maize expression vector, APAO or trAPAO Activity		
Residue position in APAO or trAPAO	C64	C109	C167	C292	C351	C359	C387	C461	C482	N201	S203	N204	S206		
Construct	Amino acid substitution											Plasmid	Act	Plasmid	Act
Wild type APAO-1												PHP13367	+	PHP	-
Glyc(-) 1a2a APAO								A	A			PHP16284	-	n/a	
Glyc(-) 1a2b APAO								A		A		PHP16285	-	n/a	
Glyc(-) 1b2a APAO								A	A			PHP16286	-	n/a	
Glyc(-) 1b2b APAO								A		A		PHP16287	-	n/a	
Glyc(-)2a N204A APAO									A			PHP16589	+/-	n/a	
Glyc(-)2b S206A APAO										A		PHP16590	+	PHP16711	-
Cys(-) #8 trAPAO							S					PHP16737	+	n/a	
Cys(-) #6,8 trAPAO						S	S					PHP16738	+		
Cys(-) #3,6,8 trAPAO			S		S	S						PHP17089	+ <sup>1</sup>		
Cys(-) #1,2,7 APAO	A	A				A									

A= alanine

S= serine

20 1 - activity against FB1 equals wild type; activity against AP1 was reduced.

#### **APAO and trAPAO polypeptide sequence, annotated. (SEQ ID NO: 47)**

The amino terminal domain is italicized. Cysteines and residues involved in putative glycosylation sites are underlined. Boxed residues represent amino acids that were successfully altered without complete loss of activity as E coli-expressed protein.

*MALAPSYINPPNVASPAGYSHVGVPDGGRYVTIAGQIGQDASGVTDPAYEKQVAQAFANLRACLAAVGATSNDVTKLNYYIVDYAPSKLTAIGDGLKATFALDRLLPCTLVPVSALSSP*

EYLFEVDATALVPGHTTPDNVADVVVVGAGLSGLETARKVQAAGLSCLVLEAMD  
RVGGKTLQSVQSGPGRTTINDLGAAWINDSNQSEVSRLFERFHLEGELQRTTGSIH  
QAQDGTTTAPYGDSSLSEEVASALAEELLVWSQLIEEHSILQDLKASPQAKRLDSV  
5 SFAHYCEKELNLPALGVANQITRALLGVEAHEISMLFLTDYIKSATGLSNIFSDKK  
DGGQYMRCKTGMQSIC~~G~~HAMSKELVPGSVHLNTPVAEIEQSASGCTVRSASAGAVF  
RSKKVVVSLPTTYPTLTFSPLPAEKQALAENSILGYYSKIVFVWDKPWWREQGF  
SGVLQSSCDPISFARDTSIDVDRQWSITCFMVGDGRKWSQQSKQVRQKSVDQL  
RAAYENAGAQVPEPANVLEIEWSKQQYFQGAPSAYGLNDLITLGSALRTPFKSV  
HFVGTETSLVWKGYMEAIRSGQRGAAEVVASLVPAA

10

APAO enzyme activity is maintained when a serine residue at position 206 is mutated to alanine, eliminating a potential glycosylation site (N204 – S206) close to the putative substrate binding site. Please see the tables entitled “Table of site-directed mutagenesis vectors and enzyme assay results” and “Glyc(-) APAO lysates from *E. coli*.”

15

The polynucleotide sequence of APAO mutated to alter the serine at position 206 to an alanine (S206A) can be seen in SEQ ID NO: 32. The resulting polypeptide is shown in SEQ ID NO: 33.

#### 20 Glyc(-) APAO lysates from *E. coli*

Sample (lysate)	Substrate	M H <sub>2</sub> O <sub>2</sub> /min <sup>1</sup>	Conclusion
WT APAO	AP1	1.92	Active (wild type)
	FB1	0.12	Slightly active (wt)
N204A	AP1	0.09	Slightly active
	FB1	0.04	Slightly active
S206A	AP1	0.85	Partially Active
	FB1	0.07	Slightly active

25 However, in transient expression assays in maize, expression of S206A resulted in no detectable enzyme activity. Please see the table above entitled “Table of site-directed mutagenesis vectors and enzyme assay results.” Thus, elimination of this glycosylation site is not in itself sufficient to have an active protein upon secretion. This could be due to glycosylation occurring at a second adjacent site (N201 - S203). However, no active APAO was recovered when either N201 or S203 is mutated along with S206. Please see the table entitled “Table of site-directed mutagenesis vectors and enzyme assay results.”

30 While not to be limited by theory, the molecule may be inactive because both N201 and S203 are buried within the tertiary structure of APAO, and any modification of side chains disrupts proper folding or conformation, or FAD binding. This is backed up by predicted solvent accessibility numbers for these residues in the 3-D model based on the

maize amine oxidase. Please see the table below entitled “Solvent accessibility for cysteine residues of truncated APAO.” The elimination of APAO glycosylation site at amino acids 204 to 206 is not sufficient to allow APAO to be secreted from the cell and retain full enzyme activity, but elimination of this site may improve chances for obtaining 5 a fully active enzyme once the other roadblock(s) to secretability have been resolved. In other words, elimination of this site may be necessary but not sufficient to produce active secretable APAO.

APAO also contains nine cysteine residues, which are likely to be unpaired in the reducing environment of the cytosol but which may crosslink unfavorably upon secretion. 10 Cysteines are present at residues 64, 109, 167, 292, 351, 359, 387, 461, and 482. The 3-D model helps predict the relative location of each amino acid in the structure, and whether it is solvent accessible or buried. Buried residues are more difficult to mutate without destroying structural integrity.

15

#### Solvent accessibility for cysteine residues of truncated APAO

APAO Position <sup>1</sup>	Position <sup>2</sup>	Cys# <sup>3</sup>	aa MPAO	-1	0	1	average	Conclusion	
Cys	26	167	3	Leu	32	0.675	0.253	0.24	0.389333 maybe partially exposed
Cys	151	292	4	Asn	147	0.069	0.122	0.147	0.112667 buried
Cys	210	351	5	Tyr	211	0.184	0.244	0.03	0.152667 buried
Cys	218	359	6	Thr	219	0.633	0.319	0.447	0.466333 maybe partially exposed
Cys	246	387	7	Val	247	0.145	0.046	0.366	0.185667 buried
Cys	320	461	8	Ser	324	0.199	0.789	0.643	0.543667 exposed
Cys	341	482	9	Leu	346	0.152	0.071	0.052	0.091667 buried

1. Relative to amino acid 1 of truncated APAO  
2. Relative to amino acid 1 of full length APAO  
20 3. Cysteine number relative to full length APAO

Proteins that are secreted to the apoplast are folded to their mature form in the highly oxidizing environment of the ER/Golgi. Among other things this promotes crosslinking of cysteine residues often found in secreted proteins. Unpaired cysteines that 25 are solvent-accessible are rare in secreted proteins, since they would rapidly be oxidized by other cysteine residues of the same protein or another protein. Although not to be limited by theory, it is possible that APAO is normally a cytosolic protein, and thus the presence of nine cysteine residues would not be unusual even though they may not be crosslinked in

the mature protein. In fact, the 3-D model predicts that they would not be crosslinked because the intermolecular distances predicted would be too great. Therefore it is possible that secretion of APAO to the apoplast results in an improper folding and crosslinking of cysteines in the Golgi, and results in inactive enzyme. Using the solvent accessibility tables from APAO modeled against MPAO, the three most solvent-exposed cysteines were identified and then eliminated by site-directed mutagenesis of the APAO cDNA. The sequence of APAO mutated at cysteine 461 and used for expression in bacteria can be seen in SEQ ID NO: 48. The resulting protein is shown in SEQ ID NO: 49. The polynucleotide and resulting polypeptide sequence of APAO mutated at both cysteines 359 and 461 and used for in the bacterial expression system can be seen in SEQ ID NOS: 50 and 51. The polynucleotide and resulting polypeptide sequence of APAO mutated at cysteines 169, 359, and 461 can be seen in SEQ ID NOS: 52 and 53.

The APAO molecules mutated at specific cysteines were tested in a bacterial expression system using the previously described Amplex Red assay. The results can be seen in the table below entitled “Cys(-) APAO lysates from *E. coli*.” The mutated APAO molecules can then be tested in maize, linked to a signal peptide, as previously described. Either one of the cysteines or two or three together could be mutated to serines without any measured loss in APAO enzyme activity of the *E. coli*-expressed enzyme. In fact, one of the *E. coli*-expressed clones (C359S + C461S; PHI16738) had more APAO activity in crude lysates than wild type enzyme and may represent a catalytic improvement. A triply Cys-mutated version of APAO does not show catalytic improvement but retains full activity of the wild type enzyme against FB1, although AP1 activity was somewhat reduced. The mutated versions of APAO operably linked to a signal sequence, which retain function when expressed as recombinant fusion proteins in *E. coli*, may also provide additional stability or foldability when expressed in plants or other secretion expression systems.

**Cys(-) APAO lysates from *E. coli***

Sample (lysate)	Substrate	M H <sub>2</sub> O <sub>2</sub> /min <sup>1</sup>	Conclusion
WT APAO	AP1	2.14	Active (wild type)
	FB1	0.11	Slightly active (wt)
C461S	AP1	2.25	Fully Active
	FB1	0.14	Slightly active
C359S, C461S	AP1	3.90	Fully/More Active
	FB1	0.16	Slightly active
C167S, C359S, C461S	AP1	0.27	Slightly active
	FB1	0.25	Slightly active

**Triple Cys(-) APAO lysates from *E. coli***

Sample (lysate)	Substrate	M H <sub>2</sub> O <sub>2</sub> /min <sup>1</sup>	Conclusion
WT APAO	AP1	1.16	Active (wild type)
	FB1	0.27	Slightly active (wt)
C167S, C359S, C461S	AP1	0.27	Slightly Active
	FB1	0.26	Slightly active

It is expected that the S206A mutations will contribute to the functionality of secreted APAO by reducing the degree of glycosylation and the C167S, C359S, and C461S mutations (or combinations thereof) will improve the functionality of secreted APAO by reducing chances for spurious disulfide formation on folding.

To determine expression of a mutated APAO in maize, three APAO constructs were introduced into maize embryos by Agrobacterium-mediated transformation (Zhao et al, 1999, US Patent 5,981,840). The three constructs were PHP17105 (Ubi:BAA:Cys(-)K-trAPAO (C359S, C461S):PinII), PHP17108 (Ubi:Cys(-)K-trAPAO (C359S,

C461S):PinII), and PHP17110 (Ubi:APAO:PinII). In addition, PHP16543 (NOS:CRC:PinII-Ubi:MO-PAT:T35) was introduced as a negative control and PHP15258 (Ubi:APAO:PinII-Ubi:BAA:ESP1:PinII-P35S:PAT:T35S) was introduced as a non-targeted positive control. One experiment with two replications was performed. Samples were assayed for both APAO activity by TLC as described previously and by Enzyme

Linked ImmunoSorbent Assay (ELISA). For a discussion of ELISA methods, please see, for example, *Current Protocols in Molecular Biology*, 2:11.1.1-11.3.4, John Wiley & Sons, Inc. (Ausubel, et al., eds. 1994). The APAO ELISA is a capture format assay for the quantitative determination of APAO protein in the presence of extracted maize tissue protein. It was performed by co-incubation of biotinylated antibody with an extract

prepared from leaf, seed, or callus in phosphate buffered saline with 0.5% Tween-20®. The detection of the antibody complex was accomplished through the added incubation of

streptavidin-alkaline phosphatase (Bio-Rad Life Sciences Products #19542-018), followed by the addition of substrate (pNPP tablets, Sigma #104-105). The resultant color intensity was quantified by determining optical density and was directly proportional to the amount of APAO protein present in the sample extracts. The assay has no matrix effects at 5  $1\mu\text{g}/\text{well}$  or below for maize leaf, seed, or callus protein. The standard curve was spiked with wild type extract at levels above  $1.0\ \mu\text{g}/\text{well}$ . The transient testing results are summarized in the table below.

10

**Transient Testing of APAO Constructs (6-8-2000)**

Experiment	Rep	Construct	APAO-TLC	APAO-ELISA (ppm)
negative control	none	none	0	-2
4350.08.01	1	php16543, as a (-) control	0	-4
4350.08.02		php15258, non-targeted APAO as a (+) control	3	out high
4350.08.03		php17105, UBI-BAA::CYS(-)K-TR-APAO (C359S, C461S)	1	107
4350.08.04		php17108, UBI-CYS(-)K-TR-APAO (C359S, C461S)	3	270
4350.08.05		php17110, UBI-APAO	3	out high
4350.08.06	2	php16543	0	-5
4350.08.07		php15258	3	313
4350.08.08		php17105	0	52
4350.08.09		php17108	2	143
4350.08.10		php17110	2	123
3477.27.01	transformed callus lines	php15258 as positive controls	1	118
3477.27.02			2	141
3477.27.03			2	187
3477.27.04			2	184

As can be seen in the Table above, the BAA-targeted APAO (PHP17105) did not 15 accumulate as much APAO as the non-BAA targeted counterpart (PHP17108). Although not to be limited by theory, the lack of APAO protein accumulation rather than APAO function may play a role in the lack of detectable APAO activity with the BAA-targeted APAO construct. It appears that only when the APAO concentration exceeds 100 ppm can 20 APAO activity be seen by TLC. Nevertheless, the double Cys(-) mutant is active in maize when expressed either cytosolically or extracellularly.

### EXAMPLE 16

#### Other APAO Polynucleotides From *Exophiala spinifera* and *Rhinocladiella atrovirens*

Using primers designed from the APAO isolated from *Exophiala spinifera*, ATCC 25 74269(Table 15), three new APAO polynucleotides were isolated from *Exophiala spinifera* (isolates ESP002 and ESP003), designated ESP002\_C2, ESP002\_C3 and ESP003\_C12

and three new APAO polynucleotides from *Rhinocladiella atrovirens* (isolate RAT011) designated RAT011\_C1, RAT011\_C2, RAT011\_C4. The strains used to isolate the polynucleotides are described below.

5

Isolate	Genus species	Source	FB1 degrader	APAO homologs isolated
ESP002	<i>Exophiala spinifera</i>	Palm, ATCC 26089	Yes	ESP002_c2 in pGEX4T1 ESP002_c3 in pGEX4T1
ESP003	<i>Exophiala spinifera</i>	Maize seed	Yes	ESP003_c12 in pGEX4T1
RAT011	<i>Rhinocladiella atrovirens</i>	Maize seed	Yes	RAT011_c1 in pGEM11Zf+ RAT011_c2 in pGEX4T1 RAT011_c4 in pGEM11Zf+

### Growth conditions and production of culture material

- 10 1. Streak 150 x 15 mm YPD plates with a glycerol aliquot of the above fungal isolates.
2. Grow at 28° C in the dark until there is sufficient growth for inoculating liquid medium usually at least two weeks.
- 15 3. Mycelia and spores were scraped from the plates or agar cubes used to inoculate 50 mls YPD broth in 250 ml baffled flasks.
4. Flasks of culture material were grown at 28° C in the dark at ~125 rpm.
5. After sufficient growth was obtained the cultures were harvested by pelleting the culture in 50 ml centrifuge tubes at 3400 rpm for 15 min.
6. The supernatant was discarded and the pellets were frozen at -20° C.

### YPD broth and agar medium

Amount per liter:	Yeast Extract	10 g
	Bactopeptone	20 g
25	Dextrose	0.5 g
	Bactoagar	15 g (for agar media only)

### 30 DNA Isolation,

The DNA was isolated according to a modified version of a plant CTAB DNA extraction protocol (Saghai-Marof MA, *et al.*, *Proc Natl Acad Sci, USA*, 81:8014-8018 (1984)) as follows.

- 35 1. Place 0.2-0.5 g (dry weight) lyophilized fungal mycelium in a 50 ml disposable centrifuge tube, break up mat with a spatula or glass rod. Shake briefly.
2. Add 10 ml (per 0.5 g mat) of CTAB extraction buffer. Gently mix to wet all the powdered mat.

3. Place in 65° C water bath for 30 minutes.  
4. Cool. Add an equal volume of phenol:chloroform. Shake briefly to mix.  
5. Centrifuge 20 minutes at 3400 rpm.  
6. To the aqueous phase add an equal volume of chloroform:isoamyl alcohol  
(24:1). Shake briefly to mix.  
5  
7. Centrifuge 15 minutes at 3400 rpm.  
8. To aqueous phase add an equal volume of isopropanol.  
9. Centrifuge for 30 minutes at 3400 rpm to pellet precipitated DNA.  
10. Rinse DNA pellet with 70% ethanol.  
10  
11. Air dry pellet.  
12. Resuspend pellet in 1-5 ml TE containing 20 ug/ml RNase A.

**CTAB Extraction Buffer**

15 0.1 M Tris, pH 7.5  
1% CTAB (mixed hexadecyl trimethyl ammonium bromide)  
0.7 M NaCl  
10 mM EDTA  
1% 2-mercaptoethanol  
20 Add proteinase K to a final concentration of 0.3 mg/ml prior to use.

**Primer Design**

25 Primers used were gene specific primers based on APAO polynucleotide sequence  
(SEQ ID NO: 22) with restriction enzymes sites for cloning. The 5'-primer, 26194,  
contained the restriction enzyme recognition site, EcoRI. The complementary 3'-primer,  
23259, contained the restriction enzyme recognition site, NotI.

30 26194  
5' ggggaattcATGGCACTTGCACCGAGCTACATCAATC 3' , 37-mer (SEQ ID NO: 34)

23259  
5' gggGCGGCCGCCTATGCTGCTGGCACCCAGGCTAG 3' , 34-mer (SEQ ID NO: 13)

35

**PCR conditions**

40

45

5	1.	The PCR cocktail: per 50 ul reaction per 0.2 ml tube	10 mM dNTPs 10X Advantage polymerase buffer HPLC water 10 uM primer 26194 10 uM primer 23259 50 X Advantage polymerase mix	1 ul 5 ul 38 ul 2 ul 2 ul 1 ul
---	----	--	---	---

10 (Clontech)  
Template, genomic DNA, 50 ng/ $\mu$ l 1  $\mu$ l

2. Thermocycling conditions:  
 MJ PTC-100 AgV Thermocycler:

Step	1	95°	2 minutes
	2	95°	30 seconds
	3	60°	1 minute
	4	72°	1 minute 30 seconds
	5	Go to step 2, 34X more	
	6	72°	5 minutes
	7	4°	Hold
	8	End	

3. PCR products were analyzed on a 1% LE-agarose, TAE plus ethidium bromide gel. Bands of about 1900 bp were seen on the gel. The band was not present in the no DNA control reaction.

## Cloning Protocols

1. DNA was extracted from excised gel fragments using a QIAGEN Gel Extraction Kit (Catalog number 28704, QIAGEN, Santa Clara, CA).
  2. PCR fragments were digested with EcoRI and Not I to free up the sites for cloning into EcoRI and Not I digested vector, either pGEX4T1 (Pharmacia) or pGEM11Zf+ (Promega).
  3. Digests were cleaned up and desalted used a QIAquick PCR Purification Kit (Catalog number 28104).
  4. Isolated fragment was quantified and checked for purity on a 1% LE-agarose, TAE + ethidium bromide gel.
  5. Fragments were ligated into compatible sites in either pGEX4T1 (Pharmacia) or pGEM11Zf+ (Promega).
  6. After heat inactivation Library efficiency DH5<sup>r</sup> competent *E. coli* were transformed with a small amount of the ligation reaction.
  7. LB + carbenicillin, 50 ug/ml, plates were spread with an aliquot of the transformation mix, grown overnight at 37° C.
  8. Colonies were screened for full-length insert using a PCR miniprep method utilizing vector primers flanking the multiple cloning region.

9. Positive clones were identified and overnight cultures grown for plasmid isolation and verification by sequencing.

10. Positive clones are identified as follows:

5 DH5 :pGEX4T1:ESP002FL\_c2 ( from palm tree isolate)

DH5 :pGEX4T1:ESP002FL\_c3 ( from palm tree isolate)

DH5 :pGEX4T1:ESP003FL\_c12 ( from maize isolate)

DH5 :pGEM11Zf+:RAT011FL\_c1 ( from maize isolate)

DH5 :pGEM11Zf+:RAT011FL\_c4 ( from maize isolate)

DH5 :pGEX4T1:RAT011FL\_c2 ( from maize isolate)

10 \*\*Important note: These are genomic clones containing two introns

## Sequence Results

Three APAO polynucleotides and related polypeptides were isolated from

15 *Exophiala spinifera* (isolates ESP002 and ESP003), designated ESP002\_C2, (SEQ ID

NOS: 35 and 36) ESP002\_C3 (SEQ ID NOS: 37 and 38) and ESP003\_C12 (SEQ ID NOS:

39 and 40). Three APAO polynucleotides were isolated from *Rhinocladiella atrovirens*

(isolate RAT011) designated RAT011\_C1 (SEQ ID NOS: 41 and 42), RAT011\_C2 (SEQ

ID NOS: 43 and 44), and RAT011\_C4 (SEQ ID NOS: 45 and 46). Introns were detected

20 by comparison of the genomic sequence with the cDNA sequence of APAO from *E.*

*spinifera* 2141.10 (SEQ ID NO: 22), and by identifying putative intron splice junctions in

the gap domains (Shah, *et al.*, *Journal of Molecular and Applied Genetics* 2:111-126

(1983)).

Plasmids containing the polynucleotide sequences of the invention were deposited

25 with American Type Culture Collection (ATCC), Manassas, Virginia, and assigned

Accession No. 98812, 98813, 98814, 98815, 98816, (all deposited on July 15, 1998) and

PTA-32 (deposited on May 7, 1999). The deposits will be maintained under the terms of

the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for

the Purposes of Patent Procedure. The deposits were made merely as a convenience for

30 those of skill in the art and are not an admission that a deposit is required under 35 U.S.C.

§ 112.

Preliminary sequence results were entered into GCG, and nucleotide and protein

alignments were done in a pileup using a software program called Genedoc for shading and

homology comparisons (Nicholas, *et al.*, *EMBNEW.NEWS* 4:14 (1997; or at the Internet

35 site <http://www.cris.com/~Ketchup/genedoc.shtml>). The first APAO (SEQ ID NO: 22)

sequence was included for comparison. Comparing the reference sequence SEQ ID NO:

22 to the other homologs sequence identities range from 96 to 99% (identities are lower  
since APAO introns were not included). Homologies are slightly higher comparing  
*Exophiala* genes sequences. At the amino acid sequence level the comparison of the  
reference sequence (SEQ ID NO: 23) to the other homologs yielded sequence identities of  
5 approximately 97%.

All publications and patent applications in this specification are indicative of the  
level of ordinary skill in the art to which this invention pertains. All publications and  
patent applications are herein incorporated by reference to the same extent as if each  
individual publication or patent application was specifically and individually indicated by  
10 reference.

The invention has been described with reference to various specific and preferred  
embodiments and techniques. However, it should be understood that many variations and  
modifications may be made while remaining within the spirit and scope of the invention.

WHAT IS CLAIMED IS:

1. An isolated polynucleotide comprising an APAO encoding polynucleotide linked to a fumonisin esterase encoding polynucleotide, wherein the APAO encoding polynucleotide comprises a member selected from:
  - a) a polynucleotide encoding a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53;
  - b) a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 33, 35, 37, 39, 41, 43, 45, 48, 50 and 52; and
  - c) a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52.
2. A recombinant expression cassette comprising a polynucleotide of claim 1 operably linked to a promoter.
3. The recombinant expression cassette of claim 2 wherein the polynucleotide is operably linked to a plant signal sequence.
4. A vector comprising the recombinant expression cassette of claim 2.
5. A host cell comprising the recombinant expression cassette of claim 2.
6. The host cell of claim 5 wherein the cell is a plant cell.
7. The host cell of claim 6 wherein the plant cell is selected from the group consisting of maize, sorghum, wheat, tomato, soybean, alfalfa, sunflower, canola, cotton, barley, millet, and rice.
8. A plant comprising a polynucleotide of claim 1.
9. A seed from a plant of claim 7.

10. An isolated polypeptide comprising a member selected from:

- a) a polypeptide comprising at least 70% sequence identity to a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53;
- b) a polypeptide encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52; and
- c) a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53.

11. The polynucleotide of claim 1 wherein the fumonisin esterase encoding polynucleotide is ESP1.

12. The polynucleotide of claim 11 wherein the polynucleotide is set forth in SEQ ID NO: 24.

13. The polynucleotide of claim 1 wherein the fumonisin esterase encoding polynucleotide is BEST1.

14. The polynucleotide of claim 13 wherein the polynucleotide is set forth in SEQ ID NO: 26.

15. A method of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a breakdown product of a structurally related mycotoxin comprising the steps of:

- a) applying an APAO enzyme as a spray or wash; and
- b) under degradation conditions allowing sufficient time for the polypeptide to degrade the fumonisin, the structurally related mycotoxin, the fumonisin breakdown product, or the breakdown product of a structurally related mycotoxin.

16. The method of claim 15 wherein the fumonisin or structurally related mycotoxin is present in harvested grain.

17. The method of claim 15 wherein degradation occurs during processing of the harvested grain.
18. The method of claim 17 wherein the harvested grain is to be used as animal feed.
19. The method of claim 15 wherein degradation occurs in silage.
20. The method of claim 15 wherein fumonisin esterase is also added at or before step (a).
21. The method of claim 15 wherein the APAO enzyme is selected from:  
a) a polypeptide comprising at least 70% sequence identity to a polypeptide selected from SEQ ID NOS: 6, 11, 23, 33, 36, 38, 40, 42, 44, 46, 49, 51 and 53;  
b) a polypeptide encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52; and  
c) a polypeptide selected from SEQ ID NOS: 6, 11, 23, 36, 38, 40, 42, 44, 46, 49, 51 and 53.
22. A method of identifying transformed plant cells comprising the steps of:  
a) introducing into a plant cell at least one copy of an expression cassette comprising an APAO encoding polynucleotide;  
b) placing the plant cell on culture media containing an AP1 or a phytotoxic analog; and  
c) identifying transformed cells as the surviving cells in the culture.
23. The method of claim 22 wherein the APAO encoding polynucleotide comprises a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, and 50.
24. The method of claim 22 wherein a fumonisin esterase encoding polynucleotide is also introduced into the plant cell.

25. A method of detecting fumonisins or structurally related toxins, the method comprising:

- a) adding APAO enzymes to a sample containing fumonisin or a structurally related toxin;
- b) reacting the sample under conditions of time and temperature sufficient to convert the toxin to the corresponding oxidized or deaminated toxin; and
- c) detecting the hydrogen peroxide or ammonia produced.

26. The method of claim 25 wherein the APAO enzyme is encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, and 50.

27. The method of claim 25 wherein fumonisin esterase is added at or before step (a).

28. A method of producing a plant capable of degrading fumonisin, a structurally related mycotoxin, a fumonisin breakdown product, or a breakdown product of a structurally related mycotoxin comprising the steps of:

- a) introducing into a plant cell at least one copy of an expression cassette comprising a polynucleotide encoding an APAO enzyme operably linked to a promoter; and
- b) under degradation conditions expressing the APAO enzyme for a time sufficient to degrade the fumonisin, the fumonisin breakdown product, the structurally related mycotoxin, AP1, or a breakdown product of a structurally related mycotoxin.

29. The method of claim 28 wherein a polynucleotide encoding a fumonisin esterase is also introduced.

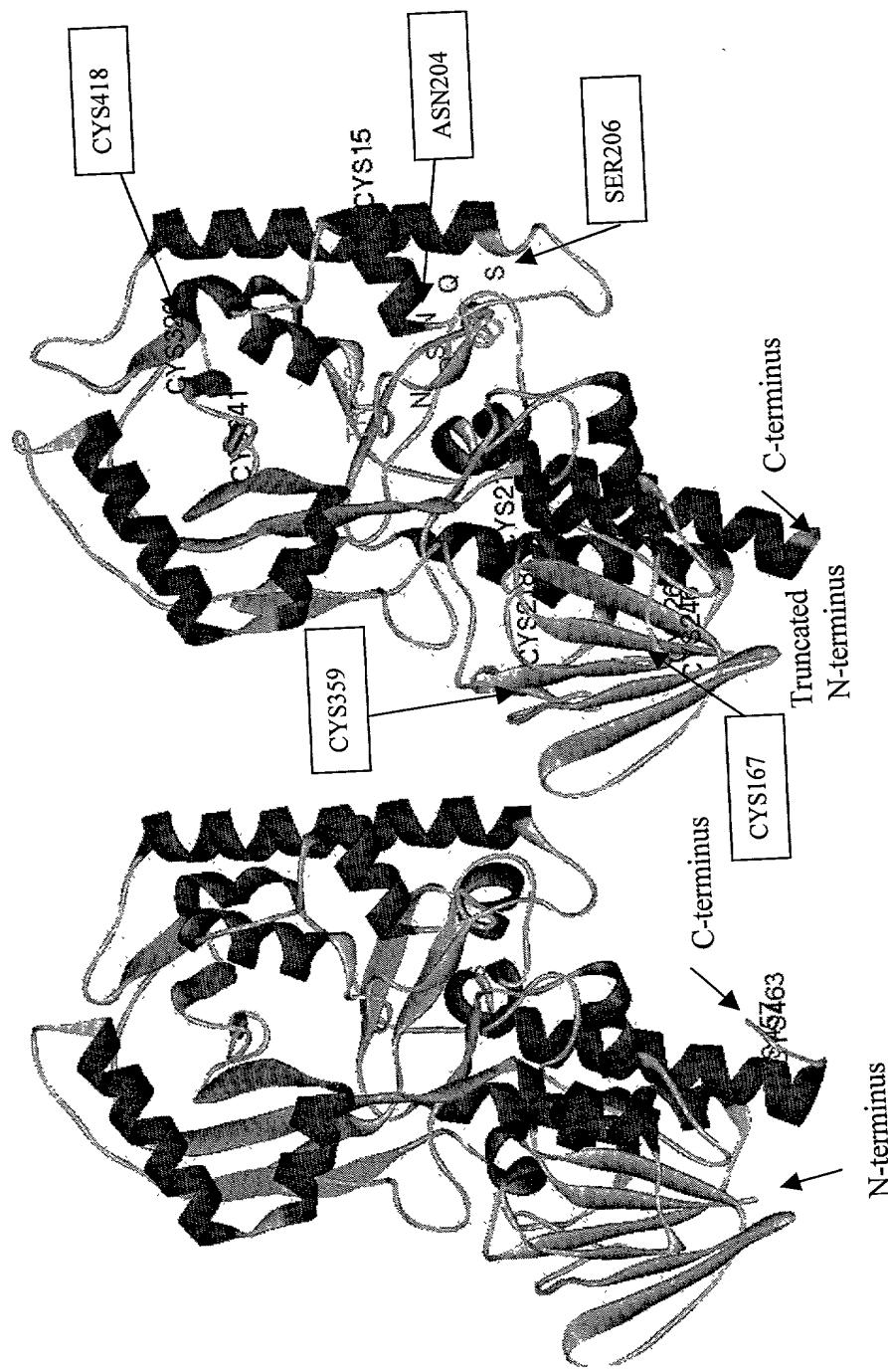
30. The method of claim 28 wherein the APAO enzyme is encoded by a polynucleotide having at least 70% sequence identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, 50 and 52.

31. The method of claim 28 wherein the plant cell is regenerated into a plant.
32. The method of claim 28 wherein a fumonisin esterase encoding polynucleotide is also introduced.
33. A host cell comprising an APAO encoding polynucleotide and a fumonisin esterase encoding polynucleotide.
34. The host cell of claim 33 wherein the APAO encoding polynucleotide comprises a polynucleotide having at least 70% identity to a polynucleotide selected from SEQ ID NOS: 5, 10, 22, 32, 35, 37, 39, 41, 43, 45, 48, and 50.
35. The host cell of claim 33 wherein the fumonisin esterase encoding polynucleotide is selected from ESP1 and BEST1.
36. The host cell of claim 33 wherein the cell is a plant cell.
37. The host cell of claim 36 wherein the cell is selected from maize, sorghum, wheat, tomato, soybean, alfalfa, sunflower, canola, cotton, and rice.
38. The host cell of claim 37 wherein the plant cell is regenerated into a plant.
39. A method of predicting possible mutagenesis sites on APAO comprising the steps of:
- a) developing a 3-dimensional model of APAO; and
  - b) identifying sites on APAO to mutate by evaluating the likelihood that particular amino acids could contribute to misfolding.
40. A 3-dimensional model of APAO generated by an automated modeling program.
41. The model of claim 40 wherein the automated modeling program is *Modeler*.

AMINO POLYOL AMINE OXIDASE POLYNUCLEOTIDES AND RELATED  
POLYPEPTIDES AND METHODS OF USE

5   **Abstract**

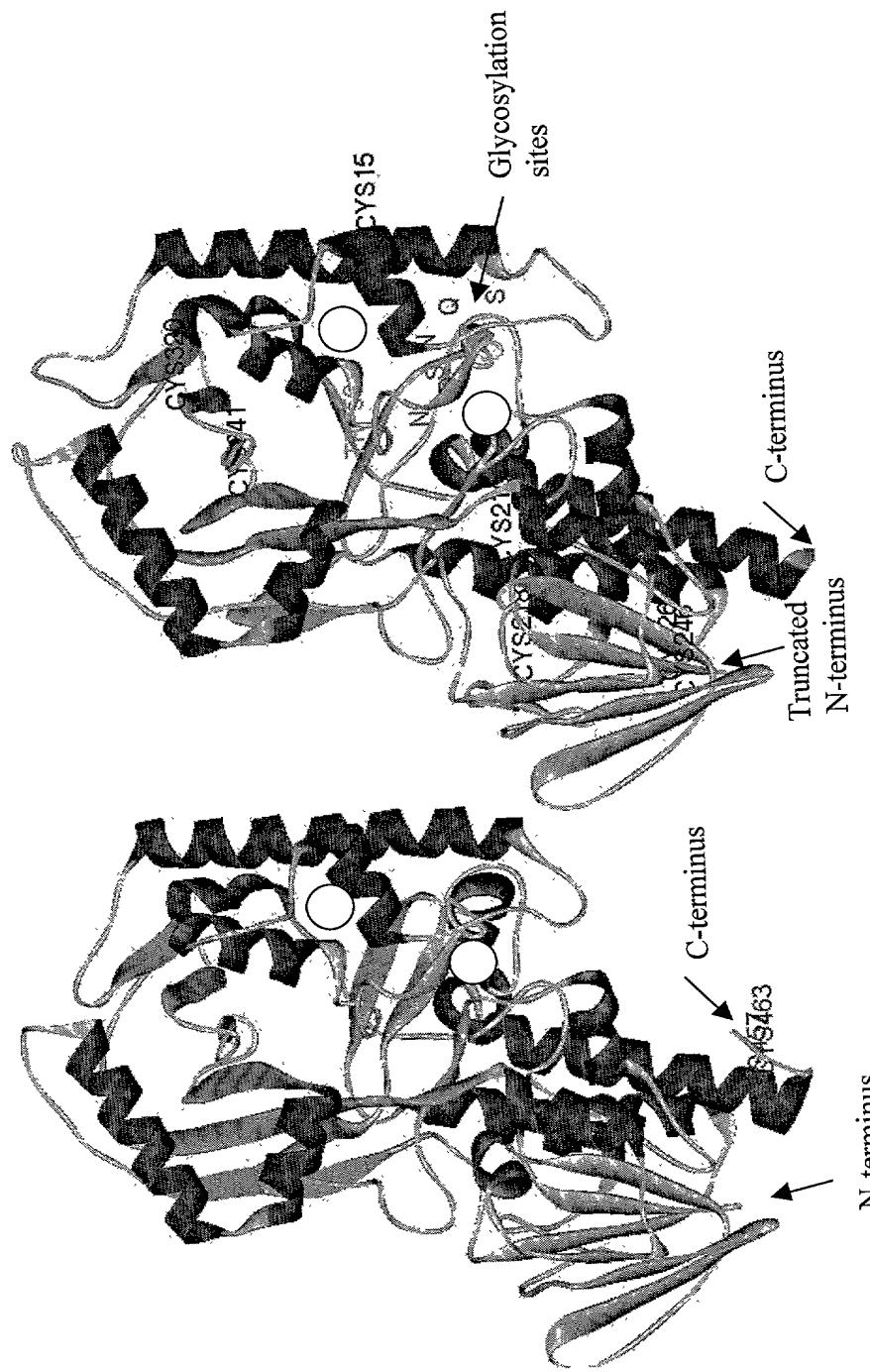
The present invention provides polynucleotides and related polypeptides of the enzyme APAO isolated from *Exophiala spinifera* and *Rhinocladiella atrovirens*. The polynucleotides may be mutated to remove glycosylation sites and cysteine residues. Additionally, the present invention provides recombinant expression cassettes, host cells, 10 transgenic plants, and transgenic seed. The present invention also provides for polynucleotides containing both APAO and a fumonisin esterase. In addition, the present invention provides methods for producing the APAO enzyme in both prokaryotic and eukaryotic systems, methods for detecting fumonisins, and methods for identifying transformed plant cells. Methods for degrading fungal toxins in plants, grain, grain 15 processing, silage, food crops and in animal feed are also disclosed.



A

Figure 1

B



B

Figure 2

A

SEQUENCE LISTING

<110> Duvick, Jonathan P.  
 Gilliam, Jacob T.  
 Maddox, Joyce R.  
 Rao, Aragula Gururaj  
 Crasta, Oswald R.  
 Folkerts, Otto

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 Polynucleotides and Related Polypeptides and Methods of Use

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ggt gct gca gaa gtt gtg gct agc ctg gtg cca gca gca tag Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala 450		455	460	1442

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 <211> 462  
 <212> PRT  
 <213> Exophiala spinifera

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 20 25 30  
 Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser Gly  
 35 40 45  
 Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp  
 50 55 60  
 Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu Glu  
 65 70 75 80  
 Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp  
 85 90 95  
 Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu

100	105	110
Val Ala Ser Ala Leu Ala Glu	Leu Pro Val Trp Ser Gln	Leu Ile
115	120	125
Glu Glu His Ser Leu Gln Asp	Leu Lys Ala Ser Pro Gln	Ala Lys Arg
130	135	140
Leu Asp Ser Val Ser Phe Ala His	Tyr Cys Glu Lys	Glu Leu Asn Leu
145	150	155
Pro Ala Val Leu Gly Val Ala Asn	Gln Ile Thr Arg Ala	Leu Leu Gly
165	170	175
Val Glu Ala His Glu Ile Ser Met	Leu Phe Leu Thr Asp Tyr	Ile Lys
180	185	190
Ser Ala Thr Gly Leu Ser Asn Ile	Phe Ser Asp Lys Lys	Asp Gly Gly
195	200	205
Gln Tyr Val Arg Cys Lys Thr	Gly Met Gln Ser Ile Cys	His Ala Met
210	215	220
Ser Lys Glu Leu Val Pro Gly	Ser Val His Leu Asn Thr	Pro Val Ala
225	230	235
Glu Ile Glu Gln Ser Ala Ser	Gly Cys Thr Val Arg	Ser Ala Ser Gly
245	250	255
Ala Val Phe Arg Ser Lys Lys	Val Val Ser Leu Pro	Thr Thr Leu
260	265	270
Tyr Pro Thr Leu Thr Phe Ser	Pro Pro Leu Pro Ala	Glu Lys Gln Ala
275	280	285
Leu Ala Glu Asn Ser Ile Leu	Gly Tyr Tyr Ser Lys Ile	Val Phe Val
290	295	300
Trp Asp Lys Pro Trp Trp Arg	Glu Gln Gly Phe Ser Gly	Val Leu Gln
305	310	315
Ser Ser Cys Asp Pro Ile Ser	Phe Ala Arg Asp Thr	Ser Ile Asp Val
325	330	335
Asp Arg Gln Trp Ser Ile Thr	Cys Phe Met Val Gly Asp	Pro Gly Arg
340	345	350
Lys Trp Ser Gln Gln Ser Lys	Gln Val Arg Gln Lys	Ser Val Trp Asp
355	360	365
Gln Leu Arg Ala Ala Tyr	Glu Asn Ala Gly Ala	Gln Val Pro Glu Pro
370	375	380
Ala Asn Val Leu Glu Ile	Glu Trp Ser Lys Gln Gln	Tyr Phe Gln Gly
385	390	395
Ala Pro Ser Ala Val Tyr	Gly Leu Asn Asp Leu Ile	Thr Leu Gly Ser
405	410	415
Ala Leu Arg Thr Pro Phe Lys	Ser Val His Phe Val	Gly Thr Glu Thr
420	425	430
Ser Leu Val Trp Lys Gly	Tyr Met Glu Gly Ala Ile	Arg Ser Gly Gln
435	440	445
Arg Gly Ala Ala Glu Val Val	Ala Ser Leu Val Pro	Ala Ala
450	455	460

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 <211> 458  
 <212> PRT  
 <213> Exophiala spinifera

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Glu Thr Ala Arg Lys Val Gln	Ala Ala Gly Leu Ser Cys	Leu Val Leu	
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Glu Ala Met Asp Arg Val	Gly Lys Thr Leu Ser Val Gln	Ser Gly	
35	40	45	
Pro Gly Arg Thr Thr Ile	Asn Asp Leu Gly Ala Ala	Trp Ile Asn Asp	
50	55	60	
Ser Asn Gln Ser Glu Val	Ser Arg Leu Phe Glu Arg	Phe His Leu Glu	
65	70	75	80

Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp  
                   85                  90                  95  
 Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu  
                   100                  105                  110  
 Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu Ile  
                   115                  120                  125  
 Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg  
                   130                  135                  140  
 Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn Leu  
                   145                  150                  155                  160  
 Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly  
                   165                  170                  175  
 Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys  
                   180                  185                  190  
 Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly Gly  
                   195                  200                  205  
 Gln Tyr Val Arg Cys Lys Thr Gly Ala Cys Gly Val Val Ser Gly Gly  
                   210                  215                  220  
 Gly Leu Val Ser Gln Trp Ser Phe Gln Val Cys Ser Arg Phe Ala Met  
                   225                  230                  235                  240  
 Pro Cys Gln Arg Asn Leu Phe Gln Ala Gln Cys Thr Ser Thr Pro Pro  
                   245                  250                  255  
 Ser Leu Lys Leu Ser Ser Arg His Pro Ala Val Gln Tyr Asp Arg Pro  
                   260                  265                  270  
 Arg Ala Pro Cys Ser Glu Ala Lys Arg Trp Trp Phe Arg Tyr Arg Gln  
                   275                  280                  285  
 Pro Cys Ile Pro Pro His Phe His His Leu Phe Pro Pro Arg Ser Lys  
                   290                  295                  300  
 His Trp Arg Lys Ile Leu Ser Trp Ala Thr Ile Ala Arg Ser Ser Tyr  
                   305                  310                  315                  320  
 Gly Thr Ser Arg Gly Gly Ala Asn Lys Ala Ser Arg Ala Ser Ser Asn  
                   325                  330                  335  
 Arg Ala Val Thr Pro Ser His Leu Pro Glu Ile Pro Ala Ser Thr Ser  
                   340                  345                  350  
 Ile Asp Asn Gly Pro Leu Pro Val Ser Trp Ser Glu Thr Arg Asp Gly  
                   355                  360                  365  
 Ser Gly Pro Asn Ser Pro Ser Arg Tyr Asp Lys Ser Leu Ser Gly Thr  
                   370                  375                  380  
 Asn Ser Ala Gln Pro Thr Arg Thr Pro Gly Pro Lys Ser Gln Ser Arg  
                   385                  390                  395                  400  
 Pro Thr Cys Ser Lys Ser Ser Gly Arg Ser Ser Ser Ile Ser Lys Glu  
                   405                  410                  415  
 Leu Arg Ala Pro Ser Met Gly Thr Ile Ser Ser His Trp Val Arg Arg  
                   420                  425                  430  
 Ser Glu Arg Arg Ser Arg Val Phe Ile Ser Leu Glu Arg Arg Arg Leu  
                   435                  440                  445  
 Phe Gly Lys Gly Ile Trp Lys Gly Pro Tyr  
                   450                  455

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<210> 10
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<212> DNA
<213> Exophiala spinifera
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1	5	10	15	
ttg gag acg gca cgc aaa gtc cag gcc gcc ggt ctg tcc tgc ctc gtt				96
Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val				
20	25	30		
ctt gag gcg atg gat cgt gta ggg gga aag act ctg agc gta caa tcg				144
Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser				
35	40	45		
ggc ccc ggc agg acg act atc aac gac ctc ggc gct gcg tgg atc aat				192
Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn				
50	55	60		
gac agc aac caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg				240
Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu				
65	70	75	80	
gag ggc gag ctc cag agg acg act gga aat tca atc cat caa gca caa				288
Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln				
85	90	95		
gac ggt aca acc act aca gct cct tat ggt gac tcc ttg ctg agc gag				336
Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu				
100	105	110		
gag gtt gca agt gca ctt gcg gaa ctc ctc ccc gta tgg tct cag ctg				384
Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu				
115	120	125		
atc gaa gag cat agc ctt caa gac ctc aag gcg agc cct cag gcg aag				432
Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys				
130	135	140		
cgg ctc gac agt gtg agc ttc gcg cac tac tgt gag aag gaa cta aac				480
Arg Leu Asp Ser Val Phe Ala His Tyr Cys Glu Lys Glu Leu Asn				
145	150	155	160	
ttg cct gct gtt ctc ggc gta gca aac cag atc aca cgc gct ctg ctc				528
Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu				
165	170	175		
ggt gtg gaa gcc cac gag atc agc atg ctt ttt ctc acc gac tac atc				576
Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile				
180	185	190		
aag agt gcc acc ggt ctc agt aat att ttc tcg gac aag aaa gac ggc				624
Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly				
195	200	205		
ggg cag tat atg cga tgc aaa aca ggt atg cag tcg att tgc cat gcc				672
Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala				
210	215	220		
atg tca aag gaa ctt gtt cca ggc tca gtg cac ctc aac acc ccc gtc				720
Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val				
225	230	235	240	

gct gaa att gag cag tcg gca tcc ggc tgt aca gta cga tcg gcc tcg Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser 245 250 255	768
ggc gcc gtg ttc cga agc aaa aag gtg gtg gtt tcg tta ccg aca acc Gly Ala Val Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr 260 265 270	816
ttg tat ccc acc ttg aca ttt tca cca cct ctt ccc gcc gag aag caa Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln 275 280 285	864
gca ttg gcg gaa aat tct atc ctg ggc tac tat agc aag ata gtc ttc Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe 290 295 300	912
gta tgg gac aag ccg tgg tgg cgc gaa caa ggc ttc tcg ggc gtc ctc Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu 305 310 315 320	960
caa tcg agc tgt gac ccc atc tca ttt gcc aga gat acc agc atc gac Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp 325 330 335	1008
gtc gat cga caa tgg tcc att acc tgt ttc atg gtc gga gac ccg gga Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly 340 345 350	1056
cg <sup>g</sup> aag tgg tcc caa cag tcc aag cag gta cga caa aag tct gtc tgg Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp 355 360 365	1104
gac caa ctc cgc gca gcc tac gag aac gcc ggg gcc caa gtc cca gag Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu 370 375 380	1152
ccg gcc aac gtg ctc gaa atc gag tgg tcg aag cag cag tat ttc caa Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln 385 390 395 400	1200
gga gct ccg agc gcc gtc tat ggg ctg aac gat ctc atc aca ctg ggt Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly 405 410 415	1248
tcg gcg ctc aga acg ccg ttc aag agt gtt cat ttc gtt gga acg gag Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu 420 425 430	1296
acg tct tta gtt tgg aaa ggg tat atg gaa ggg gcc ata cga tcg ggt Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly 435 440 445	1344
caa cga ggt gct gca gaa gtt gtg gct agc ctg gtg cca gca gca Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala 450 455 460	1389
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<211> 463
<212> PRT
<213> Exophiala spinifera

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Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser  
35 40 45  
Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn  
50 55 60  
Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu  
65 70 75 80  
Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln  
85 90 95  
Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu  
100 105 110  
Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu  
115 120 125  
Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys  
130 135 140  
Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn  
145 150 155 160  
Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu  
165 170 175  
Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile  
180 185 190  
Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly  
195 200 205  
Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala  
210 215 220  
Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val  
225 230 235 240  
Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser  
245 250 255  
Gly Ala Val Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr  
260 265 270  
Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln  
275 280 285  
Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe  
290 295 300  
Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu  
305 310 315 320  
Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp  
325 330 335  
Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly  
340 345 350  
Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp  
355 360 365  
Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu  
370 375 380  
Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln  
385 390 395 400  
Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly  
405 410 415  
Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu  
420 425 430  
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435 440 445  
Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala  
450 455 460

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-85	-80	-75	
gca tta gct gct cca gtc aac act aca aca gaa gat gaa acg gca caa Ala Leu Ala Ala Pro Val Asn Thr Thr Glu Asp Glu Thr Ala Gln			96
-70	-65	-60	
att ccg gct gaa gct gtc atc ggt tac tca gat tta gaa ggg gat ttc Ile Pro Ala Glu Ala Val Ile Gly Tyr Ser Asp Leu Glu Gly Asp Phe			144
-55	-50	-45	
gat gtt gct gtt ttg cca ttt tcc aac agc aca aat aac ggg tta ttg Asp Val Ala Val Leu Pro Phe Ser Asn Ser Thr Asn Asn Gly Leu Leu			192
-40	-35	-30	
ttt ata aat act act att gcc agc att gct gct aaa gaa gaa ggg gta Phe Ile Asn Thr Thr Ile Ala Ser Ile Ala Ala Lys Glu Glu Gly Val			240
-25	-20	-15	-10
tct ctc gag aaa aga gag gct gaa gct gaa ttc aaa gac aac gtt gcg Ser Leu Glu Lys Arg Glu Ala Glu Ala Glu Phe Lys Asp Asn Val Ala			288
-5	1	5	
gac gtg gta gtg gtc ggc gct ggc ttg agc ggt ttg gag acg gca cgc Asp Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg			336
10	15	20	
aaa gtc cag gcc gcc ggt ctg tcc tgc ctc gtt ctt gag gcg atg gat Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp			384
25	30	35	
cgt gta ggg gga aag act ctg agc gta caa tcg ggt ccc ggc agg acg Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr			432
40	45	50	55
act atc aac gac ctc ggc gct gcg tgg atc aat gac agc aac caa agc Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser			480
60	65	70	
gaa gta tcc aga ttg ttt gaa aga ttt cat ttg gag ggc gag ctc cag Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln			528
75	80	85	
agg acg act gga aat tca atc cat caa gca caa gac ggt aca acc act Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr			576
90	95	100	
aca gct cct tat ggt gac tcc ttg ctg agc gag gag gtt gca agt gca Thr Ala Pro Tyr Gly Asp Ser Leu Ser Glu Glu Val Ala Ser Ala			624
105	110	115	
ctt gcg gaa ctc ctc ccc gta tgg tct cag ctg atc gaa gag cat agc Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser			672
120	125	130	135
ctt caa gac ctc aag gcg agc cct cag gcg aag cgg ctc gac agt gtg Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val			720
140	145	150	
agc ttc gcg cac tac tgt gag aag gaa cta aac ttg cct gct gtt ctc Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu			768
155	160	165	

ggc gta gca aac cag atc aca cgc gct ctg ctc ggt gtg gaa gcc cac Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His 170 175 180	816
gag atc agc atg ctt ttt ctc acc gac tac atc aag agt gcc acc ggt Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly 185 190 195	864
ctc agt aat att ttc tcg gac aag aaa gac ggc ggg cag tat atg cga Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg 200 205 210 215	912
tgc aaa aca ggt atg cag tcg att tgc cat gcc atg tca aag gaa ctt Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu 220 225 230	960
gtt cca ggc tca gtg cac ctc aac acc ccc gtc gct gaa att gag cag Val Pro Gly Ser Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln 235 240 245	1008
tcg gca tcc ggc tgt aca gta cga tcg gcc tcg ggc gcc gtg ttc cga Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg 250 255 260	1056
agc aaa aag gtg gtg gtt tcg tta ccg aca acc ttg tat ccc acc ttg Ser Lys Lys Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu 265 270 275	1104
aca ttt tca cca cct ctt ccc gcc gag aag caa gca ttg gcg gaa aat Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn 280 285 290 295	1152
tct atc ctg ggc tac tat agc aag ata gtc ttc gta tgg gac aag ccg Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro 300 305 310	1200
tgg tgg cgc gaa caa ggc ttc tcg ggc gtc ctc caa tcg agc tgt gac Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp 315 320 325	1248
ccc atc tca ttt gcc aga gat acc agc atc gac gtc gat cga caa tgg Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp 330 335 340	1296
tcc att acc tgt ttc atg gtc gga gac ccg gga cgg aag tgg tcc caa Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln 345 350 355	1344
cag tcc aag cag gta cga caa aag tct gtc tgg gac caa ctc cgc gca Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala 360 365 370 375	1392
gcc tac gag aac gcc ggg gcc caa gtc cca gag ccg gcc aac gtg ctc Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu 380 385 390	1440
gaa atc gag tgg tcg aag cag cag tat ttc caa gga gct ccg agc gcc Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala 395 400 405	1488
gtc tat ggg ctg aac gat ctc atc aca ctg ggt tcg gcg ctc aga acg	1536

Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr			
410	415	420	
ccg ttc aag agt gtt cat ttc gtt gga acg gag acg tct tta gtt tgg			1584
Pro Phe Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp			
425	430	435	
aaa ggg tat atg gaa ggg gcc ata cga tcg ggt caa cga ggt gct gca			1632
Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala			
440	445	450	455
gaa gtt gtg gct agc ctg gtg cca gca gca taggcggccg c			1673
Glu Val Val Ala Ser Leu Val Pro Ala Ala			
460	465		
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Ile Pro Ala Glu Ala Val Ile Gly Tyr Ser Asp Leu Glu Gly Asp Phe			
-55	-50	-45	
Asp Val Ala Val Leu Pro Phe Ser Asn Ser Thr Asn Asn Gly Leu Leu			
-40	-35	-30	
Phe Ile Asn Thr Thr Ile Ala Ser Ile Ala Ala Lys Glu Glu Gly Val			
-25	-20	-15	-10
Ser Leu Glu Lys Arg Glu Ala Glu Ala Glu Phe Lys Asp Asn Val Ala			
-5	1	5	
Asp Val Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg			
10	15	20	
Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp			
25	30	35	
Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr			
40	45	50	55
Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser			
60	65	70	
Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln			
75	80	85	
Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr			
90	95	100	
Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala			
105	110	115	
Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser			
120	125	130	135
Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val			
140	145	150	
Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu			
155	160	165	
Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His			
170	175	180	
Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly			
185	190	195	

Leu Ser Asn Ile Phe Ser Asp Lys Asp Gly Gly Gln Tyr Met Arg  
 200 205 210 215  
 Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu  
 220 225 230  
 Val Pro Gly Ser Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln  
 235 240 245  
 Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg  
 250 255 260  
 Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu  
 265 270 275  
 Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn  
 280 285 290 295  
 Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro  
 300 305 310  
 Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp  
 315 320 325  
 Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp  
 330 335 340  
 Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln  
 345 350 355  
 Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala  
 360 365 370 375  
 Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu  
 380 385 390  
 Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala  
 395 400 405  
 Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr  
 410 415 420  
 Pro Phe Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp  
 425 430 435  
 Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala  
 440 445 450 455  
 Glu Val Val Ala Ser Leu Val Pro Ala Ala  
 460 465

<210> 18  
 <211> 2079  
 <212> DNA  
 <213> Unknown

<220>  
 <223> GST:K:trAPAO 2079 nt. Translation starting at nt 1  
 - 687, gst fusion + polylinker, 688-2076,  
 K:trAPAO, extra lysine underlined; 2077-2079, stop  
 codon. For bacterial expression.

<221> CDS  
 <222> (1)...(2076)

<221> misc\_feature  
 <222> (1)...(687)  
 <223> gst fusion + polylinker

<221> misc\_feature  
 <222> (688)...(2076)  
 <223> K:trAPAO

<221> misc\_feature  
 <222> (688)...(690)  
 <223> Extra lysine

DRAFT

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1 5 10 15	
act cga ctt ctt ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ttg Thr Arg Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu	96
20 25 30	
tat gag cgc gat gaa ggt gat aaa tgg cga aac aaa aag ttt gaa ttg Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu	144
35 40 45	
ggt ttg gag ttt ccc aat ctt cct tat tat att gat ggt gat gtt aaa Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys	192
50 55 60	
tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn	240
65 70 75 80	
atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu	288
85 90 95	
gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser	336
100 105 110	
aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu	384
115 120 125	
atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa aca tat tta aat Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn	432
130 135 140	
ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp	480
145 150 155 160	
gtt gtt tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu	528
165 170 175	
gtt tgt ttt aaa aaa cgt att gaa gct atc cca caa att gat aag tac Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr	576
180 185 190	
ttg aaa tcc agc aag tat ata gca tgg cct ttg cag ggc tgg caa gcc Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala	624
195 200 205	
acg ttt ggt ggt ggc gac cat cct cca aaa tcg gat ctg gtt cog cgt Thr Phe Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg	672
210 215 220	
gga tcc ccg gaa ttc aaa gac aac gtt gcg gac gtg gta gtg gtc ggc Gly Ser Pro Glu Phe Lys Asp Asn Val Ala Asp Val Val Val Gly	720
225 230 235 240	
gct ggc ttg agc ggt ttg gag acg gca cgc aaa gtc cag gcc gcc ggt	768

THE HISTORY OF THE CHURCH OF JESUS CHRIST

DRAFT

tcg tta ccg aca acc ttg tat ccc acc ttg aca ttt tca cca cct ctt Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu	500	505	510	1536
ccc gcc gag aag caa gca ttg gcg gaa aat tct atc ctg ggc tac tat Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr	515	520	525	1584
agc aag ata gtc ttc gta tgg gac aag ccg tgg tgg cgc gaa caa ggc Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Arg Glu Gln Gly	530	535	540	1632
ttc tcg ggc gtc ctc caa tcg agc tgt gac ccc atc tca ttt gcc aga Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg	545	550	555	1680
gat acc agc atc gac gtc gat cga caa tgg tcc att acc tgt ttc atg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met	565	570	575	1728
gtc gga gac ccg gga cgg aag tgg tcc caa cag tcc aag cag gta cga Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg	580	585	590	1776
caa aag tct gtc tgg gac caa ctc cgc gca gcc tac gag aac gcc ggg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly	595	600	605	1824
gcc caa gtc cca gag ccg gcc aac gtg ctc gaa atc gag tgg tcg aag Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys	610	615	620	1872
cag cag tat ttc caa gga gct ccg agc gcc gtc tat ggg ctg aac gat Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp	625	630	635	1920
ctc atc aca ctg ggt tcg gcg ctc aga acg ccg ttc aag agt gtt cat Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His	645	650	655	1968
ttc gtt gga acg gag acg tct tta gtt tgg aaa ggg tat atg gaa ggg Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly	660	665	670	2016
gcc ata cga tcg ggt caa cga ggt gct gca gaa gtt gtg gct agc ctg Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu	675	680	685	2064
gtg cca gca gca tag Val Pro Ala Ala				2079
690				

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<210> 19
<211> 692
<212> PRT
<213> Unknown

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<400> 19
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1 5 10 15

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Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu  
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 Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu  
                   35                  40                  45  
 Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys  
                   50                  55                  60  
 Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn  
                   65                  70                  75                  80  
 Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu  
                   85                  90                  95  
 Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser  
                   100                105                110  
 Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu  
                   115                120                125  
 Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn  
                   130                135                140  
 Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp  
                   145                150                155                160  
 Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu  
                   165                170                175  
 Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr  
                   180                185                190  
 Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala  
                   195                200                205  
 Thr Phe Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg  
                   210                215                220  
 Gly Ser Pro Glu Phe Lys Asp Asn Val Ala Asp Val Val Val Gly  
                   225                230                235                240  
 Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly  
                   245                250                255  
 Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr  
                   260                265                270  
 Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly  
                   275                280                285  
 Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe  
                   290                295                300  
 Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser  
                   305                310                315                320  
 Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp  
                   325                330                335  
 Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro  
                   340                345                350  
 Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala  
                   355                360                365  
 Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys  
                   370                375                380  
 Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile  
                   385                390                395                400  
 Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met Leu Phe  
                   405                410                415  
 Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser  
                   420                425                430  
 Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln  
                   435                440                445  
 Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro Gly Ser Val His  
                   450                455                460  
 Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr  
                   465                470                475                480  
 Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys Lys Val Val Val  
                   485                490                495  
 Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu  
                   500                505                510

Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr  
       515                         520                         525  
 Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly  
       530                         535                         540  
 Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg  
       545                         550                         555                         560  
 Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met  
       565                         570                         575  
 Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg  
       580                         585                         590  
 Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly  
       595                         600                         605  
 Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys  
       610                         615                         620  
 Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp  
       625                         630                         635                         640  
 Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His  
       645                         650                         655  
 Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly  
       660                         665                         670  
 Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu  
       675                         680                         685  
 Val Pro Ala Ala  
       690

<210> 20  
 <211> 1464  
 <212> DNA  
 <213> Unknown

<220>  
 <223> Nucleotide sequence of K:trAPAO translational fusion with barley alpha amylase signal sequence, for expression and secretion of the mature trAPAO in maize. Nucleotides 1-72, barley alpha amylase signal sequence, nucleotides 73-75, added lysine residue; nucleotides 76 -1464 , trAPAO cDNA.

<221> sig\_peptide  
 <222> (1)...(72)  
 <223> Barley alpha amylase signal sequence

<221> misc\_feature  
 <222> (73)...(1464)  
 <223> K:trAPAOcDNA

<221> CDS  
 <222> (1)...(1461)

<221> misc\_feature  
 <222> (73)...(75)  
 <223> Added lysine residue

<400> 20

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 Met Ala Asn Lys His Leu Ser Leu Ser Leu Phe Leu Val Leu Leu Gly  
       -20                         -15                         -10

48

ctc tcc gcc tcc ctc gcc agc ggc aaa gac aac gtt gcg gac gtg gta  
 Leu Ser Ala Ser Leu Ala Ser Gly Lys Asp Asn Val Ala Asp Val Val  
       -5                         1                         5

96

gtg	gtg	ggc	gct	ggc	ttg	agc	ggt	ttg	gag	acg	gca	cgc	aaa	gtc	cag		144
Val	Val	Gly	Ala	Gly	Leu	Ser	Gly	Leu	Glu	Thr	Ala	Arg	Lys	Val	Gln		
10					15					20							
gcc	gcc	ggt	ctg	tcc	tgc	ctc	gtt	ctt	gag	gcg	atg	gat	cgt	gta	ggg		192
Ala	Ala	Gly	Leu	Ser	Cys	Leu	Val	Leu	Glu	Ala	Met	Asp	Arg	Val	Gly		
25					30				35				40				
gga	aag	act	ctg	agc	gta	caa	tcg	ggt	ccc	ggc	agg	acg	act	atc	aac		240
Gly	Lys	Thr	Leu	Ser	Val	Gln	Ser	Gly	Pro	Gly	Arg	Thr	Thr	Ile	Asn		
					45				50				55				
gac	ctc	ggc	gct	gcg	tgg	atc	aat	gac	agc	aac	caa	agc	gaa	gta	tcc		288
Asp	Leu	Gly	Ala	Ala	Trp	Ile	Asn	Asp	Ser	Asn	Gln	Ser	Glu	Val	Ser		
					60				65				70				
aga	ttg	ttt	gaa	aga	ttt	cat	ttg	gag	ggc	gag	ctc	cag	agg	acg	act		336
Arg	Leu	Phe	Glu	Arg	Phe	His	Leu	Glu	Gly	Glu	Leu	Gln	Arg	Thr	Thr		
					75				80				85				
gga	aat	tca	atc	cat	caa	gca	caa	gac	ggt	aca	acc	act	aca	gct	cct		384
Gly	Asn	Ser	Ile	His	Gln	Ala	Gln	Asp	Gly	Thr	Thr	Thr	Thr	Ala	Pro		
					90				95				100				
tat	ggt	gac	tcc	ttg	ctg	agc	gag	gag	gtt	gca	agt	gca	ctt	gcg	gaa		432
Tyr	Gly	Asp	Ser	Leu	Leu	Ser	Glu	Glu	Val	Ala	Ser	Ala	Leu	Ala	Glu		
					105				110				115			120	
ctc	ctc	ccc	gta	tgg	tct	cag	ctg	atc	gaa	gag	cat	agc	ctt	caa	gac		480
Leu	Leu	Pro	Val	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Gln	Asp		
					125				130				135				
ctc	aag	gcg	agc	cct	cag	gcg	aag	cg	ctc	gac	agt	gtg	agc	ttc	g		528
Leu	Lys	Ala	Ser	Pro	Gln	Ala	Lys	Arg	Leu	Asp	Ser	Val	Ser	Phe	Ala		
					140				145				150				
cac	tac	tgt	gag	aag	gaa	cta	aac	ttg	cct	gct	gtt	ctc	ggc	gta	gca		576
His	Tyr	Cys	Glu	Lys	Glu	Leu	Asn	Leu	Pro	Ala	Val	Leu	Gly	Val	Ala		
					155				160				165				
aac	cag	atc	aca	cgc	gct	ctg	ctc	ggt	gtg	gaa	gcc	cac	gag	atc	agc		624
Asn	Gln	Ile	Thr	Arg	Ala	Leu	Leu	Gly	Val	Ala	His	Glu	Ile	Ser			
					170				175				180				
atg	ctt	ttt	ctc	acc	gac	tac	atc	aag	agt	gcc	acc	ggt	ctc	agt	aat		672
Met	Leu	Phe	Leu	Thr	Asp	Tyr	Ile	Lys	Ser	Ala	Thr	Gly	Leu	Ser	Asn		
					185				190				195			200	
att	tcc	tcg	gac	aag	aaa	gac	ggc	ggg	cag	tat	atg	cga	tgc	aaa	aca		720
Ile	Phe	Ser	Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys	Thr		
					205				210				215				
ggt	atg	cag	tcg	att	tgc	cat	gcc	atg	tca	aag	gaa	ctt	gtt	cca	ggc		768
Gly	Met	Gln	Ser	Ile	Cys	His	Ala	Met	Ser	Lys	Glu	Leu	Val	Pro	Gly		
					220				225				230				
tca	gtg	cac	ctc	aac	acc	ccc	gtc	gct	gaa	att	gag	cag	tcg	gca	tcc		816
Ser	Val	His	Leu	Asn	Thr	Pro	Val	Ala	Glu	Ile	Glu	Gln	Ser	Ala	Ser		
					235				240				245				
ggc	tgt	aca	gta	cga	tcg	gcc	tcg	ggc	gcc	gtg	ttc	cga	agc	aaa	aag		864

Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys Lys			
250	255	260	
gtg gtg gtt tcg tta ccg aca acc ttg tat ccc acc ttg aca ttt tca			912
Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe Ser			
265	270	275	280
cca cct ctt ccc gcc gag aag caa gca ttg gcg gaa aat tct atc ctg			960
Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile Leu			
285	290	295	
ggc tac tat agc aag ata gtc ttc gta tgg gac aag ccg tgg tgg cgc			1008
Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp Arg			
300	305	310	
gaa caa ggc ttc tcg ggc gtc ctc caa tcg agc tgt gac ccc atc tca			1056
Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser			
315	320	325	
ttt gcc aga gat acc agc atc gac gtc gat cga caa tgg tcc att acc			1104
Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile Thr			
330	335	340	
tgt ttc atg gtc gga gac ccg gga aag tgg tcc caa cag tcc aag			1152
Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys			
345	350	355	360
cag gta cga caa aag tct gtc tgg gac caa ctc cgc gca gcc tac gag			1200
Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr Glu			
365	370	375	
aac gcc ggg gcc caa gtc cca gag ccg gcc aac gtg ctc gaa atc gag			1248
Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu			
380	385	390	
tgg tcg aag cag cag tat ttc caa gga gct ccg agc gcc gtc tat ggg			1296
Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr Gly			
395	400	405	
ctg aac gat ctc atc aca ctg ggt tcg gcg ctc aga acg ccg ttc aag			1344
Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys			
410	415	420	
agt gtt cat ttc gtt gga acg gag acg tct tta gtt tgg aaa ggg tat			1392
Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr			
425	430	435	440
atg gaa ggg gcc ata cga tcg ggt caa cga ggt gct gca gaa gtt gtg			1440
Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val Val			
445	450	455	
gct agc ctg gtg cca gca gca tag			1464
Ala Ser Leu Val Pro Ala Ala			
460			

<210> 21  
 <211> 487  
 <212> PRT  
 <213> Unknown

<220>  
<221> SIGNAL  
<222> (1)...(24)

<400> 21  
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Leu Ser Ala Ser Leu Ala Ser Gly Lys Asp Asn Val Ala Asp Val Val  
-5 1 5  
Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val Gln  
10 15 20  
Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val Gly  
25 30 35 40  
Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile Asn  
45 50 55  
Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Ser  
60 65 70  
Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr Thr  
75 80 85  
Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro  
90 95 100  
Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu  
105 110 115 120  
Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln Asp  
125 130 135  
Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe Ala  
140 145 150  
His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val Ala  
155 160 165  
Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser  
170 175 180  
Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn  
185 190 195 200  
Ile Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr  
205 210 215  
Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro Gly  
220 225 230  
Ser Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser  
235 240 245  
Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys Lys  
250 255 260  
Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe Ser  
265 270 275 280  
Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile Leu  
285 290 295  
Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp Arg  
300 305 310  
Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser  
315 320 325  
Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile Thr  
330 335 340  
Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys  
345 350 355 360  
Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr Glu  
365 370 375  
Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu  
380 385 390  
Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr Gly  
395 400 405  
Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys  
410 415 420  
Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr

425                          430                          435                          440  
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     1                        5                            10                            15

gca ggg tat tct cac gtc ggc gta ggc cca gac gga ggg agg tat gtg                  96  
 Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Arg Tyr Val  
     20                        25                            30

aca ata gct gga cag att gga caa gac gct tcg ggc gtg aca gac cct                  144  
 Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro  
     35                        40                            45

gcc tac gag aaa cag gtt gcc caa gca ttc gcc aat ctg cga gct tgc                  192  
 Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys  
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 Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr  
     65                        70                            75                            80

tac atc gtc gac tac gcc ccg agc aaa ctc acc gca att gga gat ggg                  288  
 Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly  
     85                        90                            95

ctg aag gct acc ttt gcc ctt gac agg ctc cct tgc acg ctg gtg                  336  
 Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val  
     100                        105                            110

cca gtg tcg gcc ttg tct tca cct gaa tac ctc ttt gag gtt gat gcc                  384  
 Pro Val Ser Ala Leu Ser Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala  
     115                        120                            125

acg gcg ctg gtg ccg gga cac acg acc cca gac aac gtt gcg gac gtg                  432  
 Thr Ala Leu Val Pro Gly His Thr Thr Pro Asp Asn Val Ala Asp Val  
     130                        135                            140

gta gtg gtg ggc gct ggc ttg agc ggt ttg gag acg gca cgc aaa gtc                  480  
 Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val  
     145                        150                            155                            160

cag gcc gcc ggt ctg tcc tgc ctc gtt ctt gag gcg atg gat cgt gta                  528  
 Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val  
     165                        170                            175

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act gga aat tca atc cat caa gca caa gac ggt aca acc act aca gct Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala 225	230	235	720
cct tat ggt gac tcc ttg ctg agc gag gag gtt gca agt gca ctt gcg Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala 245	250	255	768
gaa ctc ctc ccc gta tgg tct cag ctg atc gaa gag cat agc ctt caa Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln 260	265	270	816
gac ctc aag gcg agc cct cag gcg aag cgg ctc gac agt gtg agc ttc Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe 275	280	285	864
gcg cac tac tgt gag aag gaa cta aac ttg cct gct gtt ctc ggc gta Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val 290	295	300	912
gca aac cag atc aca cgc gct ctg ctc ggt gtg gaa gcc cac gag atc Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile 305	310	315	960
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tcc ggc tgt aca gta cga tcg gcc tcg ggc gcc gtg ttc cga agc aaa Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys 385	390	395	1200
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ctg ggc tac tat agc aag ata gtc ttc gta tgg gac aag ccg tgg tgg Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp 435 440 445	1344																							
cgc gaa caa ggc ttc tcg ggc gtc ctc caa tcg agc tgt gac ccc atc Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile 450 455 460	1392																							
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gtg gct agc ctg gtg cca gca gca tag Val Ala Ser Leu Val Pro Ala Ala 595 600	1803																							
<p>&lt;210&gt; 23  &lt;211&gt; 600  &lt;212&gt; PRT  &lt;213&gt; <i>Exophiala spinifera</i></p> <p>&lt;400&gt; 23</p> <table border="0"> <tbody> <tr> <td>Met Ala Leu Ala Pro Ser Tyr Ile Asn Pro Pro Asn Val Ala Ser Pro</td> <td></td> </tr> <tr> <td>1 5 10 15</td> <td></td> </tr> <tr> <td>Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Gly Arg Tyr Val</td> <td></td> </tr> <tr> <td>20 25 30</td> <td></td> </tr> <tr> <td>Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro</td> <td></td> </tr> <tr> <td>35 40 45</td> <td></td> </tr> <tr> <td>Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys</td> <td></td> </tr> <tr> <td>50 55 60</td> <td></td> </tr> <tr> <td>Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr</td> <td></td> </tr> <tr> <td>65 70 75 80</td> <td></td> </tr> <tr> <td>Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly</td> <td></td> </tr> </tbody> </table>		Met Ala Leu Ala Pro Ser Tyr Ile Asn Pro Pro Asn Val Ala Ser Pro		1 5 10 15		Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Gly Arg Tyr Val		20 25 30		Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro		35 40 45		Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys		50 55 60		Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr		65 70 75 80		Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly		1803
Met Ala Leu Ala Pro Ser Tyr Ile Asn Pro Pro Asn Val Ala Ser Pro																								
1 5 10 15																								
Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Gly Arg Tyr Val																								
20 25 30																								
Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro																								
35 40 45																								
Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys																								
50 55 60																								
Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr																								
65 70 75 80																								
Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly																								

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Leu Lys Ala Thr Phe Ala Leu Asp Arg	Leu Pro Pro Cys	Thr Leu Val
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Ser Arg Leu Phe Glu Arg	Phe His Leu Glu Gly Glu Leu Gln Arg Thr	
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Thr Gly Asn Ser Ile His	Gln Ala Gln Asp Gly Thr Thr Thr Ala	
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Pro Tyr Gly Asp Ser	Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala	
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Thr Cys Phe Met Val Gly Asp Pro	Gly Arg Lys Trp Ser Gln Gln Ser	
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Gly Leu Asn Asp Leu Ile Thr	Leu Gly Ser Ala Leu Arg Thr Pro Phe	
545	550	560
Lys Ser Val His Phe Val Gly	Thr Glu Thr Ser Leu Val Trp Lys Gly	
565	570	575
Tyr Met Glu Gly Ala Ile Arg Ser	Gly Gln Arg Gly Ala Ala Glu Val	



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ccc aag gga gat ttg tcc gaa cct tgg aac acc act gtt caa gct ctc Pro Lys Gly Asp Leu Ser Glu Pro Trp Asn Thr Thr Val Gln Ala Leu	235                    240                    245	816
aac tgt acc acc agt atc gac atc ttg agt tgt atg aga aga gtc gat Asn Cys Thr Thr Ser Ile Asp Ile Leu Ser Cys Met Arg Arg Val Asp	250                    255                    260	864
ctc gcc act ctg atg aac acg atc gag caa ctc gga ctt ggg ttt gag Leu Ala Thr Leu Met Asn Thr Ile Glu Gln Leu Gly Leu Gly Phe Glu	265                    270                    275                    280	912
tac acg ttg gac aac gta acg gct gtg tac cgt tct gaa acg gct cgc Tyr Thr Leu Asp Asn Val Thr Ala Val Tyr Arg Ser Glu Thr Ala Arg	285                    290                    295	960
acg act ggt gac att gct cgt gta cct gtt ctc gtc ggg acg gtg gcc Thr Thr Gly Asp Ile Ala Arg Val Pro Val Leu Val Gly Thr Val Ala	300                    305                    310	1008
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Tyr His Ser Ser Glu Val Gly Met Val Phe Gly Thr Tyr Pro Val Ala			
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795	800	805

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Arg Glu Ile Thr Met Ala Trp Phe Asn Thr Pro Pro Pro Ser Ala Gly			
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Glu Ser Glu Asp Cys Leu Asn Leu Asn Ile Tyr Val Pro Gly Thr Glu			
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Asn Thr Asn Lys Ala Val Met Val Trp Ile Tyr Gly Gly Ala Leu Glu			
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Tyr Gly Trp Asn Ser Phe His Leu Tyr Asp Gly Ala Ser Phe Ala Ala			
125	130	135	
Asn Gln Asp Val Ile Ala Val Thr Ile Asn Tyr Arg Thr Asn Ile Leu			
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Gly Phe Pro Ala Ala Pro Gln Leu Pro Ile Thr Gln Arg Asn Leu Gly			
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Phe Leu Asp Gln Arg Phe Ala Leu Asp Trp Val Gln Arg Asn Ile Ala			
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Ala Phe Gly Gly Asp Pro Arg Lys Val Thr Ile Phe Gly Gln Ser Ala			
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Gly Gly Arg Ser Val Asp Val Leu Leu Thr Ser Met Pro His Asn Pro			
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Pro Phe Arg Ala Ala Ile Met Glu Ser Gly Val Ala Asn Tyr Asn Phe			
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Pro Lys Gly Asp Leu Ser Glu Pro Trp Asn Thr Thr Val Gln Ala Leu			
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Leu Ala Thr Leu Met Asn Thr Ile Glu Gln Leu Gly Leu Gly Phe Glu			
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Tyr Thr Leu Asp Asn Val Thr Ala Val Tyr Arg Ser Glu Thr Ala Arg			
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Thr Thr Gly Asp Ile Ala Arg Val Pro Val Leu Val Gly Thr Val Ala			
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Asn Asp Gly Leu Leu Phe Val Leu Gly Glu Asn Asp Thr Gln Ala Tyr			
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Trp Ala Ala Phe Ala Lys Asn Pro Met Asn Gly Pro Gly Trp Lys Gln			
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Val Pro Asn Val Ala Ala Leu Gly Ser Pro Gly Lys Ala Ile Gln Val			
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Tyr Tyr Thr Glu Leu Gly Thr Ile Ala Pro Arg Thr Phe Gly Gly Gly			
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Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala	Val Phe Arg Ser Lys	
765	770	775
Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr	Pro Thr Leu Thr Phe	
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Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu	Ala Glu Asn Ser Ile	
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Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp	Asp Lys Pro Trp Trp	
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Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser	Ser Cys Asp Pro Ile	
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Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp	Arg Gln Trp Ser Ile	
845	850	855
Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys	Trp Ser Gln Gln Ser	
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Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln	Leu Arg Ala Ala Tyr	
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Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala	Asn Val Leu Glu Ile	
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Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly	Ala Pro Ser Ala Val Tyr	
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Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala	Leu Arg Thr Pro Phe	
925	930	935
Lys Ser Val His Phe Val Gly Thr Glu Thr Ser	Leu Val Trp Lys Gly	
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          10                  15                  20

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Gly Ile Pro Tyr Ala Ala Pro Pro Val Gly Gly Leu Arg Trp Lys Pro
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Pro Gln His Ala Arg Pro Trp Ala Gly Val Arg Pro Ala Thr Gln Phe
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Gly Ser Asp Cys Phe Gly Ala Ala Tyr Leu Arg Lys Gly Ser Leu Ala
          60                  65                  70

ccc ggc gtg agc gag gac tgt ctt tac ctc aac gta tgg gcg ccg tca 336
Pro Gly Val Ser Glu Asp Cys Leu Tyr Leu Asn Val Trp Ala Pro Ser
          75                  80                  85

ggc gct aaa ccc ggc cag tac ccc gtc atg gtc tgg gtc tac ggc ggc 384
Gly Ala Lys Pro Gly Gln Tyr Pro Val Met Val Trp Val Tyr Gly Gly

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atc ctg ggc ttt ttc gcc cat cct ggt ctc tcg cgc gag agc ccc acc Ile Leu Gly Phe Phe Ala His Pro Gly Leu Ser Arg Glu Ser Pro Thr	140	145	528
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gga act tcg ggc aac tac ggc cta ctc gac att ctc gcc gct ctt cgg Gly Thr Ser Gly Asn Tyr Gly Leu Leu Asp Ile Leu Ala Ala Leu Arg	155	160	576
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tgg gtg cag agc aac gcc cgc gcc ttc gga ggg gac ccc ggc cga gtg Trp Val Gln Ser Asn Ala Arg Ala Phe Gly Gly Asp Pro Gly Arg Val	170	175	624
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acg gtc ttt ggt gaa tcg gcc gga gcg agc gcg atc gga ctt ctg ctc Thr Val Phe Gly Glu Ser Ala Gly Ala Ser Ala Ile Gly Leu Leu Leu	185	190	672
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275		280	
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735				740
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 Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn  
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 Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu  
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 Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly  
                   940                  945                  950  
 Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala  
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<211> 3618

<212> DNA

<213> Unknown

<220>

<223> gst:esp1:sp:K:trapao, 3618. 1-687, gst +  
polylinker; 688-2190, esp1 mat; 2191-2226 spacer;  
2227-3615, K:trAPAO, 3616-3618, stop codon. For  
bacterial expression.

<221> CDS

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<221> misc\_feature

<222> (1)...(687)

<223> gst + polylinker

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<223> Extra lysine

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Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro	
1 5 10 15	

act cga ctt ctt ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ttg	96
Thr Arg Leu Leu Glu Tyr Leu Glu Lys Tyr Glu Glu His Leu	
20 25 30	

tat gag cgc gat gaa ggt gat aaa tgg cga aac aaa aag ttt gaa ttg	144
Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu	
35 40 45	

ggt ttg gag ttt ccc aat ctt cct tat tat att gat ggt gat gtt aaa	192
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys	
50 55 60	

tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac	240
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn	
65 70 75 80	

atg ttg ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa	288
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu	
85 90 95	

gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt	336
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser	
100 105 110	

aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa	384
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu	
115 120 125	

atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa aca tat tta aat	432
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn	
130 135 140	

ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat	480
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp	
145 150 155 160	

gtt gtt tta tac atg gac cca atg tgc ctg gat gcg ttc cca aaa tta	528
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu	
165 170 175	

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gtt tgc ttt aaa aaa cgt att gaa gct atc cca caa att gat aag tac		576
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr		
180	185	190
ttg aaa tcc agc aag tat ata gca tgg cct ttg cag ggc tgg caa gcc		624
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala		
195	200	205
acg ttt ggt ggt ggc gac cat cct cca aaa tcg gat ctg gtt ccg cgt		672
Thr Phe Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg		
210	215	220
gga tcc ccg gaa ttc gct cct act gtc aag att gat gct ggg atg gtg		720
Gly Ser Pro Glu Phe Ala Pro Thr Val Lys Ile Asp Ala Gly Met Val		
225	230	235
240		
gtc ggc acg act act gtc ccc ggc acc act gcg acc gtc agc gag		768
Val Gly Thr Thr Thr Val Pro Gly Thr Thr Ala Thr Val Ser Glu		
245	250	255
ttc ttg ggc gtt cct ttt gcc gcc tct ccg aca cga ttt gcg cct cct		816
Phe Leu Gly Val Pro Phe Ala Ala Ser Pro Thr Arg Phe Ala Pro Pro		
260	265	270
act cgt ccc gtg cct tgg tca acg cct ttg caa gcc act gca tat ggt		864
Thr Arg Pro Val Pro Trp Ser Thr Pro Leu Gln Ala Thr Ala Tyr Gly		
275	280	285
cca gca tgc cct caa caa ttc aat tac ccc gaa gaa ctc cgt gag att		912
Pro Ala Cys Pro Gln Gln Phe Asn Tyr Pro Glu Glu Leu Arg Glu Ile		
290	295	300
acg atg gcc tgg ttc aat aca ccc ccc ccg tca gct ggt gaa agt gag		960
Thr Met Ala Trp Phe Asn Thr Pro Pro Pro Ser Ala Gly Glu Ser Glu		
305	310	315
320		
gac tgc ctg aac ctc aac atc tac gtc cca gga act gag aac aca aac		1008
Asp Cys Leu Asn Leu Asn Ile Tyr Val Pro Gly Thr Glu Asn Thr Asn		
325	330	335
aaa gcc gtc atg gtt tgg ata tac ggt gga gcg ctg gaa tat ggt tgg		1056
Lys Ala Val Met Val Trp Ile Tyr Gly Gly Ala Leu Glu Tyr Gly Trp		
340	345	350
aat tca ttc cac ctt tac gac ggg gct agt ttc gca gcc aat cag gat		1104
Asn Ser Phe His Leu Tyr Asp Gly Ala Ser Phe Ala Ala Asn Gln Asp		
355	360	365
gtc atc gcc gtg acc atc aac tac aga acg aac att ctg ggg ttc cct		1152
Val Ile Ala Val Thr Ile Asn Tyr Arg Thr Asn Ile Leu Gly Phe Pro		
370	375	380
gct gcc cct cag ctt cca ata aca cag cga aat ctg ggg ttc cta gac		1200
Ala Ala Pro Gln Leu Pro Ile Thr Gln Arg Asn Leu Gly Phe Leu Asp		
385	390	395
400		
caa agg ttt gct ttg gat tgg gta cag cgg aac atc gca gcc ttt ggc		1248
Gln Arg Phe Ala Leu Asp Trp Val Gln Arg Asn Ile Ala Ala Phe Gly		
405	410	415
gtt gat cct caa aag gtc aca ata ttt qqq caq aqt qcq qqq qqc aqa		1296

Gly Asp Pro Arg Lys Val Thr Ile Phe Gly Gln Ser Ala Gly Gly Arg			
420	425	430	
agt gtc gac gtc ctc ttg acg tct atg cca cac aac cca ccc ttc cga			1344
Ser Val Asp Val Leu Leu Thr Ser Met Pro His Asn Pro Pro Phe Arg			
435	440	445	
gca gca atc atg gag tcc ggt gtg gct aac tac aac ttc ccc aag gga			1392
Ala Ala Ile Met Glu Ser Gly Val Ala Asn Tyr Asn Phe Pro Lys Gly			
450	455	460	
gat ttg tcc gaa cct tgg aac acc act gtt caa gct ctc aac tgt acc			1440
Asp Leu Ser Glu Pro Trp Asn Thr Thr Val Gln Ala Leu Asn Cys Thr			
465	470	475	480
acc agt atc gac atc ttg agt tgt atg aga aga gtc gat ctc gcc act			1488
Thr Ser Ile Asp Ile Leu Ser Cys Met Arg Arg Val Asp Leu Ala Thr			
485	490	495	
ctg atg aac acg atc gag caa ctc gga ctt ggg ttt gag tac acg ttg			1536
Leu Met Asn Thr Ile Glu Gln Leu Gly Leu Gly Phe Glu Tyr Thr Leu			
500	505	510	
gac aac gta acg gct gtg tac cgt tct gaa acg gct cgc acg act ggt			1584
Asp Asn Val Thr Ala Val Tyr Arg Ser Glu Thr Ala Arg Thr Thr Gly			
515	520	525	
gac att gct cgt gta cct gtt ctc gtc ggg acg gtg gcc aac gac gga			1632
Asp Ile Ala Arg Val Pro Val Leu Val Gly Thr Val Ala Asn Asp Gly			
530	535	540	
ctt ctc ttt gtc ctc ggg gag aat gac acc caa gca tat ctc gag gag			1680
Leu Leu Phe Val Leu Gly Glu Asn Asp Thr Gln Ala Tyr Leu Glu Glu			
545	550	555	560
gca atc ccg aat cag ccc gac ctt tac cag act ctc ctt gga gca tat			1728
Ala Ile Pro Asn Gln Pro Asp Leu Tyr Gln Thr Leu Leu Gly Ala Tyr			
565	570	575	
ccc att gga tcc cca ggg atc gga tcg cct caa gat cag att gcc gcc			1776
Pro Ile Gly Ser Pro Gly Ile Gly Ser Pro Gln Asp Gln Ile Ala Ala			
580	585	590	
att gag acc gag gta aga ttc cag tgt cct tct gcc atc gtg gct cag			1824
Ile Glu Thr Glu Val Arg Phe Gln Cys Pro Ser Ala Ile Val Ala Gln			
595	600	605	
gac tcc cgg aat cgg ggt atc cct tct tgg cgc tac tac tac aat gcg			1872
Asp Ser Arg Asn Arg Gly Ile Pro Ser Trp Arg Tyr Tyr Tyr Asn Ala			
610	615	620	
acc ttt gag aat ctg gag ctt ttc cct ggg tcc gaa gtg tac cac agc			1920
Thr Phe Glu Asn Leu Glu Leu Phe Pro Gly Ser Glu Val Tyr His Ser			
625	630	635	640
tct gaa gtc ggg atg gtg ttt ggc acg tat cct gtc gca agt gcg acc			1968
Ser Glu Val Gly Met Val Phe Gly Thr Tyr Pro Val Ala Ser Ala Thr			
645	650	655	
gcc ttg gag gcc cag acg agc aaa tac atg cag ggt gcc tgg gcg gcc			2016
Ala Leu Glu Ala Gln Thr Ser Lys Tyr Met Gln Gly Ala Trp Ala Ala			
660	665	670	

ttt gcc aaa aac ccc atg aat ggg cct ggg tgg aaa caa gtg ccg aat Phe Ala Lys Asn Pro Met Asn Gly Pro Gly Trp Lys Gln Val Pro Asn 675 680 685	2064
gtc gcg gcg ctt ggc tca cca ggc aaa gcc atc cag gtt gac gtc tct Val Ala Ala Leu Gly Ser Pro Gly Lys Ala Ile Gln Val Asp Val Ser 690 695 700	2112
cca gcg aca ata gac caa cga tgt gcc ttg tac acg cgt tat tat act Pro Ala Thr Ile Asp Gln Arg Cys Ala Leu Tyr Thr Arg Tyr Tyr Thr 705 710 715 720	2160
gag ttg ggc aca atc gcg ccg agg aca ttt ggc gga ggc agc ggc gga Glu Leu Gly Thr Ile Ala Pro Arg Thr Phe Gly Gly Ser Gly Gly 725 730 735	2208
ggc agc ggc gga ggc agc aaa gac aac gtt gcg gac gtg gta gtg gtg Gly Ser Gly Gly Ser Lys Asp Asn Val Ala Asp Val Val Val Val 740 745 750	2256
ggc gct ggc ttg agc ggt ttg gag acg gca cgc aaa gtc cag gcc gcc Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val Gln Ala Ala 755 760 765	2304
ggt ctg tcc tgc ctc gtt ctt gag gcg atg gat cgt gta ggg gga aag Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val Gly Gly Lys 770 775 780	2352
act ctg agc gta caa tcg ggt ccc ggc agg acg act atc aac gac ctc Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu 785 790 795 800	2400
ggc gct gcg tgg atc aat gac agc aac caa agc gaa gta tcc aga ttg Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Ser Arg Leu 805 810 815	2448
ttt gaa aga ttt cat ttg gag ggc gag ctc cag agg acg act gga aat Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn 820 825 830	2496
tca atc cat caa gca caa gac ggt aca acc act aca gct cct tat ggt Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro Tyr Gly 835 840 845	2544
gac tcc ttg ctg agc gag gag gtt gca agt gca ctt gcg gaa ctc ctc Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu Leu Leu 850 855 860	2592
ccc gta tgg tct cag ctg atc gaa gag cat agc ctt caa gac ctc aag Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln Asp Leu Lys 865 870 875 880	2640
gcg agc cct cag gcg aag cgg ctc gac agt gtg agc ttc gcg cac tac Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe Ala His Tyr 885 890 895	2688
tgt gag aag gaa cta aac ttg cct gct gtt ctc ggc gta gca aac cag Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val Ala Asn Gln 900 905 910	2736
atc aca cgc gct ctg ctc ggt gtg gaa gcc cac gag atc agc atg ctt	2784

Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met Leu			
915	920	925	
ttt ctc acc gac tac atc aag agt gcc acc ggt ctc agt aat att ttc			2832
Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe			
930	935	940	
tcg gac aag aaa gac ggc ggg cag tat atg cga tgc aaa aca ggt atg			2880
Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr Gly Met			
945	950	955	960
cag tcg att tgc cat gcc atg tca aag gaa ctt gtt cca ggc tca gtg			2928
Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro Gly Ser Val			
965	970	975	
cac ctc aac acc ccc gtc gct gaa att gag cag tcg gca tcc ggc tgt			2976
His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys			
980	985	990	
aca gta cga tcg gcc tcg ggc gcc gtg ttc cga agc aaa aag gtg gtg			3024
Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys Lys Val Val			
995	1000	1005	
gtt tcg tta ccg aca acc ttg tat ccc acc ttg aca ttt tca cca cct			3072
Val Ser Leu Pro Thr Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro			
1010	1015	1020	
ctt ccc gcc gag aag caa gca ttg gcg gaa aat tct atc ctg ggc tac			3120
Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr			
1025	1030	1035	1040
tat agc aag ata gtc ttc gta tgg gac aag ccg tgg tgg cgc gaa caa			3168
Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp Arg Glu Gln			
1045	1050	1055	
ggc ttc tcg ggc gtc ctc caa tcg agc tgt gac ccc atc tca ttt gcc			3216
Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala			
1060	1065	1070	
aga gat acc agc atc gac gtc gat cga caa tgg tcc att acc tgt ttc			3264
Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile Thr Cys Phe			
1075	1080	1085	
atg gtc gga gac ccg gga cgg aag tgg tcc caa cag tcc aag cag gta			3312
Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys Gln Val			
1090	1095	1100	
cga caa aag tct gtc tgg gac caa ctc cgc gca gcc tac gag aac gcc			3360
Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala			
1105	1110	1115	1120
ggg gcc caa gtc cca gag ccg gcc aac gtg ctc gaa atc gag tgg tcg			3408
Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu Trp Ser			
1125	1130	1135	
aag cag cag tat ttc caa gga gct ccg agc gcc gtc tat ggg ctg aac			3456
Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn			
1140	1145	1150	
gat ctc atc aca ctg ggt tcg gcg ctc aga acg ccg ttc aag agt gtt			3504
Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys Ser Val			
1155	1160	1165	

cat ttc gtt gga acg gag acg tct tta gtt tgg aaa ggg tat atg gaa	3552
His Phe Val Gly Thr Ser Leu Val Trp Lys Gly Tyr Met Glu	
1170 1175 1180	
ggg gcc ata cga tcg ggt caa cga ggt gct gca gaa gtt gtg gct agc	3600
Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val Val Ala Ser	
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ctg gtg cca gca gca tag	3618
Leu Val Pro Ala Ala	
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Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu	
35 40 45	
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys	
50 55 60	
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn	
65 70 75 80	
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu	
85 90 95	
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser	
100 105 110	
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu	
115 120 125	
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn	
130 135 140	
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp	
145 150 155 160	
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu	
165 170 175	
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr	
180 185 190	
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala	
195 200 205	
Thr Phe Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg	
210 215 220	
Gly Ser Pro Glu Phe Ala Pro Thr Val Lys Ile Asp Ala Gly Met Val	
225 230 235 240	
Val Gly Thr Thr Thr Val Pro Gly Thr Thr Ala Thr Val Ser Glu	
245 250 255	
Phe Leu Gly Val Pro Phe Ala Ala Ser Pro Thr Arg Phe Ala Pro Pro	
260 265 270	
Thr Arg Pro Val Pro Trp Ser Thr Pro Leu Gln Ala Thr Ala Tyr Gly	
275 280 285	
Pro Ala Cys Pro Gln Gln Phe Asn Tyr Pro Glu Glu Leu Arg Glu Ile	
290 295 300	
Thr Met Ala Trp Phe Asn Thr Pro Pro Pro Ser Ala Gly Glu Ser Glu	
305 310 315 320	
Asp Cys Leu Asn Leu Asn Ile Tyr Val Pro Gly Thr Glu Asn Thr Asn	
325 330 335	

Lys Ala Val Met Val Trp Ile Tyr Gly Gly Ala Leu Glu Tyr Gly Trp  
                   340                  345                  350  
 Asn Ser Phe His Leu Tyr Asp Gly Ala Ser Phe Ala Ala Asn Gln Asp  
                   355                  360                  365  
 Val Ile Ala Val Thr Ile Asn Tyr Arg Thr Asn Ile Leu Gly Phe Pro  
                   370                  375                  380  
 Ala Ala Pro Gln Leu Pro Ile Thr Gln Arg Asn Leu Gly Phe Leu Asp  
                   385                  390                  395                  400  
 Gln Arg Phe Ala Leu Asp Trp Val Gln Arg Asn Ile Ala Ala Phe Gly  
                   405                  410                  415  
 Gly Asp Pro Arg Lys Val Thr Ile Phe Gly Gln Ser Ala Gly Gly Arg  
                   420                  425                  430  
 Ser Val Asp Val Leu Leu Thr Ser Met Pro His Asn Pro Pro Phe Arg  
                   435                  440                  445  
 Ala Ala Ile Met Glu Ser Gly Val Ala Asn Tyr Asn Phe Pro Lys Gly  
                   450                  455                  460  
 Asp Leu Ser Glu Pro Trp Asn Thr Thr Val Gln Ala Leu Asn Cys Thr  
                   465                  470                  475                  480  
 Thr Ser Ile Asp Ile Leu Ser Cys Met Arg Arg Val Asp Leu Ala Thr  
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 Leu Met Asn Thr Ile Glu Gln Leu Gly Leu Gly Phe Glu Tyr Thr Leu  
                   500                  505                  510  
 Asp Asn Val Thr Ala Val Tyr Arg Ser Glu Thr Ala Arg Thr Thr Gly  
                   515                  520                  525  
 Asp Ile Ala Arg Val Pro Val Leu Val Gly Thr Val Ala Asn Asp Gly  
                   530                  535                  540  
 Leu Leu Phe Val Leu Gly Glu Asn Asp Thr Gln Ala Tyr Leu Glu Glu  
                   545                  550                  555                  560  
 Ala Ile Pro Asn Gln Pro Asp Leu Tyr Gln Thr Leu Leu Gly Ala Tyr  
                   565                  570                  575  
 Pro Ile Gly Ser Pro Gly Ile Gly Ser Pro Gln Asp Gln Ile Ala Ala  
                   580                  585                  590  
 Ile Glu Thr Glu Val Arg Phe Gln Cys Pro Ser Ala Ile Val Ala Gln  
                   595                  600                  605  
 Asp Ser Arg Asn Arg Gly Ile Pro Ser Trp Arg Tyr Tyr Tyr Asn Ala  
                   610                  615                  620  
 Thr Phe Glu Asn Leu Glu Leu Phe Pro Gly Ser Glu Val Tyr His Ser  
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 Ser Glu Val Gly Met Val Phe Gly Thr Tyr Pro Val Ala Ser Ala Thr  
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 Ala Leu Glu Ala Gln Thr Ser Lys Tyr Met Gln Gly Ala Trp Ala Ala  
                   660                  665                  670  
 Phe Ala Lys Asn Pro Met Asn Gly Pro Gly Trp Lys Gln Val Pro Asn  
                   675                  680                  685  
 Val Ala Ala Leu Gly Ser Pro Gly Lys Ala Ile Gln Val Asp Val Ser  
                   690                  695                  700  
 Pro Ala Thr Ile Asp Gln Arg Cys Ala Leu Tyr Thr Arg Tyr Tyr Thr  
                   705                  710                  715                  720  
 Glu Leu Gly Thr Ile Ala Pro Arg Thr Phe Gly Gly Ser Gly Gly  
                   725                  730                  735  
 Gly Ser Gly Gly Ser Lys Asp Asn Val Ala Asp Val Val Val Val  
                   740                  745                  750  
 Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val Gln Ala Ala  
                   755                  760                  765  
 Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val Gly Gly Lys  
                   770                  775                  780  
 Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu  
                   785                  790                  795                  800  
 Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Ser Arg Leu  
                   805                  810                  815  
 Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn  
                   820                  825                  830

Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro Tyr Gly  
       835                  840                  845  
 Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu Leu Leu  
       850                  855                  860  
 Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln Asp Leu Lys  
       865                  870                  875                  880  
 Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe Ala His Tyr  
       885                  890                  895  
 Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val Ala Asn Gln  
       900                  905                  910  
 Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met Leu  
       915                  920                  925  
 Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe  
       930                  935                  940  
 Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr Gly Met  
       945                  950                  955                  960  
 Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro Gly Ser Val  
       965                  970                  975  
 His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys  
       980                  985                  990  
 Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys Lys Val Val  
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 Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro  
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 Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr  
       1025                1030                1035                1040  
 Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp Arg Glu Gln  
       1045                1050                1055  
 Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala  
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 Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile Thr Cys Phe  
       1075                1080                1085  
 Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys Gln Val  
       1090                1095                1100  
 Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala  
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 Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu Trp Ser  
       1125                1130                1135  
 Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn  
       1140                1145                1150  
 Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys Ser Val  
       1155                1160                1165  
 His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr Met Glu  
       1170                1175                1180  
 Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val Val Ala Ser  
       1185                1190                1195                1200  
 Leu Val Pro Ala Ala  
       1205

<210> 30

<211> 3591

<212> DNA

<213> Unknown

<220>

<223> Open reading frame of BEST1:K:trAPAO fusion for bacterial expression vector pGEX-4T-1 or similar vector. gst:BEST1:sp:K:trAPAO fusion, 3591 nt.  
 1-687 gst  
 + polylinker, 688-2163, BEST1 mature; 2164-2199, spacer, 2200-3588, K:trAPAO

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 <222> (1)...(687)  
 <223> gst + polylinker

<221> mat\_peptide  
 <222> (688)...(2163)  
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<221> misc\_feature  
 <222> (2164)...(2199)  
 <223> spacer sequence

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 <222> (2200)...(3588)  
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<221> CDS  
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<221> misc\_feature  
 <222> (2200)...(2202)  
 <223> Extra lysine

<400> 30

atg tcc cct ata cta ggt tat tgg aaa att aag ggc ctt gtg caa ccc	48
Met Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro	
1                   5                   10                   15	

act cga ctt ctt ttg gaa tat ctt gaa gaa aaa tat gaa gag cat ttg

Thr Arg Leu Leu Glu Tyr Leu Glu Lys Tyr Glu Glu His Leu	96
20               25                   30	

tat gag cgc gat gaa ggt gat aaa tgg cga aac aaa aag ttt gaa ttg

Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu	144
35               40                   45	

ggt ttg gag ttt ccc aat ctt cct tat tat att gat ggt gat gtt aaa

Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys	192
50               55                   60	

tta aca cag tct atg gcc atc ata cgt tat ata gct gac aag cac aac

Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn	240
65               70                   75                   80	

atg ttg ggt ggt tgt cca aaa gag cgt gca gag att tca atg ctt gaa

Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu	288
85               90                   95	

gga gcg gtt ttg gat att aga tac ggt gtt tcg aga att gca tat agt

Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser	336
100              105                   110	

aaa gac ttt gaa act ctc aaa gtt gat ttt ctt agc aag cta cct gaa

Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu	384
115              120                   125	

atg ctg aaa atg ttc gaa gat cgt tta tgt cat aaa aca tat tta aat

Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn	432
130              135                   140	

ggt gat cat gta acc cat cct gac ttc atg ttg tat gac gct ctt gat

	480
--	-----

Gly	Asp	His	Val	Thr	His	Pro	Asp	Phe	Met	Leu	Tyr	Asp	Ala	Leu	Asp	
145						150				155					160	
gtt	gtt	tta	tac	atg	gac	cca	atg	tgc	ctg	gat	gcg	ttc	cca	aaa	tta	528
Val	Val	Leu	Tyr	Met	Asp	Pro	Met	Cys	Leu	Asp	Ala	Phe	Pro	Lys	Leu	
														165	170	175
gtt	tgt	ttt	aaa	aaa	cgt	att	gaa	gct	atc	cca	caa	att	gat	aag	tac	576
Val	Cys	Phe	Lys	Lys	Arg	Ile	Glu	Ala	Ile	Pro	Gln	Ile	Asp	Lys	Tyr	
														180	185	190
ttg	aaa	tcc	agc	aag	tat	ata	gca	tgg	cct	ttg	cag	ggc	tgg	caa	gcc	624
Leu	Lys	Ser	Ser	Lys	Tyr	Ile	Ala	Trp	Pro	Leu	Gln	Gly	Trp	Gln	Ala	
														195	200	205
acg	ttt	ggt	ggt	ggc	gac	cat	cct	cca	aaa	tcg	gat	ctg	gtt	ccg	cgt	672
Thr	Phe	Gly	Gly	Gly	Asp	His	Pro	Pro	Lys	Ser	Asp	Leu	Val	Pro	Arg	
														210	215	220
gga	tcc	ccg	gaa	ttc	acg	gat	ttt	ccg	gtc	cgc	agg	acc	gat	ctg	ggc	720
Gly	Ser	Pro	Glu	Phe	Thr	Asp	Phe	Pro	Val	Arg	Arg	Thr	Asp	Leu	Gly	
														225	230	235
cag	gtt	cag	gga	ctg	gcc	ggg	gac	gtg	atg	agc	ttt	cgc	gga	ata	ccc	768
Gln	Val	Gln	Gly	Leu	Ala	Gly	Asp	Val	Met	Ser	Phe	Arg	Gly	Ile	Pro	
														245	250	255
tat	gca	gcf	ccg	ccg	gtg	ggc	ggg	ctg	cgt	tgg	aag	ccg	ccc	caa	cac	816
Tyr	Ala	Ala	Pro	Pro	Val	Gly	Gly	Leu	Arg	Trp	Lys	Pro	Pro	Gln	His	
														260	265	270
gcc	cgf	ccc	tgg	gcf	ggc	gtt	cgc	ccc	gcc	acc	caa	ttt	ggc	tcc	gac	864
Ala	Arg	Pro	Trp	Ala	Gly	Val	Arg	Pro	Ala	Thr	Gln	Phe	Gly	Ser	Asp	
														275	280	285
tgc	ttc	ggc	gcf	gcc	tat	ctt	cgc	aaa	ggc	agc	ctc	gcc	ccc	ggc	gtg	912
Cys	Phe	Gly	Ala	Ala	Tyr	Leu	Arg	Lys	Gly	Ser	Leu	Ala	Pro	Gly	Val	
														290	295	300
agc	gag	gac	tgt	ctt	tac	ctc	aac	gta	tgg	gcf	ccg	tca	ggc	gct	aaa	960
Ser	Glu	Asp	Cys	Leu	Tyr	Leu	Asn	Val	Trp	Ala	Pro	Ser	Gly	Ala	Lys	
														305	310	315
ccc	ggc	cag	tac	ccc	gtc	atg	gtc	tgg	gtc	tac	ggc	ggc	ggc	ttc	gcc	1008
Pro	Gly	Gln	Tyr	Pro	Val	Met	Val	Trp	Val	Tyr	Gly	Gly	Phe	Ala		
														325	330	335
ggc	ggc	acg	gcc	gcc	atg	ccc	tac	tac	gac	ggc	gag	gcf	ctt	gcf	cga	1056
Gly	Gly	Thr	Ala	Ala	Met	Pro	Tyr	Tyr	Asp	Gly	Glu	Ala	Leu	Ala	Arg	
														340	345	350
cag	ggc	gtc	gtc	gtg	gtg	acg	ttt	aat	cgg	acg	aac	atc	ctg	ggc		1104
Gln	Gly	Val	Val	Val	Val	Thr	Phe	Asn	Tyr	Arg	Thr	Asn	Ile	Leu	Gly	
														355	360	365
ttt	ttc	gcc	cat	cct	ggf	ctc	tgc	cgc	gag	agc	ccc	acc	gga	act	tcg	1152
Phe	Phe	Ala	His	Pro	Gly	Leu	Ser	Arg	Glu	Ser	Pro	Thr	Gly	Thr	Ser	
														370	375	380
ggc	aac	tac	ggc	cta	ctc	gac	att	ctc	gcc	gct	ctt	cgf	tgg	gtg	cag	1200
Gly	Asn	Tyr	Gly	Leu	Leu	Asp	Ile	Leu	Ala	Ala	Leu	Arg	Trp	Val	Gln	
														385	390	395

agc aac gcc cgc gcc ttc gga ggg gac ccc ggc cga gtg acg gtc ttt Ser Asn Ala Arg Ala Phe Gly Gly Asp Pro Gly Arg Val Thr Val Phe 405 410 415	1248
ggt gaa tcg gcc gga gcg agc gcg atc gga ctt ctg ctc acc tcg ccg Gly Glu Ser Ala Gly Ala Ser Ala Ile Gly Leu Leu Leu Thr Ser Pro 420 425 430	1296
ctg agc aag ggt ctc ttc cgt ggc gct atc ctc gaa agt cca ggg ctg Leu Ser Lys Gly Leu Phe Arg Gly Ala Ile Leu Glu Ser Pro Gly Leu 435 440 445	1344
acg cga ccg ctc gcg acg ctc gcc gac agc gcc gcc tcg ggc gag cgc Thr Arg Pro Leu Ala Thr Leu Ala Asp Ser Ala Ala Ser Gly Glu Arg 450 455 460	1392
ctc gac gcc gat ctt tcg cga ctg cgc tcg acc gac cca gcc acc ctg Leu Asp Ala Asp Leu Ser Arg Leu Arg Ser Thr Asp Pro Ala Thr Leu 465 470 475 480	1440
atg gcg cgc gcc gac gcg gcc cgc ccg gca tcg cgg gac ctg cgc agg Met Ala Arg Ala Asp Ala Ala Arg Pro Ala Ser Arg Asp Leu Arg Arg 485 490 495	1488
ccg cgt ccg acc gga ccg atc gtc gat ggc cat gtg ctg ccg cag acc Pro Arg Pro Thr Gly Pro Ile Val Asp Gly His Val Leu Pro Gln Thr 500 505 510	1536
gac agc gcg gcg atc gcg gcg ggg cag ctg gcg ccg gtt cgg gtc ctg Asp Ser Ala Ala Ile Ala Gly Gln Leu Ala Pro Val Arg Val Leu 515 520 525	1584
atc gga acc aat gcc gac gaa ggc cgc gcc ttc ctc ggg cgc gcg ccg Ile Gly Thr Asn Ala Asp Glu Gly Arg Ala Phe Leu Gly Arg Ala Pro 530 535 540	1632
atg gag acg cca gcg gac tac caa gcc tat ctg gag gcg cag ttt ggc Met Glu Thr Pro Ala Asp Tyr Gln Ala Tyr Leu Glu Ala Gln Phe Gly 545 550 555 560	1680
gac caa gcc gcc gtg gcg tgc tat ccc ctc gac ggc cgg gcc Asp Gln Ala Ala Val Ala Ala Cys Tyr Pro Leu Asp Gly Arg Ala 565 570 575	1728
acg ccc aag gaa atg gtc gcg cgc atc ttc ggc gac aat cag ttc aat Thr Pro Lys Glu Met Val Ala Arg Ile Phe Gly Asp Asn Gln Phe Asn 580 585 590	1776
cgg ggg gtc tcg gcc ttc tcg gaa gcg ctt gtg cgc cag ggc gcg ccc Arg Gly Val Ser Ala Phe Ser Glu Ala Leu Val Arg Gln Gly Ala Pro 595 600 605	1824
gtg tgg cgt tat cag ttc aac ggt aat acc gag ggt gga aga gcg ccg Val Trp Arg Tyr Gln Phe Asn Gly Asn Thr Glu Gly Arg Ala Pro 610 615 620	1872
gct acc cac gga gcc gaa att ccc tac gtt ttc ggg gtg ttc aag ctc Ala Thr His Gly Ala Glu Ile Pro Tyr Val Phe Gly Val Phe Lys Leu 625 630 635 640	1920
gac gag ttg ggt ctg ttc gat tgg ccg ccc gag ggg ccc acg ccc gcc	1968

Asp Glu Leu Gly Leu Phe Asp Trp Pro Pro Glu Gly Pro Thr Pro Ala				
645	650	655		
gac cgt gcg ctg ggc caa ctg atg tcc tcc gcc tgg gtc cggttccgcc				2016
Asp Arg Ala Leu Gly Gln Leu Met Ser Ser Ala Trp Val Arg Phe Ala				
660	665	670		
aag aat ggc gac ccc gcc ggg gac gcc ctt acc tgg cct gcc tat tct				2064
Lys Asn Gly Asp Pro Ala Gly Asp Ala Leu Thr Trp Pro Ala Tyr Ser				
675	680	685		
acg ggc aag tcg acc atg aca ttc ggt ccc gag ggc cgc gcg gct				2112
Thr Gly Lys Ser Thr Met Thr Phe Gly Pro Glu Gly Arg Ala Ala Val				
690	695	700		
gtg tcg ccc gga cct tcc atc ccc cct tgc gcg gat ggc gcc aag gcg				2160
Val Ser Pro Gly Pro Ser Ile Pro Pro Cys Ala Asp Gly Ala Lys Ala				
705	710	715	720	
ggg ggc gga ggc agc ggc gga ggc agc ggc gga ggc agc aaa gac aac				2208
Gly Gly Gly Ser Gly Gly Ser Gly Gly Ser Lys Asp Asn				
725	730	735		
gtt gcg gac gtg gta gtg gtc gct ggc ttg agc ggt ttg gag acg				2256
Val Ala Asp Val Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr				
740	745	750		
gca cgc aaa gtc cag gcc ggc ggt ctg tcc tgc ctc gtt ctt gag gcg				2304
Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala				
755	760	765		
atg gat cgt gta ggg gga aag act ctg agc gta caa tcg ggt ccc ggc				2352
Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly				
770	775	780		
agg acg act atc aac gac ctc ggc gct gcg tgg atc aat gac agc aac				2400
Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn				
785	790	795	800	
caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg gag ggc gag				2448
Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu				
805	810	815		
ctc cag agg acg act gga aat tca atc cat caa gca caa gac ggt aca				2496
Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr				
820	825	830		
acc act aca gct cct tat ggt gac tcc ttg ctg agc gag gag gtt gca				2544
Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala				
835	840	845		
agt gca ctt gcg gaa ctc ctc ccc gta tgg tct cag ctg atc gaa gag				2592
Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu				
850	855	860		
cat agc ctt caa gac ctc aag gcg agc cct cag gcg aag cgg ctc gac				2640
His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp				
865	870	875	880	
agt gtg agc ttc gcg cac tac tgt gag aag gaa cta aac ttg cct gct				2688
Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala				
885	890	895		

gtt ctc ggc gta gca aac cag atc aca cgc gct ctg ctc ggt gtg gaa Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu 900 905 910	2736
gcc cac gag atc agc atg ctt ttt ctc acc gac tac atc aag agt gcc Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala 915 920 925	2784
acc ggt ctc agt aat att ttc tcg gac aag aaa gac ggc ggg cag tat Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr 930 935 940	2832
atg cga tgc aaa aca ggt atg cag tcg att tgc cat gcc atg tca aag Met Arg Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys 945 950 955 960	2880
gaa ctt gtt cca ggc tca gtg cac ctc aac acc ccc gtc gct gaa att Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val Ala Glu Ile 965 970 975	2928
gag cag tcg gca tcc ggc tgt aca gta cga tcg gcc tcg ggc gcc gtg Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val 980 985 990	2976
ttc cga agc aaa aag gtg gtg gtt tcg tta ccg aca acc ttg tat ccc Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro 995 1000 1005	3024
acc ttg aca ttt tca cca cct ctt ccc gcc gag aag caa gca ttg gcg Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala 1010 1015 1020	3072
gaa aat tct atc ctg ggc tac tat agc aag ata gtc ttc gta tgg gac Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp 1025 1030 1035 1040	3120
aag ccg tgg tgg cgc gaa caa ggc ttc tcg ggc gtc ctc caa tcg agc Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser 1045 1050 1055	3168
tgt gac ccc atc tca ttt gcc aga gat acc agc atc gac gtc gat cga Cys Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg 1060 1065 1070	3216
caa tgg tcc att acc tgt ttc atg gtc gga gac ccg gga cg aag tgg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp 1075 1080 1085	3264
tcc caa cag tcc aag cag gta cga caa aag tct gtc tgg gac caa ctc Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu 1090 1095 1100	3312
cgc gca gcc tac gag aac gcc ggg gcc caa gtc cca gag ccg gcc aac Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn 1105 1110 1115 1120	3360
gtg ctc gaa atc gag tgg tcg aag cag cag tat ttc caa gga gct ccg Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro 1125 1130 1135	3408
agc gcc gtc tat ggg ctg aac gat ctc atc aca ctg ggt tcg gcg ctc	3456

Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu		
1140	1145	1150
aga acg ccg ttc aag agt gtt cat ttc gtt gga acg gag acg tct tta		3504
Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu		
1155	1160	1165
gtt tgg aaa ggg tat atg gaa ggg gcc ata cga tcg ggt caa cga ggt		3552
Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly		
1170	1175	1180
gct gca gaa gtt gtg gct agc ctg gtg cca gca gca tag		3591
Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala		
1185	1190	1195
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Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu		
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Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu		
35 40 45		
Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys		
50 55 60		
Leu Thr Gln Ser Met Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn		
65 70 75 80		
Met Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser Met Leu Glu		
85 90 95		
Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser		
100 105 110		
Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu		
115 120 125		
Met Leu Lys Met Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn		
130 135 140		
Gly Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp		
145 150 155 160		
Val Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu		
165 170 175		
Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr		
180 185 190		
Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala		
195 200 205		
Thr Phe Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg		
210 215 220		
Gly Ser Pro Glu Phe Thr Asp Phe Pro Val Arg Arg Thr Asp Leu Gly		
225 230 235 240		
Gln Val Gln Gly Leu Ala Gly Asp Val Met Ser Phe Arg Gly Ile Pro		
245 250 255		
Tyr Ala Ala Pro Pro Val Gly Gly Leu Arg Trp Lys Pro Pro Gln His		
260 265 270		
Ala Arg Pro Trp Ala Gly Val Arg Pro Ala Thr Gln Phe Gly Ser Asp		
275 280 285		
Cys Phe Gly Ala Ala Tyr Leu Arg Lys Gly Ser Leu Ala Pro Gly Val		
290 295 300		
Ser Glu Asp Cys Leu Tyr Leu Asn Val Trp Ala Pro Ser Gly Ala Lys		
305 310 315 320		

Pro Gly Gln Tyr Pro Val Met Val Trp Val Tyr Gly Gly Gly Phe Ala  
                   325                  330                  335  
 Gly Gly Thr Ala Ala Met Pro Tyr Tyr Asp Gly Glu Ala Leu Ala Arg  
                   340                  345                  350  
 Gln Gly Val Val Val Val Thr Phe Asn Tyr Arg Thr Asn Ile Leu Gly  
                   355                  360                  365  
 Phe Phe Ala His Pro Gly Leu Ser Arg Glu Ser Pro Thr Gly Thr Ser  
                   370                  375                  380  
 Gly Asn Tyr Gly Leu Leu Asp Ile Leu Ala Ala Leu Arg Trp Val Gln  
                   385                  390                  395                  400  
 Ser Asn Ala Arg Ala Phe Gly Gly Asp Pro Gly Arg Val Thr Val Phe  
                   405                  410                  415  
 Gly Glu Ser Ala Gly Ala Ser Ala Ile Gly Leu Leu Leu Thr Ser Pro  
                   420                  425                  430  
 Leu Ser Lys Gly Leu Phe Arg Gly Ala Ile Leu Glu Ser Pro Gly Leu  
                   435                  440                  445  
 Thr Arg Pro Leu Ala Thr Leu Ala Asp Ser Ala Ala Ser Gly Glu Arg  
                   450                  455                  460  
 Leu Asp Ala Asp Leu Ser Arg Leu Arg Ser Thr Asp Pro Ala Thr Leu  
                   465                  470                  475                  480  
 Met Ala Arg Ala Asp Ala Ala Arg Pro Ala Ser Arg Asp Leu Arg Arg  
                   485                  490                  495  
 Pro Arg Pro Thr Gly Pro Ile Val Asp Gly His Val Leu Pro Gln Thr  
                   500                  505                  510  
 Asp Ser Ala Ala Ile Ala Ala Gly Gln Leu Ala Pro Val Arg Val Leu  
                   515                  520                  525  
 Ile Gly Thr Asn Ala Asp Glu Gly Arg Ala Phe Leu Gly Arg Ala Pro  
                   530                  535                  540  
 Met Glu Thr Pro Ala Asp Tyr Gln Ala Tyr Leu Glu Ala Gln Phe Gly  
                   545                  550                  555                  560  
 Asp Gln Ala Ala Ala Val Ala Ala Cys Tyr Pro Leu Asp Gly Arg Ala  
                   565                  570                  575  
 Thr Pro Lys Glu Met Val Ala Arg Ile Phe Gly Asp Asn Gln Phe Asn  
                   580                  585                  590  
 Arg Gly Val Ser Ala Phe Ser Glu Ala Leu Val Arg Gln Gly Ala Pro  
                   595                  600                  605  
 Val Trp Arg Tyr Gln Phe Asn Gly Asn Thr Glu Gly Gly Arg Ala Pro  
                   610                  615                  620  
 Ala Thr His Gly Ala Glu Ile Pro Tyr Val Phe Gly Val Phe Lys Leu  
                   625                  630                  635                  640  
 Asp Glu Leu Gly Leu Phe Asp Trp Pro Pro Glu Gly Pro Thr Pro Ala  
                   645                  650                  655  
 Asp Arg Ala Leu Gly Gln Leu Met Ser Ser Ala Trp Val Arg Phe Ala  
                   660                  665                  670  
 Lys Asn Gly Asp Pro Ala Gly Asp Ala Leu Thr Trp Pro Ala Tyr Ser  
                   675                  680                  685  
 Thr Gly Lys Ser Thr Met Thr Phe Gly Pro Glu Gly Arg Ala Ala Val  
                   690                  695                  700  
 Val Ser Pro Gly Pro Ser Ile Pro Pro Cys Ala Asp Gly Ala Lys Ala  
                   705                  710                  715                  720  
 Gly Gly Gly Ser Gly Gly Ser Gly Gly Ser Lys Asp Asn  
                   725                  730                  735  
 Val Ala Asp Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr  
                   740                  745                  750  
 Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala  
                   755                  760                  765  
 Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly  
                   770                  775                  780  
 Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn  
                   785                  790                  795                  800  
 Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu  
                   805                  810                  815

Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr  
       820                  825                  830  
 Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala  
       835                  840                  845  
 Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu  
       850                  855                  860  
 His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp  
       865                  870                  875                  880  
 Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala  
       885                  890                  895  
 Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu  
       900                  905                  910  
 Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala  
       915                  920                  925  
 Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr  
       930                  935                  940  
 Met Arg Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys  
       945                  950                  955                  960  
 Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val Ala Glu Ile  
       965                  970                  975  
 Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val  
       980                  985                  990  
 Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro  
       995                  1000                1005  
 Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala  
       1010                1015                1020  
 Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp  
       1025                1030                1035                1040  
 Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser  
       1045                1050                1055  
 Cys Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg  
       1060                1065                1070  
 Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp  
       1075                1080                1085  
 Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu  
       1090                1095                1100  
 Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn  
       1105                1110                1115                1120  
 Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro  
       1125                1130                1135  
 Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu  
       1140                1145                1150  
 Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu  
       1155                1160                1165  
 Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly  
       1170                1175                1180  
 Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala  
       1185                1190                1195

<210> 32  
 <211> 1803  
 <212> DNA  
 <213> Unknown

<220>  
 <221> CDS  
 <222> (1)...(1803)

<223> Glyc(-)APAO coding sequence; mutation in putative  
 glycosylation sites

<400> 32

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1 5 10 15	
gca ggg tat tct cac gtc ggc gta ggc cca gac gga ggg agg tat gtg Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Arg Tyr Val	96
20 25 30	
aca ata gct gga cag att gga caa gac gct tcg ggc gtg aca gac cct Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro	144
35 40 45	
gcc tac gag aaa cag gtt gcc caa gca ttc gcc aat ctg cga gct tgc Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys	192
50 55 60	
ctt gct gca gtt gga gcc act tca aac gac gtc acc aag ctc aat tac Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr	240
65 70 75 80	
tac atc gtc gac tac gcc ccg agc aaa ctc acc gca att gga gat ggg Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly	288
85 90 95	
ctg aag gct acc ttt gcc ctt gac agg ctc cct cct tgc acg ctg gtg Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val	336
100 105 110	
cca gtg tcg gcc ttg tct tca cct gaa tac ctc ttt gag gtt gat gcc Pro Val Ser Ala Leu Ser Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala	384
115 120 125	
acg gcg ctg gtg ccg gga cac acg acc cca gac aac gtt gct gac gtg Thr Ala Leu Val Pro Gly His Thr Thr Pro Asp Asn Val Ala Asp Val	432
130 135 140	
gta gtg gtg ggc gct ttg agc ggt ttg gag acg gca cgc aaa gtc Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val	480
145 150 155 160	
cag gcc ggc ggt ctg tcc tgc ctc gtt gag gct atg gat cgt gta Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val	528
165 170 175	
ggg gga aag act ctg agc gta caa tcg ggt ccc ggc agg acg act atc Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile	576
180 185 190	
aac gac ctc ggc gct gcg tgg atc aat gat agc aat cag gcc gaa gta Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ala Glu Val	624
195 200 205	
tcc aga ttg ttt gaa aga ttt cat ttg gag ggc gag ctc cag agg acg Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr	672
210 215 220	
act gga aat tca atc cat caa gca caa gac ggt aca acc act aca gct Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala	720
225 230 235 240	
cct tat ggt gac tcc ttg ctg agc gag gag gtt gca agt gca ctt gcg Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala	768

	245	250	255	
gaa ctc ctc ccc gta tgg tct cag ctg atc gaa gag cat agc ctt caa Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln	260	265	270	816
gac ctc aag gcg agc cct cag gcg aag cg <sup>g</sup> ctc gac agt gtg agc ttc Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe	275	280	285	864
gcg cac tac tgt gag aag gaa cta aac ttg cct gct gtt ctc ggc gta Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val	290	295	300	912
gca aac cag atc aca cgc gct ctg ctc ggt gtg gaa gcc cac gag atc Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile	305	310	315	960
agc atg ctt ttt ctc acc gac tac atc aag agt gcc acc ggt ctc agt Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser	325	330	335	1008
aat att ttc tcg gac aag aaa gac ggc ggg cag tat atg cga tgc aaa Asn Ile Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys	340	345	350	1056
aca ggt atg cag tcg att tgc cat gcc atg tca aag gaa ctt gtt cca Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro	355	360	365	1104
ggc tca gtg cac ctc aac acc ccc gtc gct gaa att gag cag tcg gca Gly Ser Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala	370	375	380	1152
tcc ggc tgt aca gta cga tcg gcc tcg ggc gcc gtg ttc cga agc aaa Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys	385	390	395	1200
aag gtg gtg gtt tcg tta ccg aca acc ttg tat ccc acc ttg aca ttt Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe	405	410	415	1248
tca cca cct ctt ccc gcc gag aag caa gca ttg gcg gaa aat tct atc Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile	420	425	430	1296
ctg ggc tac tat agc aag ata gtc ttc gta tgg gac aag ccg tgg tgg Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp	435	440	445	1344
cgc gaa caa ggc ttc tcg ggc gtc ctc caa tcg agc tgt gac ccc atc Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile	450	455	460	1392
tca ttt gcc aga gat acc agc atc gac gtc gat cga caa tgg tcc att Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile	465	470	475	1440
acc tgt ttc atg gtc gga gac ccg gga cg <sup>g</sup> aag tgg tcc caa cag tcc Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser	485	490	495	1488

aag cag gta cga caa aag tct gtc tgg gac caa ctc cgc gca gcc tac Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr 500 505 510	1536
gag aac gcc ggg gcc caa gtc cca gag ccg gcc aac gtg ctc gaa atc Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile 515 520 525	1584
gag tgg tcg aag cag cag tat ttc caa gga gct ccg agc gcc gtc tat Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr 530 535 540	1632
ggg ctg aac gat ctc atc aca ctg ggt tcg gcg ctc aga acg ccg ttc Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe 545 550 555 560	1680
aag agt gtt cat ttc gtt gga acg gag acg tct tta gtt tgg aaa ggg Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly 565 570 575	1728
tat atg gaa ggg gcc ata cga tcg ggt caa cga ggt gct gca gaa gtt Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val 580 585 590	1776
gtg gct agc ctg gtg cca gca gca tag Val Ala Ser Leu Val Pro Ala Ala * 595 600	1803

<210> 33  
 <211> 600  
 <212> PRT  
 <213> Unknown

<400> 33  
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 Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Gly Arg Tyr Val  
 20 25 30  
 Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro  
 35 40 45  
 Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys  
 50 55 60  
 Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr  
 65 70 75 80  
 Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly  
 85 90 95  
 Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val  
 100 105 110  
 Pro Val Ser Ala Leu Ser Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala  
 115 120 125  
 Thr Ala Leu Val Pro Gly His Thr Thr Pro Asp Asn Val Ala Asp Val  
 130 135 140  
 Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val  
 145 150 155 160  
 Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val  
 165 170 175  
 Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile  
 180 185 190  
 Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ala Glu Val  
 195 200 205  
 Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr

210	215	220
Thr	Gly	Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Thr Ala
225	230	235
Pro	Tyr	Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala
		245
Glu	Leu	Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln
		260
Asp	Leu	Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe
		275
Ala	His	Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val
		290
Ala	Asn	Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile
305	310	315
Ser	Met	Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser
		325
Asn	Ile	Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys
		340
Thr	Gly	Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro
		355
Gly	Ser	Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala
		370
Ser	Gly	Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys
385	390	395
Lys	Val	Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe
		405
Ser	Pro	Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile
		420
Leu	Gly	Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp
		435
Arg	Glu	Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile
		450
Ser	Phe	Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile
465	470	475
Thr	Cys	Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser
		485
Lys	Gln	Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr
		500
Glu	Asn	Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile
		515
Glu	Trp	Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr
		530
Gly	Leu	Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe
545	550	555
Lys	Ser	Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly
		565
Tyr	Met	Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val
		580
Val	Ala	Ser Leu Val Pro Ala Ala
		595
		600

<210> 34  
 <211> 37  
 <212> DNA  
 <213> Artificial Sequence

<220>  
 <223> 37-mer oligonucleotide

<400> 34  
 ggggaattca tggcacttgc accgagctac atcaatc

<211> 1929  
 <212> DNA  
 <213> Exophiala spinifera

<220>  
 <221> intron  
 <222> (739) ... (811)

<221> intron  
 <222> (1134) ... (1186)

<400> 35

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cacatcgccg	taggccccaaa	cgaagcgagg	tatgtgacaa	tagctggaca	gattggacaa	120
gacgcgttgg	gcgtgacaga	cccagccctac	gagaaacagg	ttgccccaa	gatcgccaa	180
ctgcgagctt	gccttgcgtc	agttggagcc	tcttcaaacs	acgtcacca	gctcaattac	240
tacatcgctg	actacgcccc	gagcaaactc	accgcaattt	gagatgggt	gaagtctacc	300
tttgccttg	acaggctccc	tccttgcacg	ctggtgccag	taccggcctt	ggcttcacct	360
gaataccctt	ttgagggtta	tgccacggcg	ctggtgccag	gacactcgac	cccagacaac	420
gttgcggacg	tggttagtgg	gggcgctggc	ttgagcgggt	tggagacggc	acgcaaagtc	480
caggccgccc	gtctgtctt	cctcgttctt	gaggcgttgg	atcgtgttgg	ggaaaagact	540
ctgagcgtac	aatcggttcc	cggcaggacy	actatcaacg	acctcgccgc	tgcgtggatc	600
aatgacagca	acccaaagcga	agtatccaga	ttgtttgaaa	gatttcattt	ggagggcggag	660
ctccagagga	cgaccggaaa	ttcaatccat	caagcacaag	acggtacaac	cactacagct	720
ccttatggtg	actcccccgt	aagcacaatc	ccactttgtt	atgagacctc	tgtcgagtgt	780
agaatacagt	cactgactcc	acttcgttca	gctgagcggag	gagggttgc	gtgcacttgc	840
ggaactcctc	cccttatggt	ctcagctgt	cgaagagtat	agccttgc	acccaaaggc	900
gagccctcag	gcgaagcggc	tcgacagtgt	gagcttcgc	cactactgt	agaaggacct	960
aaacttgcct	gcttctca	gcgtggccaa	ccagatcaca	cgcgtctgc	tgcgtgttgg	1020
agcccaacgg	atcagcatgc	tttttctcac	cgactacatc	aagagtgc	ccgtctcag	1080
taatattgtc	tcggacaaga	aagacggcg	gcagtatatg	cgatgc	aaaaa	1140
cggtgtctc	tcaggttaggg	gactcggtt	tttagtgc	ttccaggat	gcagtcgatt	1200
tgccatgcc	tgctaaagga	acttgcgtt	ggctcagtgc	acctcaacac	cccgtcgct	1260
ggaattgagc	agtcggcg	cggctgtata	gtacgatcg	cctcggccgc	cgtgttccga	1320
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cctcttcccg	ccgagaagca	agcattggcg	aaaaaatct	tcctcggct	ctatagcaag	1440
atagtcttcg	tatgggacaa	cccggtgtt	cgcgaacaag	gcttcgc	cgtcctccaa	1500
tcgagctgt	acccatctc	atttgcac	gataccagca	tcgaagtgc	tcgcaatgg	1560
tccattacct	gtttcatgg	cggagacccg	ggacggaat	gttccaaaca	gtccaaggcag	1620
gtacgacaaa	agtctgtctg	ggaccaactc	cgcgcagct	acgagaacgc	cggggcccaa	1680
gtcccagagc	cggccaaacgt	gctcgaaatc	gagtggcga	agcagcagta	tttcaagga	1740
gctccgagcg	cggtctatgg	gctgaacat	ctcatcac	tgggttccgc	gctcagaac	1800
ccgttcaagt	gttttcattt	cggtggaaacg	gagacgtt	tagtttggaa	agggtatatg	1860
gaagggggca	tacgatcg	tcaacgagg	gctcagaag	ttgtggct	cctggcaca	1920
gcagcatag						1929

<210> 36  
 <211> 600  
 <212> PRT  
 <213> Exophiala spinifera

<400> 36

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Ala	Gly	Tyr	Ser	His	Ile	Gly	Val	Gly	Pro	Asn	Glu	Ala	Arg	Tyr	Val
					20			25			30				
Thr	Ile	Ala	Gly	Gln	Ile	Gly	Gln	Asp	Ala	Leu	Gly	Val	Thr	Asp	Pro
					35			40			45				
Ala	Tyr	Glu	Lys	Gln	Val	Ala	Gln	Ala	Phe	Ala	Asn	Leu	Arg	Ala	Cys
					50			55			60				
Leu	Ala	Ala	Val	Gly	Ala	Ser	Ser	Asn	Asp	Val	Thr	Lys	Leu	Asn	Tyr
					65			70			75				
															80

Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly  
                   85                  90                  95  
 Leu Lys Ser Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val  
                   100              105              110  
 Pro Val Pro Ala Leu Ala Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala  
                   115              120              125  
 Thr Ala Leu Val Pro Gly His Ser Thr Pro Asp Asn Val Ala Asp Val  
                   130              135              140  
 Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val  
                   145              150              155              160  
 Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val  
                   165              170              175  
 Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile  
                   180              185              190  
 Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val  
                   195              200              205  
 Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr  
                   210              215              220  
 Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala  
                   225              230              235              240  
 Pro Tyr Gly Asp Ser Pro Leu Ser Glu Glu Val Ala Ser Ala Leu Ala  
                   245              250              255  
 Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu Tyr Ser Leu Glu  
                   260              265              270  
 Asp Pro Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe  
                   275              280              285  
 Ala His Tyr Cys Glu Lys Asp Leu Asn Leu Pro Ala Val Leu Ser Val  
                   290              295              300  
 Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile  
                   305              310              315              320  
 Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser  
                   325              330              335  
 Asn Ile Val Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys  
                   340              345              350  
 Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro  
                   355              360              365  
 Gly Ser Val His Leu Asn Thr Pro Val Ala Gly Ile Glu Gln Ser Ala  
                   370              375              380  
 Ser Gly Cys Ile Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys  
                   385              390              395              400  
 Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe  
                   405              410              415  
 Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Lys Ser Ile  
                   420              425              430  
 Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Asn Pro Trp Trp  
                   435              440              445  
 Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile  
                   450              455              460  
 Ser Phe Ala Arg Asp Thr Ser Ile Glu Val Asp Arg Gln Trp Ser Ile  
                   465              470              475              480  
 Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser  
                   485              490              495  
 Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr  
                   500              505              510  
 Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile  
                   515              520              525  
 Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr  
                   530              535              540  
 Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe  
                   545              550              555              560  
 Lys Cys Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly  
                   565              570              575

Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val  
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 Val Ala Ser Leu Val Pro Ala Ala  
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<210> 37  
<211> 1929  
<212> DNA  
<213> *Exophiala spinifera*

<220>  
<221> intron  
<222> (739)...(811)

<221> intron  
<222> (1134)...(1186)

<400> 37

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gacgcTTgg	gcgtgacaga	cccagcc tac	gagaaaacagg	ttgccc aagc	attcgccaat
ctgcgagctt	gccttgc tgc	agttggagcc	tcttcaa acg	acgtcac caa	gctcaattac
tacatcgctg	actacgcccc	gagcaaactc	accgcaattg	gagatggc t	gaagtctacc
tttgcCTTg	acaggctccc	tccttgc acg	ctggtgc cag	taccggc tt	ggcttcac ct
gaataccctt	ttgagggtga	cgc acgc g	ctggtgc cag	gacactcgac	cccagacaac
gttgcggacg	tggtagtgg	gggcgct gg	ttgagcggc t	tggagacggc	acgcaaa agtc
caggccgccc	gtctgtcctg	cctcg ttctt	gaggcgtatgg	atcg tga tag	gggaaaagact
ctgagcgtac	aatcggttcc	cggcaggacg	actatcaacg	acctcggcgc	tgcgtggatc
aatgacagca	accaaaagcg	agtatccaga	ttgtttgaaa	gatttcattt	ggagggc gag
ctccagagga	cgacccggaa	ttcaatccat	caagcacaag	acggtacaa	cactacagct
ccttatggtg	actccccgg	aagcacaatc	ccactttgt	atgagaccc	tgtcgagtgt
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ggaactcctc	cccgatgg	ctcagctgat	cgaagagtat	agccttga	accccaaggc
gagccctcag	gcgaagcggc	tcgacagtgt	gagcttcgc	cactactgt	agaaggacct
aaacttgcct	gctgttctca	gcgtggcaaa	ccagatcaca	cgcgctctgc	tcggtgtgga
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taatattgtc	tcggacaaga	aagacggcgg	gcagtatatg	cgatgc	aaaaa cagg tgcgt
cggtgtcctc	tcaaggtaggg	gactcgttc	tttagtggtca	ttccagg	ttat gcagtcgatt
tgccatgcc	tgtcaaagg	acttgttcca	ggctcagtgc	accta	acac ccccgctcg
ggaattgagc	agtccggcgtc	cggctgtata	gtacgatcgg	cctcgggcgc	cgtgttccga
agcaaaaagg	tggtggttc	gttaccgaca	acattgtatc	ccaccttgc	attttccacca
cctcttcccg	ccgagaagca	agcattggcg	aaaaaatcta	tcctcggt	ctatagcaag
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tgcagctgt	accccatctc	atttgc caga	gataccagca	tcgaagtcg	tcggcaatgg
tccattacct	gttcatgg	cggagaccc	ggacggaa	gttcccaaca	gtccaagcag
gtacgacaaa	agtctgtctg	ggaccaactc	cgccgcagc	acgagaacgc	cggggcccaa
gtcccagagc	cggccaacgt	gctcgaaatc	gagtggtcg	agcagcag	gtttccaagga
gctccgagcg	ccgtctatgg	gctgaacgat	ctcatcacac	tgggttccgg	gc tcagaac
ccgttcaagt	gtttcattt	cgttggaa	gagacgtctt	tagtttggaa	agggtatatg
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<211> 600  
<212> PRT  
<213> *Exophiala spinifera*

<400> 38

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      20          25          30

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Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Leu Gly Val Thr Asp Pro  
                   35                  40                  45  
 Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys  
                   50                  55                  60  
 Leu Ala Ala Val Gly Ala Ser Ser Asn Asp Val Thr Lys Leu Asn Tyr  
                   65                  70                  75                  80  
 Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly  
                   85                  90                  95  
 Leu Lys Ser Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val  
                   100                105                110  
 Pro Val Pro Ala Leu Ala Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala  
                   115                120                125  
 Thr Ala Leu Val Pro Gly His Ser Thr Pro Asp Asn Val Ala Asp Val  
                   130                135                140  
 Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val  
                   145                150                155                160  
 Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val  
                   165                170                175  
 Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile  
                   180                185                190  
 Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val  
                   195                200                205  
 Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr  
                   210                215                220  
 Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala  
                   225                230                235                240  
 Pro Tyr Gly Asp Ser Pro Leu Ser Glu Glu Val Ala Ser Ala Leu Ala  
                   245                250                255  
 Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu Tyr Ser Leu Glu  
                   260                265                270  
 Asp Pro Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe  
                   275                280                285  
 Ala His Tyr Cys Glu Lys Asp Leu Asn Leu Pro Ala Val Leu Ser Val  
                   290                295                300  
 Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile  
                   305                310                315                320  
 Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser  
                   325                330                335  
 Asn Ile Val Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys  
                   340                345                350  
 Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro  
                   355                360                365  
 Gly Ser Val His Leu Asn Thr Pro Val Ala Gly Ile Glu Gln Ser Ala  
                   370                375                380  
 Ser Gly Cys Ile Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys  
                   385                390                395                400  
 Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe  
                   405                410                415  
 Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Lys Ser Ile  
                   420                425                430  
 Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Asn Pro Trp Trp  
                   435                440                445  
 Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile  
                   450                455                460  
 Ser Phe Ala Arg Asp Thr Ser Ile Glu Val Asp Arg Gln Trp Ser Ile  
                   465                470                475                480  
 Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser  
                   485                490                495  
 Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr  
                   500                505                510  
 Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile  
                   515                520                525

Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr  
 530 535 540  
 Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe  
 545 550 555 560  
 Lys Cys Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly  
 565 570 575  
 Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val  
 580 585 590  
 Val Ala Ser Leu Val Pro Ala Ala  
 595 600

<210> 39  
 <211> 1930  
 <212> DNA  
 <213> *Exophiala spinifera*

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<221> intron  
 <222> (1134) ... (1187)  
  
 <221> misc\_feature  
 <222> (648) ... (648)  
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gacgcttcgg gcgtgacaga ccctgcctac gagaacacagg ttgcccagc attcgccat	180
ctgcgagtt gccttgcgc agttggagcc acttcaaacg acgtcaccaa gctcaattac	240
tacatcgctcg actacgcccc gagcaaactc accgaattt gagatggct gaaggctacc	300
tttgccttg acaggctccc tccttgcacg ctgtgtccag tgtcggcctt gtcttcac	360
gaataacctt tttaggttga tgccacggcg ctgtgtccgg gacacacgac cccagacaac	420
gttgcggacg tggtagtggt gggcgctggc ttgagcgggtt tggagacggc acgaaagtc	480
caggccggcg gtctgtcctt cctcgtttctt gagggcatgg atcgtgttgg gggaaagact	540
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aatgacagca accaaagcga agtatccaga ttgtttgaaa gatttcatnt ggagggcgag	660
ctccagagga cgactggaaa ttcaatccat caagcacaag acggtaaac cactacagct	720
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agaatacagt cactgattcc acttcgtcca gctgagcgag gaggttgcaa gtgcacttgc	840
ggaactcctc cccgtatggt ctcagctgtat cgaagagcat agccttcaag acctcaaggc	900
gagccctcag gcgaaagcgcc tcgacagtgt gagcttgcgc cactactgtg agaaggaact	960
aaacttgcct gctgttctcg gcgttagcaaa ccagatcaca cgcgtctgc tcgtgttgg	1020
agcccacag atcagcatgc ttttctcac cgactacatc aagagtgcca ccgtctcag	1080
taatattttc tcggacaaga aagacggcg gcagtatatg cgatgcaaaa caggtgcgt	1140
tggtgtcgtc tcaggtgggg gactcgtttcaagtggc atttcaggta tgcaagtgc	1200
ttgccatgcc atgtcaaagg aacttgcctt aggctcgttgc cacctcaaca ccccgctgc	1260
tgaaatttgc cagtcggcat cccgctgtac agtacgtatcg gcctcgccgc ccgtgttcc	1320
aagaaaaaag gtgggtgtt cgttaccgac aacccgttat cccaccttga cattttcacc	1380
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gatagtcttc gtatggaca agccgtgtg gcgcaacaa ggcttctcg gcgtcctcca	1500
atcgagctgt gacccatct catttgcac agataccagc atcgacgtcg atcgacaatg	1560
gtccattacc tggatcatgg tcggagaccc gggacggaa tggtcccaac agtccaagca	1620
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agtcccagag ccggccaacg tgctcgaaat cgagtggctg aagcagcagt attccaagg	1740
agctccgagc gccgtctatg ggctgaacga tctcatcaca ctgggttcgg cgctcagaac	1800
cccggtcaag agtggcatt tcgttggaa ggagacgtct ttagtttggaa aagggttat	1860
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<210> 40  
<211> 598  
<212> PRT  
<213> Exophiala spinifera

<220>  
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Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys  
50 55 60  
Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr  
65 70 75 80  
Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly  
85 90 95  
Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val  
100 105 110  
Pro Val Ser Ala Leu Ser Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala  
115 120 125  
Thr Ala Leu Val Pro Gly His Thr Thr Pro Asp Asn Val Ala Asp Val  
130 135 140  
Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val Gln  
145 150 155 160  
Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val Gly  
165 170 175  
Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile Asn  
180 185 190  
Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Ser  
195 200 205  
Arg Leu Phe Glu Arg Phe His Xaa Glu Gly Glu Leu Gln Arg Thr Thr  
210 215 220  
Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro  
225 230 235 240  
Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu  
245 250 255  
Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln Asp  
260 265 270  
Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe Ala  
275 280 285  
His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val Asn  
290 295 300  
Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met  
305 310 315 320  
Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn Ile  
325 330 335  
Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr Gly  
340 345 350  
Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro Gly Ser  
355 360 365  
Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser Gly  
370 375 380  
Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys Lys Val  
385 390 395 400  
Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe Ser Pro

405	410	415
Pro Leu Pro Ala Glu Lys Gln Ala	Leu Ala Glu Asn Ser Ile	Leu Gly
420	425	430
Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro	Trp Trp Arg	Glu
435	440	445
Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys	Asp Pro Ile Ser Phe	
450	455	460
Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln	Trp Ser Ile Thr Cys	
465	470	475
Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln	Gln Ser Lys Gln	
485	490	495
Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala	Ala Tyr Glu Asn	
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Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val	Leu Glu Ile Glu Trp	
515	520	525
Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala	Val Tyr Gly Leu	
530	535	540
Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr	Pro Phe Lys Ser	
545	550	555
Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp	Lys Gly Tyr Met	
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Ser Leu Val Pro Ala Ala		
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<210> 41  
 <211> 1928  
 <212> DNA  
 <213> Rhinocladiella atrovirens

<220>  
 <221> intron  
 <222> (739) ... (811)  
  
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 <222> (1134) ... (1185)

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ctgcgagctt gtcttgcgc agttggagcc acttcaaacg acattaccaa gctcaattac	240
tacatcgctcg actacaaccc gagcaaactc accgcaattt gagatgggct gaaggctacc	300
tttgccttg acaggctccc tccttgcacg ctgtgcccag tgccggccct ggcttcacct	360
gaataccctt ttgaggttga tgccacggcg ctgttccag gacactcaac cccagacaat	420
gttgcggacg tggcgttgtt gggcgcttgc ttgagcggtt tggagacggc acgaaagtc	480
caggctgccg ggctgtccct cctcggttctt gaggcgtatgg atcgtgtggg gggaaagact	540
ctgagcgtaa aatcggtcc cggcaggacg gctatcaatg acctcggegc tgctgtggatc	600
aatgacagca accaaagcga agtattcaaa ttatttgaaa gatttcattt ggagggcgag	660
ctccagagga cgaccggaaa ttcaatccat caagcacaag acggtacaac cactacagct	720
ccttatgttgc attcccttgtt aagcacaatt ccattttgtt atgagacctc tgtctgtgt	780
agaatacagt cgctgactcc acatcggtcca gctgagcggag gaggttgcaa gtgcactcgc	840
ggaactcctt ccccatggt ctcagctgtatcgatcagaagacat agtcttgcggacc accccaaggc	900
gagccctcaa gcgaaaggcgc tcgacagtgt gagcttcgca cactactgtg agaaggatct	960
aagcttgcct gctgttctcg gcgtggcaaaa ccagatcaca cgcgccttcgc tcgggtgtgg	1020
agcccacggat atcagcatgc tttttctcac cgactacatc aagagtgcac ccgtctcag	1080
taatattgttc tcggataaga aagacggtgg gcagttatcgatcgatgcaaaa caggtgcgtg	1140
tggtgttctc tcagtggtggactcgatctt tagtggatcat tccaggtatgcgtcgatctt	1200
gcccattccat gtcaaaggaa cttgttccag gctcgtgcac cctcaacacc cccgtcgccg	1260
aaatttgagca gtccggatcc ggctgtacag tacgatcgac ctcggggccgc gtgttccgaa	1320
gtaaaaaggt ggtggtttcg ttaccgacaa ccttgcatac caccatgtatac ttttcaccac	1380

ctttccgc	cgagaagcaa	gcattggctg	aaaaatccat	cctgggctac	tatagcaaga	1440
tagtctcgt	atgggacaag	ccgtggggc	gcgaacaagg	cttctgggc	gtcctccaat	1500
cgagctgt	ccccatctca	tttgccagag	ataccagcat	cgaagtgcgt	cgcaatgg	1560
ccattacct	tttcatggc	ggagaccgg	gacggaagt	gtcccaacag	tccaaagg	1620
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cgccgagcgt	cgtctatggg	ctgaactgtc	tcaacacact	gggttcggcg	ctcagaacgc	1800
cgttcaaggg	tgttcatttc	gttggAACGG	agacgtctt	ggtttgaaa	ggtatatgg	1860
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cagcatag						1928

<210> 42  
 <211> 598  
 <212> PRT  
 <213> Rhinocladiella atrovirens

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					20				25					30			
Thr	Ile	Ala	Gly	Gln	Ile	Gly	Gln	Asp	Ala	Ser	Ala	Val	Thr	Asp	Pro		
					35			40						45			
Ala	Tyr	Glu	Lys	Gln	Val	Ala	Gln	Ala	Phe	Ala	Asn	Leu	Arg	Ala	Cys		
					50			55						60			
Leu	Ala	Ala	Val	Gly	Ala	Thr	Ser	Asn	Asp	Ile	Thr	Lys	Leu	Asn	Tyr		
					65			70			75			80			
Tyr	Ile	Val	Asp	Tyr	Asn	Pro	Ser	Lys	Leu	Thr	Ala	Ile	Gly	Asp	Gly		
					85			90			95						
Leu	Lys	Ala	Thr	Phe	Ala	Leu	Asp	Arg	Leu	Pro	Pro	Cys	Thr	Leu	Val		
					100			105						110			
Pro	Val	Pro	Ala	Leu	Ala	Ser	Pro	Glu	Tyr	Pro	Phe	Glu	Val	Asp	Ala		
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Thr	Ala	Leu	Val	Pro	Gly	His	Ser	Thr	Pro	Asp	Asn	Val	Ala	Asp	Val		
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Val	Val	Val	Gly	Ala	Gly	Leu	Ser	Gly	Leu	Glu	Thr	Ala	Arg	Lys	Val		
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Gln	Ala	Ala	Gly	Leu	Ser	Cys	Leu	Val	Leu	Glu	Ala	Met	Asp	Arg	Val		
					165			170			175						
Gly	Gly	Lys	Thr	Leu	Ser	Val	Gln	Ser	Gly	Pro	Gly	Arg	Thr	Ala	Ile		
					180			185			190						
Asn	Asp	Leu	Gly	Ala	Ala	Trp	Ile	Asn	Asp	Ser	Asn	Gln	Ser	Glu	Val		
					195			200			205						
Phe	Lys	Leu	Phe	Glu	Arg	Leu	Glu	Gly	Glu	Leu	Gln	Arg	Thr	Thr	Gly		
					210			215			220						
Asn	Ser	Ile	His	Gln	Ala	Gln	Asp	Gly	Thr	Thr	Thr	Thr	Ala	Pro	Tyr		
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Gly	Asp	Ser	Leu	Leu	Ser	Glu	Glu	Val	Ala	Ser	Ala	Leu	Ala	Glu	Leu		
					245			250			255						
Leu	Pro	Ala	Trp	Ser	Gln	Leu	Ile	Glu	Glu	His	Ser	Leu	Glu	Asp	Pro		
					260			265			270						
Lys	Ala	Ser	Pro	Gln	Ala	Lys	Gln	Leu	Asp	Ser	Val	Ser	Phe	Ala	His		
					275			280			285						
Tyr	Cys	Glu	Lys	Asp	Leu	Ser	Leu	Pro	Ala	Val	Leu	Gly	Val	Ala	Asn		
					290			295			300						
Gln	Ile	Thr	Arg	Ala	Leu	Leu	Gly	Val	Glu	Ala	His	Glu	Ile	Ser	Met		
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Val	Ser	Asp	Lys	Lys	Asp	Gly	Gly	Gln	Tyr	Met	Arg	Cys	Lys	Thr	Gly		
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355	360	365
Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser Gly		
370	375	380
Cys Thr Val Arg Ser Ala Ser Gly Gly Val Phe Arg Ser Lys Lys Val		
385	390	395
400		400
Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Ile Phe Ser Pro		
405	410	415
Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Lys Ser Ile Leu Gly		
420	425	430
Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp Arg Glu		
435	440	445
Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser Phe		
450	455	460
Ala Arg Asp Thr Ser Ile Glu Val Asp Arg Gln Trp Ser Ile Thr Cys		
465	470	475
480		480
Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser Lys Gln		
485	490	495
Val Arg Gln Lys Ser Val Trp Asn Gln Leu Arg Ala Ala Tyr Glu Asn		
500	505	510
Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile Glu Trp		
515	520	525
Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Val Val Tyr Gly Leu		
530	535	540
Asn Cys Leu Asn Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe Lys Gly		
545	550	555
560		560
Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly Tyr Met		
565	570	575
Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val Val Ala		
580	585	590
Ser Leu Val Pro Ala Ala		
595		

<210> 43  
 <211> 1928  
 <212> DNA  
 <213> Rhinocladiella atrovirens

<220>  
 <221> intron  
 <222> (739)...(811)

<221> intron  
 <222> (1134)...(1186)

<400> 43

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gacgcttcgg	ccgtgacaga	ccctgcctac	gagaaacagg	ttgcccggc	attcgccaaac	180
ctgcgagctt	gtcttgcgtc	agttggagcc	acttcaaacg	acattaccaa	gctcaattac	240
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tttgccttgc	acaggctccc	tccttgcacg	ctgggtcccg	tgccggccct	ggcttcacct	360
gaataccctt	tttagggttga	tgccacggcg	ctgttcccg	gacactcaac	cccagacaat	420
gttgcgga	tggtcggtgt	gggcgctggc	ttgagcgggtt	tggagacggc	acgcaaagtc	480
caggctggcg	ggctgtccctg	cctcggttctt	gaggcgatgg	atcggtgtgg	ggggaaagact	540
ctgagcgatc	aatcggttcc	cggcaggacg	actatcaatg	acctcgccgc	tgctgtggatc	600
aatgacagca	acccaaagcg	agtattcaaa	ttatttgaaa	gatttcattt	ggagggcgag	660
ctccagagga	cgaccggaaa	ttcaatccat	caagcacaag	acggtacaac	cactacagct	720
ccttatgtgt	attccctgtgt	aagcacaattt	ccatcttgcgt	atgagacactc	tgtcggtgtgt	780
agaatacagt	cgctgactcc	acatcggtcca	gctgagcgag	gagggttgcaa	gtgcactcgcc	840
ggaactcctt	cccgcatgtt	ctcagctgtat	cgaagagcat	agtcttgaag	accccaaggc	900
gagccctcaa	gcgaaggcgc	tgcacagtgt	gagcttcgca	cactactgtg	agaaggatct	960
aaacttgctt	gctgttctcg	gcgtggccaaa	ccagatcaca	cgcgctctgc	tcgggtgtgg	1020

agccccacgag atcagcatgt	tttttctcac	cgactacatc	aagagtgc	cca	ccgg	tctc	cag	1080
taatattgtc tcggataaga	aagacgg	tg	gcag	tat	atg	caaaa	cagg	1140
tggtgttctc tcagtg	ggag	actcg	tttct	tag	gg	tcat	tcc	1200
gccatg	ccat	gtcaaa	aggaa	ctt	gtt	ccag	gtat	1260
aaattg	agca	gtcg	gcatcc	gg	ctg	tacag	tac	1320
gtaaaa	aggt	gg	ttcg	ttacc	cg	ac	ttt	ttt
ctcttccc	gc	gaa	gag	cata	cc	ttt	gtata	1380
tagtctt	cg	aaa	atcc	at	ct	cc	at	1440
cgagctgt	atgg	aca	ag	cc	gtt	gg	cc	1500
ccccatct	ca	ttt	g	cc	ca	ag	at	1560
ccattac	ctg	ttt	cat	gg	gg	ac	cc	1620
tacgac	aga	gg	gac	cc	gg	aa	ac	1680
tc	cc	ca	gag	cc	gg	gg	cc	1740
cgcc	gag	cc	g	ct	ca	gg	gg	1800
cg	tt	ca	ag	tt	cc	gg	gg	1860
aagg	gg	cc	at	ac	cc	aa	gg	1920
ac	gat	cc	gg	gt	cc	aa	gt	1928
cac	atag	g	ca	at	cc	gg	cc	

<210> 44

<211> 591

<212> PRT

<213> Rhinocladiella atrovirens

<400> 44

Met Ala Leu Ala Pro Ser Tyr Ile Asn Pro Pro Asn Leu Ala Ser Pro	1	5	10	15
Ala Gly Tyr Ser Tyr Val Gly Val Gly Pro Asn Gly Gly Arg Tyr Val	20	25	30	
Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Ala Val Thr Asp Pro	35	40	45	
Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys	50	55	60	
Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Ile Thr Lys Leu Asn Tyr	65	70	75	80
Tyr Ile Val Asp Tyr Asn Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly	85	90	95	
Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val	100	105	110	
Pro Val Pro Ala Leu Ala Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala	115	120	125	
Thr Ala Leu Val Pro Gly His Ser Thr Pro Asp Asn Val Ala Asp Val	130	135	140	
Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val	145	150	155	160
Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val	165	170	175	
Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Gly Arg Thr Thr Ile Asn	180	185	190	
Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Lys	195	200	205	
Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr Thr Gly	210	215	220	
Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro Tyr	225	230	235	240
Gly Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu Leu Leu	245	250	255	
Pro Ala Ser Gln Leu Ile Glu Glu His Ser Leu Glu Asp Pro Lys Ala	260	265	270	
Ser Pro Gln Ala Lys Gln Leu Asp Ser Val Ser Phe Ala His Tyr Cys	275	280	285	
Glu Lys Leu Asn Leu Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg	290	295	300	
Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser Met Phe Phe Leu Thr				

305	310	315	320
Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn Ile Val Ser Asp Lys			
325	330	335	
Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Leu			
340	345	350	
Cys His Ala Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn			
355	360	365	
Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg			
370	375	380	
Ser Ala Ser Gly Gly Val Phe Arg Ser Lys Lys Val Val Leu Pro Thr			
385	390	395	400
Leu Tyr Pro Thr Leu Ile Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln			
405	410	415	
Ala Leu Ala Glu Lys Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe			
420	425	430	
Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu			
435	440	445	
Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Glu			
450	455	460	
Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly			
465	470	475	480
Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp			
485	490	495	
Asn Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu			
500	505	510	
Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln			
515	520	525	
Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Cys Leu Asn Thr Leu Gly			
530	535	540	
Ser Ala Leu Arg Thr Pro Phe Lys Gly Val His Phe Val Gly Thr Glu			
545	550	555	560
Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly			
565	570	575	
Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala			
580	585	590	

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<210> 45
<211> 1928
<212> DNA
<213> Rhinocladiella atrovirens
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<221> intron  
<222> (1134) ... (1185)

<400> 45

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cacgtcgccg	taggccccaa	cgaggaggagg	tatgtgacaa	tagctggaca	gattggacaa	120
gacgcttcgg	ccgtgacaga	ccctgcctac	gagaaacagg	ttgccccaa	gcttgcacac	180
ctgcgagctt	gttttgcgtc	agttggagcc	acttcaaacg	acattaccaa	gctcaattac	240
tacatgtcg	actacaaccc	gagcaaactc	accgcaattt	gagatgggct	gaaggctacc	300
tttgccttgc	acaggctccc	tccttgcacg	ctgggtccag	tgccggccct	ggttcacct	360
gaataacctt	ttgaggttga	tgctacggcg	ctggttccag	gacactcaac	cccgagacaat	420
gttgcggacg	tggtcgttgt	gggctgttgc	ttgagcgggtt	tggagacggc	acgcaaagtcc	480
caggctgccc	ggctgtcctt	cctcgttctt	gaggcgatgg	atcggtgtgg	ggaaagact	540
ctgagcgtac	aatcggttcc	cggtcggacg	actatcaatg	acctcggcgc	tgcgtggatc	600
aatgacagca	acccaaagcga	agtattcaaa	ttatgtggaaa	gatttcaattt	ggagggcggag	660
ctccagagga	cgaccggaaa	ttcaatccat	caagcacaag	acggtacaac	cactacagct	720
ccttatggtg	attcccttgt	aggcacaatt	ccatcttgt	atgagacetc	tgtcggtgt	780

agaatacagt	cgctgactcc	acatcgtocca	gctgagcgag	gagggttgc当地	gtgcactc当地	840
ggaactccctt	cccgcatgg	ctcagctgat	cgaagagcat	agtcttgaaag	accccaaggc	900
gagccctcaa	gogaagcagc	tcgacagtt	gagcttc当地	cactactgt	agaaggatct	960
aaacttgctt	gctgttctcg	gctgtggcaa	ccagatcaca	cgcgctctgc	tcgtgtgga	1020
agcccacgag	atcagcatgc	tttttctcac	cgactacatc	aagagtgc当地	ccgtctc当地	1080
taatattgtc	tcggataaga	aagacggtgg	gcagtatatg	cgatgc当地	cagggtcg	1140
tggtgttctc	tcagtgggag	actcgtttct	tagtggat	tccaggtatg	cagtc当地	1200
gccatgccc	gtcaaaggaa	cttggccag	gctcagtgca	cctcaacacc	ccgtcgccg	1260
aaattgagca	gtcgccatcc	ggctgtacag	tacgatcggc	ctcggccggc	gtgttccgaa	1320
gtaaaaagg	gtgtgttcc	ttaccgacaa	ccttgatcc	cacccatgata	ttttcaccac	1380
ctcttccgc	cgagaagcaa	gcattggctg	aaaatccat	cctgggc当地	tatagcaaga	1440
tagtcttcgt	atgggacaag	ctgtgttggc	gcaacaagg	cttctcgccg	gtcctccat	1500
cgagctgtg	ccccatctca	tttgc当地	ataccagcat	cgaagtc当地	cggcaatgg	1560
ccattacctg	tttcatggc	ggagacc	gacggaa	gtcccaacag	tccaa	1620
tacgacagaa	gtctgtctgg	aaccaactcc	gogcagcc	cgagaacg	ggggcccaag	1680
tcccagagcc	ggccaacgtg	ctcgagatcg	agtgg	gcagc当地	ttccaaggag	1740
cgccgagcgc	cgtctatggg	ctgaactg	tcaacacact	gggttccg	ctcagaacgc	1800
cgttcaagg	tgttcattt	gttggaa	agacgtctt	gttttggaaa	gggttatatgg	1860
aaggggccat	acgatcg	cagcagg	ctgcaga	gtgtcctag	ctgg	1920
cagcatag						1928

<210> 46  
 <211> 591  
 <212> PRT  
 <213> Rhinocladiella atrovirens

<400> 46  
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 Gly Tyr Ser His Val Gly Val Gly Pro Asn Gly Gly Arg Tyr Val Thr  
 20 25 30  
 Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Ala Val Thr Asp Pro Ala  
 35 40 45  
 Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys Leu  
 50 55 60  
 Ala Ala Val Gly Ala Thr Ser Asn Asp Ile Thr Lys Leu Asn Tyr Tyr  
 65 70 75 80  
 Ile Val Asp Tyr Asn Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly Leu  
 85 90 95  
 Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val Pro  
 100 105 110  
 Val Pro Ala Leu Ala Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala Thr  
 115 120 125  
 Ala Leu Val Pro Gly His Ser Thr Pro Asp Asn Val Ala Asp Val Val  
 130 135 140  
 Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val Gln  
 145 150 155 160  
 Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val Gly  
 165 170 175  
 Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile Asn  
 180 185 190  
 Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val Phe  
 195 200 205  
 Lys Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr Thr  
 210 215 220  
 Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala Pro  
 225 230 235 240  
 Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala Glu  
 245 250 255  
 Leu Leu Pro Ala Trp Ser Gln Leu Ile Glu Glu His Ser Leu Glu Asp  
 260 265 270  
 Pro Lys Ala Ser Pro Gln Ala Lys Gln Leu Asp Ser Val Phe Ala

275	280	285
His Tyr Cys Glu Lys Asp Leu Asn Leu Pro Ala Val Leu Gly Val Ala		
290	295	300
Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile Ser		
305	310	315
Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser Asn		
325	330	335
Ile Val Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys Thr		
340	345	350
Gly Met Gln Ser Leu Cys His Ala Met Ser Lys Glu Leu Val Pro Gly		
355	360	365
Ser Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala Ser		
370	375	380
Gly Cys Thr Val Arg Ser Ala Ser Gly Gly Val Phe Arg Ser Lys Lys		
385	390	395
Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Ile Phe Ser Pro Leu		
405	410	415
Pro Ala Glu Lys Gln Ala Leu Ala Glu Lys Ser Ile Gly Tyr Tyr Ser		
420	425	430
Lys Ile Val Phe Val Asp Lys Leu Trp Trp Arg Glu Gln Gly Phe Ser		
435	440	445
Gly Val Leu Gln Ser Ser Cys Asp Pro Ile Ser Phe Ala Arg Asp Thr		
450	455	460
Ser Ile Glu Val Asp Arg Gln Ser Ile Thr Cys Phe Met Val Gly Asp		
465	470	475
Pro Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val		
485	490	495
Trp Asn Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro		
500	505	510
Glu Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe		
515	520	525
Gln Ala Pro Ser Ala Val Tyr Gly Leu Asn Cys Leu Asn Thr Leu Gly		
530	535	540
Ser Ala Leu Arg Thr Pro Phe Lys Gly Val His Phe Val Gly Thr Glu		
545	550	555
Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly		
565	570	575
Gln Arg Gly Ala Ala Glu Val Val Pro Ser Leu Val Pro Ala Ala		
580	585	590

<210> 47  
 <211> 600  
 <212> PRT  
 <213> Exophiala spinifera

<400> 47		
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Ala Gly Tyr Ser His Val Gly Val Gly Pro Asp Gly Gly Arg Tyr Val		
20	25	30
Thr Ile Ala Gly Gln Ile Gly Gln Asp Ala Ser Gly Val Thr Asp Pro		
35	40	45
Ala Tyr Glu Lys Gln Val Ala Gln Ala Phe Ala Asn Leu Arg Ala Cys		
50	55	60
Leu Ala Ala Val Gly Ala Thr Ser Asn Asp Val Thr Lys Leu Asn Tyr		
65	70	75
Tyr Ile Val Asp Tyr Ala Pro Ser Lys Leu Thr Ala Ile Gly Asp Gly		
85	90	95
Leu Lys Ala Thr Phe Ala Leu Asp Arg Leu Pro Pro Cys Thr Leu Val		
100	105	110
Pro Val Ser Ala Leu Ser Ser Pro Glu Tyr Leu Phe Glu Val Asp Ala		
115	120	125

Thr Ala Leu Val Pro Gly His Thr Thr Pro Asp Asn Val Ala Asp Val  
 130 135 140  
 Val Val Val Gly Ala Gly Leu Ser Gly Leu Glu Thr Ala Arg Lys Val  
 145 150 155 160  
 Gln Ala Ala Gly Leu Ser Cys Leu Val Leu Glu Ala Met Asp Arg Val  
 165 170 175  
 Gly Gly Lys Thr Leu Ser Val Gln Ser Gly Pro Gly Arg Thr Thr Ile  
 180 185 190  
 Asn Asp Leu Gly Ala Ala Trp Ile Asn Asp Ser Asn Gln Ser Glu Val  
 195 200 205  
 Ser Arg Leu Phe Glu Arg Phe His Leu Glu Gly Glu Leu Gln Arg Thr  
 210 215 220  
 Thr Gly Asn Ser Ile His Gln Ala Gln Asp Gly Thr Thr Thr Ala  
 225 230 235 240  
 Pro Tyr Gly Asp Ser Leu Leu Ser Glu Glu Val Ala Ser Ala Leu Ala  
 245 250 255  
 Glu Leu Leu Pro Val Trp Ser Gln Leu Ile Glu Glu His Ser Leu Gln  
 260 265 270  
 Asp Leu Lys Ala Ser Pro Gln Ala Lys Arg Leu Asp Ser Val Ser Phe  
 275 280 285  
 Ala His Tyr Cys Glu Lys Glu Leu Asn Leu Pro Ala Val Leu Gly Val  
 290 295 300  
 Ala Asn Gln Ile Thr Arg Ala Leu Leu Gly Val Glu Ala His Glu Ile  
 305 310 315 320  
 Ser Met Leu Phe Leu Thr Asp Tyr Ile Lys Ser Ala Thr Gly Leu Ser  
 325 330 335  
 Asn Ile Phe Ser Asp Lys Lys Asp Gly Gly Gln Tyr Met Arg Cys Lys  
 340 345 350  
 Thr Gly Met Gln Ser Ile Cys His Ala Met Ser Lys Glu Leu Val Pro  
 355 360 365  
 Gly Ser Val His Leu Asn Thr Pro Val Ala Glu Ile Glu Gln Ser Ala  
 370 375 380  
 Ser Gly Cys Thr Val Arg Ser Ala Ser Gly Ala Val Phe Arg Ser Lys  
 385 390 395 400  
 Lys Val Val Val Ser Leu Pro Thr Thr Leu Tyr Pro Thr Leu Thr Phe  
 405 410 415  
 Ser Pro Pro Leu Pro Ala Glu Lys Gln Ala Leu Ala Glu Asn Ser Ile  
 420 425 430  
 Leu Gly Tyr Tyr Ser Lys Ile Val Phe Val Trp Asp Lys Pro Trp Trp  
 435 440 445  
 Arg Glu Gln Gly Phe Ser Gly Val Leu Gln Ser Ser Cys Asp Pro Ile  
 450 455 460  
 Ser Phe Ala Arg Asp Thr Ser Ile Asp Val Asp Arg Gln Trp Ser Ile  
 465 470 475 480  
 Thr Cys Phe Met Val Gly Asp Pro Gly Arg Lys Trp Ser Gln Gln Ser  
 485 490 495  
 Lys Gln Val Arg Gln Lys Ser Val Trp Asp Gln Leu Arg Ala Ala Tyr  
 500 505 510  
 Glu Asn Ala Gly Ala Gln Val Pro Glu Pro Ala Asn Val Leu Glu Ile  
 515 520 525  
 Glu Trp Ser Lys Gln Gln Tyr Phe Gln Gly Ala Pro Ser Ala Val Tyr  
 530 535 540  
 Gly Leu Asn Asp Leu Ile Thr Leu Gly Ser Ala Leu Arg Thr Pro Phe  
 545 550 555 560  
 Lys Ser Val His Phe Val Gly Thr Glu Thr Ser Leu Val Trp Lys Gly  
 565 570 575  
 Tyr Met Glu Gly Ala Ile Arg Ser Gly Gln Arg Gly Ala Ala Glu Val  
 580 585 590  
 Val Ala Ser Leu Val Pro Ala Ala  
 595 600

<211> 1392  
 <212> DNA  
 <213> Unknown  
  
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 <221> CDS  
 <222> (1) ... (1392)  
  
 <223> Cys (-) APAO; removal of cysteine 461  
  
 <400> 48  
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 Lys Asp Asn Val Ala Asp Val Val Val Val Gly Ala Gly Leu Ser Gly  
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 ttg gag acg gca cgc aaa gtc cag gcc gcc ggt ctg tcc tgc ctc gtt 96  
 Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val  
 20 25 30  
  
 ctt gag gcg atg gat cgt gta ggg gga aag act ctg agc gta caa tcg 144  
 Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser  
 35 40 45  
  
 ggt ccc ggc agg acg act atc aac gac ctc ggc gct gcg tgg atc aat 192  
 Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn  
 50 55 60  
  
 gac agc aac caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg 240  
 Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu  
 65 70 75 80  
  
 gag ggc gag ctc cag agg acg act gga aat tca atc cat caa gca caa 288  
 Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln  
 85 90 95  
  
 gac ggt aca acc act aca gct cct tat ggt gac tcc ttg ctg agc gag 336  
 Asp Gly Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu  
 100 105 110  
  
 gag gtt gca agt gca ctt gcg gaa ctc ctc ccc gta tgg tct cag ctg 384  
 Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu  
 115 120 125  
  
 atc gaa gag cat agc ctt caa gac ctc aag gcg agc cct cag gcg aag 432  
 Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys  
 130 135 140  
  
 cgg ctc gac agt gtg agc ttc gcg cac tac tgt gag aag gaa cta aac 480  
 Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn  
 145 150 155 160  
  
 ttg cct gct gtt ctc ggc gta gca aac cag atc aca cgc gct ctg ctc 528  
 Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu  
 165 170 175  
  
 ggt gtg gaa gcc cac gag atc agc atg ctt ttt ctc acc gac tac atc 576  
 Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile  
 180 185 190  
  
 aag agt gcc acc ggt ctc agt aat att ttc tcg gac aag aaa gac ggc 624  
 Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly  
 195 200 205

ggg cag tat atg cga tgc aaa aca ggt atg cag tcg att tgc cat gcc Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala 210 215 220	672
atg tca aag gaa ctt gtt cca ggc tca gtg cac ctc aac acc ccc gtc Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val 225 230 235 240	720
gct gaa att gag cag tcg gca tcc ggc tgt aca gta cga tcg gcc tcg Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser 245 250 255	768
ggc gcc gtg ttc cga agc aaa aag gtg gtg gtt tcg tta ccg aca acc Gly Ala Val Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr 260 265 270	816
ttg tat ccc acc ttg aca ttt tca cca cct ctt ccc gcc gag aag caa Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln 275 280 285	864
gca ttg gcg gaa aat tct atc ctg ggc tac tat agc aag ata gtc ttc Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe 290 295 300	912
gta tgg gac aag ccg tgg tgg cgc gaa caa ggc ttc tcg ggc gtc ctc Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu 305 310 315 320	960
caa tcg agc tcc gac ccc atc tca ttt gcc aga gat acc agc atc gac Gln Ser Ser Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp 325 330 335	1008
gtc gat cga caa tgg tcc att acc tgt ttc atg gtc gga gac ccg gga Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly 340 345 350	1056
cgg aag tgg tcc caa cag tcc aag cag gta cga caa aag tct gtc tgg Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp 355 360 365	1104
gac caa ctc cgc gca gcc tac gag aac gcc ggg gcc caa gtc cca gag Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu 370 375 380	1152
ccg gcc aac gtg ctc gaa atc gag tgg tcg aag cag cag tat ttc caa Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln 385 390 395 400	1200
gga gct ccg agc gcc gtc tat ggg ctg aac gat ctc atc aca ctg ggt Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly 405 410 415	1248
tcg gcg ctc aga acg ccg ttc aag agt gtt cat ttc gtt gga acg gag Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu 420 425 430	1296
acg tct tta gtt tgg aaa ggg tat atg gaa ggg gcc ata cga tcg ggt Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly 435 440 445	1344
caa cga ggt gct gca gaa gtt gtg gct agc ctg gtg cca gca gca tag	1392

Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala \*  
450 455 460

<210> 49  
<211> 463  
<212> PRT  
<213> Unknown

<220>  
<223> Cys (-) APAO; removal of cysteine 461

<400> 49  
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20 25 30  
Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser  
35 40 45  
Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn  
50 55 60  
Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu  
65 70 75 80  
Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln  
85 90 95  
Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu  
100 105 110  
Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu  
115 120 125  
Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys  
130 135 140  
Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn  
145 150 155 160  
Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu  
165 170 175  
Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile  
180 185 190  
Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly  
195 200 205  
Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Ile Cys His Ala  
210 215 220  
Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val  
225 230 235 240  
Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser  
245 250 255  
Gly Ala Val Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr  
260 265 270  
Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln  
275 280 285  
Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe  
290 295 300  
Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu  
305 310 315 320  
Gln Ser Ser Ser Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp  
325 330 335  
Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly  
340 345 350  
Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp  
355 360 365  
Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu  
370 375 380  
Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln

385	390	395	400
Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly			
405	410		415
Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu			
420	425		430
Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly			
435	440		445
Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala			
450	455		460
<210> 50			
<211> 1392			
<212> DNA			
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<221> CDS			
<222> (1)...(1392)			
<223> Cys (-) APAO; removal of cysteines 359 and 461			
<400> 50			
aaa gac aac gtt gcg gac gtc gta gtg gtg ggc gct ggc ttg agc ggt			
Lys Asp Asn Val Ala Asp Val Val Val Val Gly Ala Gly Leu Ser Gly			
1	5	10	15
ttg gag acg gca cgc aaa gtc cag gcc gcc ggt ctg tcc tgc ctc gtt			
Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val			
20	25	30	
ctt gag gcg atg gat cgt gta ggg gga aag act ctg agc gta caa tcg			
Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser			
35	40	45	
ggt ccc ggc agg acg act atc aac gac ctc ggc gct gcg tgg atc aat			
Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn			
50	55	60	
gac agc aac caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg			
Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu			
65	70	75	80
gag ggc gag ctc cag agg acg act gga aat tca atc cat caa gca caa			
Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln			
85	90	95	
gac ggt aca acc act aca gct cct tat ggt gac tcc ttg ctg agc gag			
Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu			
100	105	110	
gag gtt gca agt gca ctt gcg gaa ctc ctc ccc gta tgg tct cag ctg			
Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu			
115	120	125	
atc gaa gag cat agc ctt caa gac ctc aag gcg agc cct cag gcg aag			
Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys			
130	135	140	
cgg ctc gac agt gtg agc ttc gcg cac tac tgt gag aag gaa cta aac			
Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn			
145	150	155	160

ttg cct gct gtt ctc ggc gta gca aac cag atc aca cgc gct ctg ctc Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu 165 170 175	528
ggt gtg gaa gcc cac gag atc agc atg ctt ttt ctc acc gac tac atc Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile 180 185 190	576
aag agt gcc acc ggt ctc agt aat att ttc tcg gac aag aaa gac ggc Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly 195 200 205	624
ggg cag tat atg cga tgc aaa aca ggt atg cag tcg att tcg cat gcc Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Ile Ser His Ala 210 215 220	672
atg tca aag gaa ctt gtt cca ggc tca gtg cac ctc aac acc ccc gtc Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val 225 230 235 240	720
gct gaa att gag cag tcg gca tcc ggc tgt aca gta cga tcg gcc tcg Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser 245 250 255	768
ggc gcc gtg ttc cga agc aaa aag gtg gtg gtt tcg tta ccg aca acc Gly Ala Val Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr 260 265 270	816
ttg tat ccc acc ttg aca ttt tca cca cct ctt ccc gcc gag aag caa Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln 275 280 285	864
gca ttg gcg gaa aat tct atc ctg ggc tac tat agc aag ata gtc ttc Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe 290 295 300	912
gta tgg gac aag ccg tgg tgg cgc gaa caa ggc ttc tcg ggc gtc ctc Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu 305 310 315 320	960
caa tcg agc tcc gac ccc atc tca ttt gcc aga gat acc agc atc gac Gln Ser Ser Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp 325 330 335	1008
gtc gat cga caa tgg tcc att acc tgt ttc atg gtc gga gac ccg gga Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly 340 345 350	1056
ccg aag tgg tcc caa cag tcc aag cag gta cga caa aag tct gtc tgg Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp 355 360 365	1104
gac caa ctc cgc gca gcc tac gag aac gcc ggg gcc caa gtc cca gag Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu 370 375 380	1152
ccg gcc aac gtg ctc gaa atc gag tgg tcg aag cag cag tat ttc caa Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln 385 390 395 400	1200
gga gct ccg agc gcc gtc tat ggg ctg aac gat ctc atc aca ctg ggt Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly	1248

405	410	415	
tcg gcg ctc aga acg ccg ttc aag agt gtt cat ttc gtt gga acg gag Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu 420	425	430	1296
acg tct tta gtt tgg aaa ggg tat atg gaa ggg gcc ata cga tcg ggt Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly 435	440	445	1344
caa cga ggt gct gca gaa gtt gtg gct agc ctg gtg cca gca gca tag Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala * 450	455	460	1392
<p>&lt;210&gt; 51  &lt;211&gt; 463  &lt;212&gt; PRT  &lt;213&gt; Unknown</p> <p>&lt;220&gt;  &lt;223&gt; Cys (-) APAO; removal of cysteines 359 and 461</p> <p>&lt;400&gt; 51  Lys Asp Asn Val Ala Asp Val Val Val Gly Ala Gly Leu Ser Gly  1 5 10 15  Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Cys Leu Val  20 25 30  Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser  35 40 45  Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn  50 55 60  Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu  65 70 75 80  Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln  85 90 95  Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu  100 105 110  Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu  115 120 125  Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys  130 135 140  Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn  145 150 155 160  Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu  165 170 175  Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile  180 185 190  Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly  195 200 205  Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Ile Ser His Ala  210 215 220  Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val  225 230 235 240  Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser  245 250 255  Gly Ala Val Phe Arg Ser Lys Lys Val Val Ser Leu Pro Thr Thr  260 265 270  Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln  275 280 285  Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe  290 295 300</p>			

Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu  
 305 310 315 320  
 Gln Ser Ser Ser Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp  
 325 330 335  
 Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly  
 340 345 350  
 Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp  
 355 360 365  
 Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu  
 370 375 380  
 Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln  
 385 390 395 400  
 Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly  
 405 410 415  
 Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu  
 420 425 430  
 Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly  
 435 440 445  
 Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala  
 450 455 460

<210> 52

<211> 1392

<212> DNA

<213> Unknown

<220>

<221> CDS

<222> (1)...(1392)

<223> Cys (-) APAO; removal of cysteines 169, 359, and  
461

<400> 52

aaa gac aac gtt gct gac gtg gta gtg gtg ggc gct ggc ttg agc ggt	48
Lys Asp Asn Val Ala Asp Val Val Val Val Gly Ala Gly Leu Ser Gly	
1 5 10 15	

ttg gag acg gca cgc aaa gtc cag gcc gcc ggt ctg agc tcc ctc gtt	96
Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Ser Leu Val	
20 25 30	

ctt gag gcg atg gat cgt gta ggg gga aag act ctg agc gta caa tcg	144
Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser	
35 40 45	

ggt ccc ggc agg acg act atc aac gac ctc ggc gct gcg tgg atc aat	192
Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn	
50 55 60	

gac agc aac caa agc gaa gta tcc aga ttg ttt gaa aga ttt cat ttg	240
Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu	
65 70 75 80	

gag ggc gag ctc cag agg acg act gga aat tca atc cat caa gca caa	288
Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln	
85 90 95	

gac ggt aca acc act aca gct cct tat ggt gac tcc ttg ctg agc gag	336
Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu	
100 105 110	

gag gtt gca agt gca ctt gcg gaa ctc ctc ccc gta tgg tct cag ctg Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu 115 120 125	384
atc gaa gag cat agc ctt caa gac ctc aag gcg agc cct cag gcg aag Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys 130 135 140	432
cgg ctc gac agt gtg agc ttc gcg cac tac tgt gag aag gaa cta aac Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn 145 150 155 160	480
ttg cct gct gtt ctc ggc gta gca aac cag atc aca cgc gct ctg ctc Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu 165 170 175	528
ggt gtg gaa gcc cac gag atc agc atg ctt ttt ctc acc gac tac atc Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile 180 185 190	576
aag agt gcc acc ggt ctc agt aat att ttc tcg gac aag aaa gac ggc Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly 195 200 205	624
ggg cag tat atg cga tgc aaa aca ggt atg cag tcg att tcg cat gcc Gly Gln Tyr Met Arg Cys Lys Thr Gly Met Gln Ser Ile Ser His Ala 210 215 220	672
atg tca aag gaa ctt gtt cca ggc tca gtg cac ctc aac acc ccc gtc Met Ser Lys Glu Leu Val Pro Gly Ser Val His Leu Asn Thr Pro Val 225 230 235 240	720
gct gaa att gag cag tcg gca tcc ggc tgt aca gta cga tcg gcc tcg Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys Thr Val Arg Ser Ala Ser 245 250 255	768
ggc gcc gtg ttc cga agc aaa aag gtg gtg gtt tcg tta ccg aca acc Gly Ala Val Phe Arg Ser Lys Lys Val Val Ser Leu Pro Thr Thr 260 265 270	816
ttg tat ccc acc ttg aca ttt tca cca cct ctt ccc gcc gag aag caa Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln 275 280 285	864
gca ttg gcg gaa aat tct atc ctg ggc tac tat agc aag ata gtc ttc Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe 290 295 300	912
gta tgg gac aag ccg tgg tgg cgc gaa caa ggc ttc tcg ggc gtc ctc Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu 305 310 315 320	960
caa tcg agc tcc gac ccc atc tca ttt gcc aga gat acc agc atc gac Gln Ser Ser Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp 325 330 335	1008
gtc gat cga caa tgg tcc att acc tgt ttc atg gtc gga gac ccg gga Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly 340 345 350	1056
cgg aag tgg tcc caa cag tcc aag cag gta cga caa aag tct gtc tgg Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp	1104

355	360	365	
gac caa ctc cgc gca gcc tac gag aac gcc ggg gcc caa gtc cca gag Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu 370	375	380	1152
ccg gcc aac gtg ctc gaa atc gag tgg tcg aag cag cag tat ttc caa Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln 385	390	395	1200
gga gct ccg agc gcc gtc tat ggg ctg aac gat ctc atc aca ctg ggt Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly 405	410	415	1248
tcg gcg ctc aga acg ccg ttc aag agt gtt cat ttc gtt gga acg gag Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu 420	425	430	1296
acg tct tta gtt tgg aaa ggg tat atg gaa ggg gcc ata cga tcg ggt Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly 435	440	445	1344
caa cga ggt gct gca gaa gtt gtg gct agc ctg gtg cca gca gca tag Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala * 450	455	460	1392
<p>&lt;210&gt; 53  &lt;211&gt; 463  &lt;212&gt; PRT  &lt;213&gt; Unknown</p> <p>&lt;220&gt;  &lt;223&gt; Cys (-) APAO; removal of cysteines 169, 359, and  461</p> <p>&lt;400&gt; 53  Lys Asp Asn Val Ala Asp Val Val Val Gly Ala Gly Leu Ser Gly  1 5 10 15  Leu Glu Thr Ala Arg Lys Val Gln Ala Ala Gly Leu Ser Ser Leu Val  20 25 30  Leu Glu Ala Met Asp Arg Val Gly Gly Lys Thr Leu Ser Val Gln Ser  35 40 45  Gly Pro Gly Arg Thr Thr Ile Asn Asp Leu Gly Ala Ala Trp Ile Asn  50 55 60  Asp Ser Asn Gln Ser Glu Val Ser Arg Leu Phe Glu Arg Phe His Leu  65 70 75 80  Glu Gly Glu Leu Gln Arg Thr Thr Gly Asn Ser Ile His Gln Ala Gln  85 90 95  Asp Gly Thr Thr Thr Ala Pro Tyr Gly Asp Ser Leu Leu Ser Glu  100 105 110  Glu Val Ala Ser Ala Leu Ala Glu Leu Leu Pro Val Trp Ser Gln Leu  115 120 125  Ile Glu Glu His Ser Leu Gln Asp Leu Lys Ala Ser Pro Gln Ala Lys  130 135 140  Arg Leu Asp Ser Val Ser Phe Ala His Tyr Cys Glu Lys Glu Leu Asn  145 150 155 160  Leu Pro Ala Val Leu Gly Val Ala Asn Gln Ile Thr Arg Ala Leu Leu  165 170 175  Gly Val Glu Ala His Glu Ile Ser Met Leu Phe Leu Thr Asp Tyr Ile  180 185 190  Lys Ser Ala Thr Gly Leu Ser Asn Ile Phe Ser Asp Lys Lys Asp Gly</p>			

195	200	205
Gly Gln Tyr Met Arg Cys Lys Thr	Gly Met Gln Ser Ile Ser His Ala	
210	215	220
Met Ser Lys Glu Leu Val Pro Gly Ser Val His	Leu Asn Thr Pro Val	
225	230	235
Ala Glu Ile Glu Gln Ser Ala Ser Gly Cys	Thr Val Arg Ser Ala Ser	
245	250	255
Gly Ala Val Phe Arg Ser Lys Lys Val Val Val Ser Leu Pro Thr Thr		
260	265	270
Leu Tyr Pro Thr Leu Thr Phe Ser Pro Pro Leu Pro Ala Glu Lys Gln		
275	280	285
Ala Leu Ala Glu Asn Ser Ile Leu Gly Tyr Tyr Ser Lys Ile Val Phe		
290	295	300
Val Trp Asp Lys Pro Trp Trp Arg Glu Gln Gly Phe Ser Gly Val Leu		
305	310	315
Gln Ser Ser Ser Asp Pro Ile Ser Phe Ala Arg Asp Thr Ser Ile Asp		
325	330	335
Val Asp Arg Gln Trp Ser Ile Thr Cys Phe Met Val Gly Asp Pro Gly		
340	345	350
Arg Lys Trp Ser Gln Gln Ser Lys Gln Val Arg Gln Lys Ser Val Trp		
355	360	365
Asp Gln Leu Arg Ala Ala Tyr Glu Asn Ala Gly Ala Gln Val Pro Glu		
370	375	380
Pro Ala Asn Val Leu Glu Ile Glu Trp Ser Lys Gln Gln Tyr Phe Gln		
385	390	395
Gly Ala Pro Ser Ala Val Tyr Gly Leu Asn Asp Leu Ile Thr Leu Gly		
405	410	415
Ser Ala Leu Arg Thr Pro Phe Lys Ser Val His Phe Val Gly Thr Glu		
420	425	430
Thr Ser Leu Val Trp Lys Gly Tyr Met Glu Gly Ala Ile Arg Ser Gly		
435	440	445
Gln Arg Gly Ala Ala Glu Val Val Ala Ser Leu Val Pro Ala Ala		
450	455	460